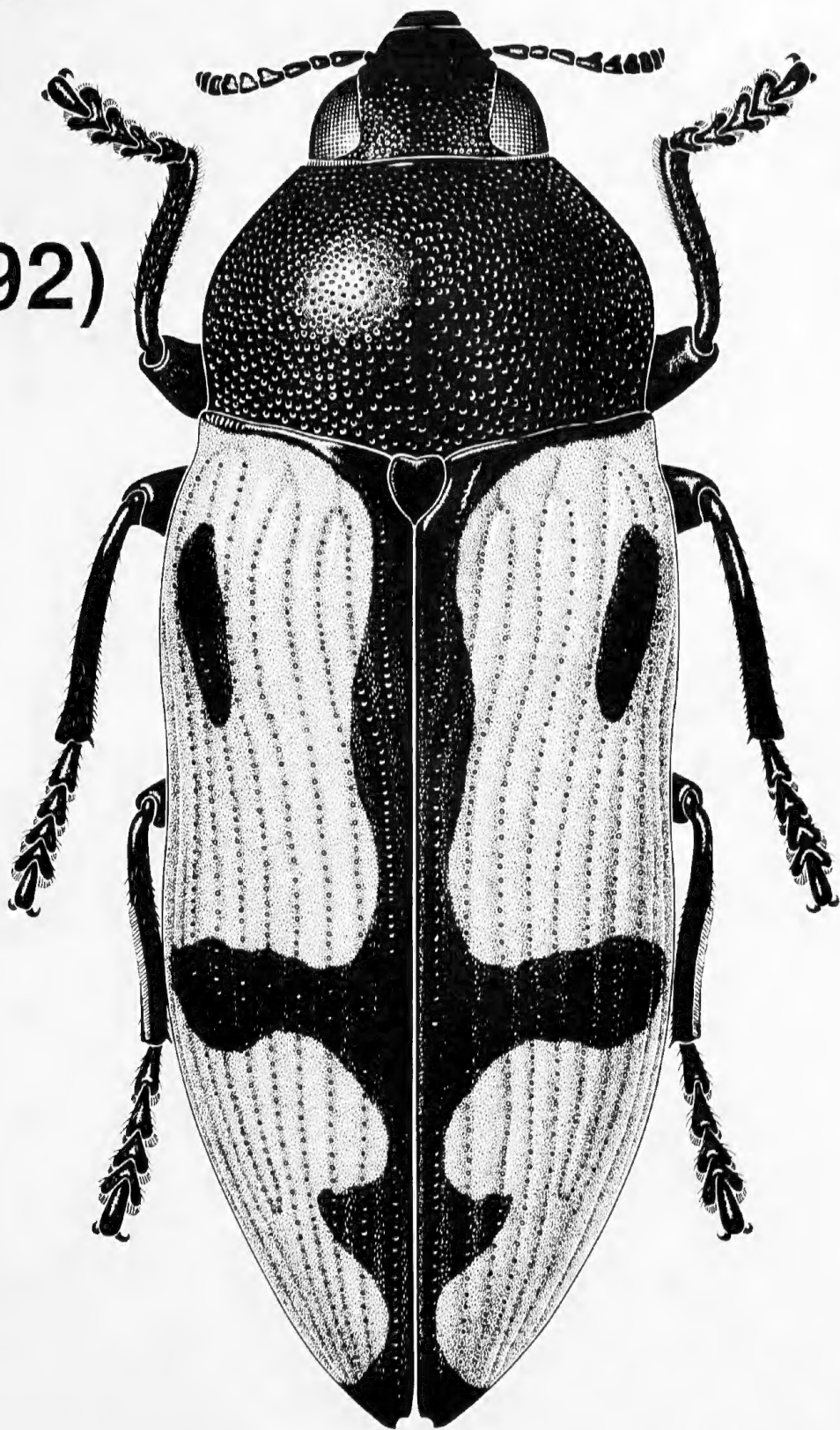


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Front cover : *Castiarina burnsi* (Barker) was named after Mr Gordon Burns in recognition of his enormous contribution to the study of burprestid beetles. The species occurs in the Big and Little Deserts in Victoria and Gawler in South Australia. Illustration by Graham Milledge.

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Manuscripts must be typed on A4 paper, double-spaced, on one side of the paper and with ample margins. Final manuscripts on floppy discs are encouraged. Tables, captions to text figures and plates must be attached to the manuscript as final pages. Underlining in the text should be restricted to generic and specific names. Measurements must be in the metric system (SI units).

References should be listed alphabetically at the end of the manuscript. Journal citations must be in full. References to books must give the year of publication, edition, name of publisher and city of publication. Titles of books and names of journals should be underlined.

Photographs must have clear definition and may be submitted as either glossy or flat prints at the actual size for reproduction. Line drawings for text figures should be in black ink on white card or drawing film. Maximum full page size is 177 mm wide by 220 mm, single column width is 86 mm. Clear lettering must be inserted. Original drawings up to twice final size are acceptable.

The distribution of Victorian jewel beetles (Coleoptera: Buprestidae) – an ENTRECS project

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Abstract. Burns, G.G. and Burns, A.J., 1992. The distribution of Victorian jewel beetles (Coleoptera: Buprestidae) – an ENTRECS project. *Occasional Papers from the Museum of Victoria* 5: 1–53.

A checklist of the 300 species of Victorian Buprestidae is presented with maps showing the distribution of each species.

Introduction

The ENTRECS PROJECT of the Entomological Society of Victoria (Inc.) aims to record data on the distribution of insects in Victoria in a form which could be readily expanded to cover the whole of Australia. An explanatory handbook has been produced and data solicited from collectors. At its inception donations were received from two companies to assist in furthering the project. A first book with distribution maps of butterflies of Victoria was produced (Entomological Society of Victoria, 1986).

This, the second contribution to the project, is a checklist of the Victorian Buprestidae with maps showing the distribution of each species within Victoria. The text and maps have been compiled for the Entomological Society of Victoria (Inc.) on a Macintosh computer and are presented as a starting point for future workers. The compilation relies on the identifications of many collectors.

Systematics

Buprestidae, or jewel beetles, are, as their common name suggests, probably the most colourful of all beetles. Although they show great variation of size and colour they are all of much the same shape. The prothorax is closely joined to the rest of the body and the head fits neatly so that the beetle is compact, narrow and oval. They can be recognized by a transverse metasternal suture and the partly fused first two abdominal sternites (Matthews, 1985). The adults are very active in hot weather and fly readily in sunlight. Many can be found on nectar bearing flowers such as, *Eucalyptus*, *Leptospermum*, *Bursaria* and *Baecchia*.

The family name Buprestidae was introduced by Leach (1815) and used later by Eschscholtz (1829). The name is from Greek: “*bous*” (cattle) and “*prethein*” (to blow up). The beetles were evidently thought to be poisonous to animals and were probably confused with meloids (Bright, 1987). There are about 12 000 species worldwide with approximately 850 species known from Australia. Bellamy (1986) recognised seven subfamilies and 22 tribes for Australian Buprestidae. All

subfamilies and 17 of the tribes are represented from Victoria by 300 species.

Bellamy's (1986) classification of the higher taxa and his synonymies are followed in this contribution. More recent changes to his systematic arrangement have not been followed. Cobos (1986) included *Merimna* Thomson in Melanophilini rather than in Actenodini as is usual. Toyama (1987) synonymised Chalcophorinae with Buprestinae but did not elaborate on how the tribes of Chalcophorinae should be considered. For some species dealt with in this paper more complete synonymies than are given here can be found in Carter and Théry (1929).

The only taxonomic changes made here involve *Pachycisseis* and *Neospades* (Bellamy, 1988: 417) and *Themognatha* and *Castiarina* (Gardner, 1990: 338). All four names have been elevated from subgeneric to generic rank.

Distribution maps

The ENTRECS scheme divides the State into major blocks, each corresponding to a map in the Commonwealth of Australia 1:250,000 series R502. Each block is given an alphabetical code. These major blocks are further subdivided into 54 areas each 10 minutes of latitude (18 km) by 10 minutes of longitude (15 km).

Map A shows the overall distribution of all buprestids recorded to 31 March 1990. The remaining maps are distributional maps for individual species. They represent data from 5735 records obtained from specimens in museums, private collections, literature, from private communication, or visually identified in the field. Each “record” represents one or more beetles in a single collection made at one time and place, or a unique field observation. Specimens come from 27 collections made by over 210 collectors. They date from the earliest specimens that we have been able to locate, 6 specimens of *Melobasis nervosa* Boisdual collected at Prahran in October 1865, until 31 March 1990.

The material for this report is based on collections in: Australian Museum, Sydney; Australian National Insect Collection, CSIRO, Canberra; Museum of Victoria, Melbourne; Queensland Museum, Brisbane; Entomology Department, University of Queensland, Brisbane; South Australian Museum,

¹ Gordon G. Burns died on 12 December 1990 shortly after completing this manuscript.

Adelaide; Tasmanian Museum, Hobart; Victorian Agricultural Insect Collection, Burnley; and the following private collections: E.E. Adams, G.W. Anderson, I. Faithfull, F. Hallgarten, J. Harslett, K. Hateley, D.R. Holmes, Y. Komiya, J.C. LeSouef, M.W. Mules, S.K. Smith, A. Sundholme, R.G. Thompson, G. and T. Williams and the authors' collection.

Maps are not provided for species recorded from Victoria only in the literature, or for species represented by specimens labelled simply as Victoria, Mallee dist., Western dist. or Gippsland. These are: *Neobubastes aureocincta* Blackburn, (Mallee dist.); *Neobubastes orientalis* Carter, (Vic.); *Balthasarella melandryoides* Obenberger, (literature); *Melobasis nais* Obenberger, (literature); *Melobasis occidentalis* Carter, (Western dist.); *Melobasis prominens* Obenberger, (literature); *Melobasis semimarginata* Obenberger, (literature); *Melobasis victoriae* Obenberger, (literature); *Themognatha aestimata* (Kerremans), (Vic.); *Themognatha conspicillata* (White), (Mallee dist.); *Themognatha pascoei* (Saunders), (Mallee dist.); *Themognatha sanguinea* (Saunders), (Mallee); *Castiarina arida* (Barker), (Vic.); *Castiarina commixta* (Carter), (Gippsland); *Neospades simplex* Blackburn, (Mallee dist.) and *Agrilus frenchi* Blackburn, (Vic.).

Some taxa described as "varieties" have not been investigated critically; *Torresita cuprifera* var. *limbata* Carter, *Melobasis gratiosissima* var. *aurora* Obenberger, *Melobasis gratiosissima* var. *amara* Obenberger, *Melobasis gratiosissima* var. *amabilior* Obenberger, *Melobasis obscurella* var. *chalcosoma* Obenberger, *Melobasis picticollis* var. *signatipennis* Obenberger, *Melobasis purpurascens* var. *anchoralis* Obenberger, *Melobasis vittata* var. *incipiens* Obenberger and *Stigmodera yarrelli* var. *coerulescens* Carter. As their precise status is unknown they are listed under the name of the species in which they were first described.

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CHECKLIST OF THE VICTORIAN BUPRESTIDAE

Family BUPRESTIDAE Eschscholtz

Buprestides Eschscholtz, 1829: 239.

Subfamily POLYCESTINAE Lacordaire

Polycestides Lacordaire, 1857: 61.

Tribe ASTRAEUSINI Cobos

Astraeusini Cobos, 1980: 28.

Buprestes (part) Carter and Théry, 1929: 270.

ASTRAEUS Laporte and Gory

Astraeus Laporte and Gory, 1837: 1.

Asthraeus Imhoff, 1856: 46 (misspelling).

Conognatha auctorum.

Subgenus ASTRAEUS Laporte and Gory

- Astraeus* (*Astraeus*) *badeni* van de Poll (Map 1)
Astraeus badeni van de Poll, 1889: 93.
Astraeus badeni var. *disjunctus* Obenberger, 1928: 204 (Barker, 1989: 188).
- Astraeus* (*Astraeus*) *dilutipes* van de Poll (Map 2)
Astraeus samouelli var. *dilutipes* van de Poll, 1886b: 180.
Astraeus strandi Obenberger, 1928: 205 (Barker, 1975: 123).
- Astraeus* (*Astraeus*) *jansonii* van de Poll (Map 3)
Astraeus jansonii van de Poll, 1889: 94.
Astraeus tepperi Blackburn, 1890a: 1258 (Barker, 1975: 133).
- Astraeus* (*Astraeus*) *major* Blackburn (Map 4)
Astraeus major Blackburn, 1890a: 1257.
Astraeus navarchis var. *major*.—Carter, 1929: 282 (Barker, 1975: 127).
- Astraeus* (*Astraeus*) *mastersii* Macleay (Map 5)
Astraeus mastersii Macleay, 1872: 239.
Astraeus samouelli van de Poll, 1886b: 176 (Barker, 1975: 122).
- Astraeus splendens* van de Poll, 1889: 108 (Barker, 1975: 122).
- Astraeus simplex* Blackburn, 1892d: 211 (Barker, 1975: 122).
- Astraeus mastersi* auctorum.
- Astraeus* (*Astraeus*) *navarchis* (Thomson) (Map 6)
Conognatha navarchis Thomson, 1856: 115, pl. 6, fig. 2.
Astraeus (*Astraeus*) *navarchis* (Thomson) (Barker, 1975: 114, 127).

- Astraeus (Astraeus) pygmaeus** van de Poll (Map 7)
Astraeus pygmaeus van de Poll, 1886b: 178.
Astraeus pygmaeus var. *subfasciatus* van de Poll, 1886b: 178 (Barker, 1975: 118).
Astraeus samouellei var. *pygmaeus*. — Blackburn, 1890a: 1256.
Astraeus samouelli var. *pygmaeus*. — Kerremans, 1892: 102 (misspelling).
Astraeus pygmaeus. — Kerremans, 1903a: 148 (misspelling).
Astraeus pygmaeus var. *subfasciatus*. — Kerremans, 1903a: 148 (misspelling).
Astraeus pygmaeus. — Carter, 1929: 282 (misspelling).
Astraeus pygmaeus var. *subfasciatus*. — Carter, 1929: 282 (misspelling).

Subgenus **DEPOLLUS** Barker
Depollus Barker, 1975: 105.

- Astraeus (Depollus) irregularis** van de Poll (Map 8)
Astraeus irregularis van de Poll, 1889: 86.

Tribe **PROSPHERINI** Cobos
 Prospherini Cobos, 1980: 84.
 Polycetae (part) Carter and Théry, 1929: 269.

PROSPHERES Saunders
Prospheres Saunders, 1868b: 11.
Phospheres Thomson, 1878: 61 (misspelling).

- Prospheres aurantiopicta** (Gory and Laporte) (Map 9)
Buprestis aurantiopicta Gory and Laporte, 1837: 132.
Prospheres aurantiopicta (Gory and Laporte) (Saunders, 1868b: 7).
Stigmodera gulielmi White, 1859b: 120 (Levey, 1978a: 713, 721).
Buprestis moesta Carter, 1915: 77 (Levey, 1978a: 713, 721).
Prospheres aurantiopictus var. *caledonicus* Obenberger, 1924a: 32 (Levey, 1978a: 713, 721).

Subfamily **MASTOGENINAE** Leconte and Horn
 Mastogeninae Leconte and Horn, 1883: 199 (as tribe).

Tribe **MASTOGENINI**

HELPERELLA Cobos
Helperella Cobos 1957: 91.

- Helperella frenchi** (Théry) (Map 10)
Mastogenius frenchi Théry, 1928: 456.
Helperella frenchi (Théry) (Williams and Weir, 1987: 159).

Subfamily **CHALCOPHORINAE** Lacordaire
 Chalcophorides Lacordaire, 1857: 14.

Tribe **EPISTOMENTINI** Levey
 Epistomentini Levey, 1978b: 155.

CYRIA Solier
Cyria Solier, 1833: 269.
Euchloris Billberg teste Mannerheim, 1837: 30.
Cyrioides Carter, 1920: 222.
Xenocyria Obenberger, 1947: 129.

- Cyria imperialis** (Fabricius) (Map 11)
Buprestis imperialis Fabricius, 1801: 204.
Cyria imperialis (Fabricius) (Saunders, 1868b: 3).

DIADOXUS Thomson
Diadoxus Thomson, 1878: 15.
Stigmodera auctorum.

Anthaxia auctorum.

- Diadoxus erythrurus** (White) (Map 12)
Stigmodera erythrura White, 1843b: 507, pl.1, fig. 7.
Diadoxus erythrurus (White) (Saunders, 1868b: 4).

- Diadoxus jungi** Blackburn (Map 13)
Diadoxus jungi Blackburn, 1899: 28.

- Diadoxus scalaris** (Gory and Laporte) (Map 14)
Buprestis scalaris Gory and Laporte, 1837: 141.
Diadoxus scalaris (Gory and Laporte) (Saunders, 1868b: 4).
Diadoxus scalaris var. *blackburni* Obenberger, 1923b: 72.

Tribe **CHALCOPHORINI**

IRIDOTAENIA Deyrolle
Iridotaenia Deyrolle, 1864: 25.
Paracupta auctorum.

- Iridotaenia albivittis** (Hope) (Map 15)
Buprestis albivittis Hope, 1846: 214.
Iridotaenia albivittis (Hope) (Carter, 1929: 301).
Chrysodema pyritosa Boheman, 1858: 58.

CHALCOTAENIA Deyrolle
Chalcotaenia Deyrolle, 1864: 12.
Chalcophora auctorum.

- Chalcotaenia exilis** (Blackburn) (Map 16)
Chalcophora exilis Blackburn, 1894: 98.

Tribe **JULODIMORPHINI** Kerremans
 Julodimorphites Kerremans, 1902: 16.
 Julodimorphae Carter and Théry, 1929: 271.

JULODIMORPHA Thomson

Julodimorpha Thomson, 1878: 51.

- Julodimorpha bakewellii** (White) (Map 17)
Stigmodera bakewellii White, 1859a: 290.
Julodimorpha bakewellii (White) (Thomson, 1878: 51).
Julodimorpha saundersii Thomson, 1878: 51 (Obenberger, 1926: 35).
Julodimorpha bakewellii auctorum.

Subfamily **BUPESTINAE** Lacordaire
 Buprestides Lacordaire, 1857: 33.

Tribe **BUBASTINI** Obenberger
 Bubastini Obenberger, 1920a: 89.
 Buprestes (part) Carter and Théry, 1929: 270.

NEOBUBASTES Blackburn
Neobubastes Blackburn, 1892d: 213.
Castelnaudina Obenberger, 1924b: 17.
Castelnaudia Obenberger, 1923a: 14 (preocc.).

- Neobubastes aureocincta** Blackburn
Neobubastes aureocincta Blackburn, 1892d: 213.
Neobubastes aureocincta var. *scutalis* Blackburn, 1892d: 213 (Obenberger, 1930: 303).
Castelnaudia australasiae Obenberger, 1923a: 15.

NOTOBUBASTES Carter
Notobubastes Carter, 1924a: 24.

- Notobubastes orientalis** Carter
Notobubastes orientalis Carter, 1924a: 25.

BUBASTES Laporte and Gory

Bubastes Laporte and Gory, 1836: 1.
Neraldus Théry, 1911: 17.

Bubastes globicollis Thomson

(Map 18)

Bubastes globicollis Thomson, 1879a: 14.
Bubastes simillimus Obenberger, 1922b: 81.

Tribe DICERCINI Kerremans

Dicercites Kerremans, 1903a: 124.
Buprestes (part) Carter and Théry, 1929: 270.

MICROCASTALIA Heller

Microcastalia Heller, 1891: 135.
Bubastodes Blackburn, 1892d: 212.

Microcastalia globithorax (Thomson)

(Map 19)

Castalia globithorax Thomson, 1878: 46.
Microcastalia globithorax (Thomson) (Kerremans, 1906: 421).
Bubastodes sulcicollis Blackburn, 1892d: 212.

Tribe BUPRESTINI

Buprestes (part) Carter and Théry, 1929: 270.

NEOBUPRESTIS Kerremans

Neobuprestis Kerremans, 1903a: 136.
Sphenoptera auctorum.
Strigoptera auctorum.

Neobuprestis frenchi (Blackburn)

(Map 20)

Strigoptera frenchi Blackburn, 1892a: 500.
Neobuprestis frenchi (Blackburn) (Carter, 1924b: 524).

Neobuprestis marmorata (Blackburn)

(Map 21)

Strigoptera marmorata Blackburn, 1892a: 501.
Neobuprestis marmorata (Blackburn) (Carter, 1924b: 524).

Neobuprestis peroni (Gory and Laporte)

(Map 22)

Stigmodera peroni Gory and Laporte, 1837: 48.
Neobuprestis peroni (Gory and Laporte) (Obenberger, 1930: 363).
Strigoptera australis Blackburn, 1892a: 501 (Obenberger, 1930: 363).

NASCIO Laporte and Gory

Nascio Laporte and Gory, 1837: 1.
Geronia Dejean, 1837: 89.

Nascio vetusta (Boisduval)

(Map 23)

Buprestis vetusta Boisduval, 1835: 85.
Nascio vetusta (Boisduval) (Gory and Laporte, 1837: 2).
Nascio lunaris Kerremans, 1900: 294 (Obenberger, 1930: 367).
Nascio vetusta var. *brunneipuncta* Obenberger, 1928: 204 (Obenberger, 1930: 368).

NASCIODES Kerremans

Nascioides Kerremans, 1903a: 147.

Nascioides parryi (Hope)

(Map 24)

Stigmodera parryi Hope, 1845: 103.
Nascioides parryi (Hope) (Kerremans, 1903a: 147).
Stigmodera saundersii Hope, 1846: 213 (Williams, 1987: 137).

Nascioides quadrinotatus (van de Poll)

(Map 25)

Nascio quadrinotata van de Poll, 1886a: 123.
Nascioides quadrinotatus (van de Poll) (Williams, 1987: 139).

BALTHASARELLA Obenberger

Balthasarella Obenberger, 1958: 487.

Balthasarella melandryoides Obenberger

Balthasarella melandryoides Obenberger, 1958: 489.

Tribe MELANOPHILINI Bedel

Melanophilini Bedel, 1921: 171.
Anthaxiae (part) Carter and Théry, 1929: 270.

TORRESITA Gemminger and Harold

Torresita Gemminger and Harold, 1869: 1382 (not described).
Plagiopoe Saunders. 1868b: 12 (not described, preocc.).

Torresita cuprifera (Kirby)

(Map 26)

Buprestis cuprifera Kirby, 1818b: 457.
Torresita cuprifera (Kirby) (Gemminger and Harold, 1869: 1382).
Buprestis chrysochloris Gory and Laporte, 1837: 122.
Melobasis dilatata Redtenbacher, 1867: 85.
Torresita aenea Thomson, 1879a: 23.
Torresita cuprifera var. *limbata* Carter, 1934: 258.

PSEUDANILARA Théry

Pseudanilara Théry, 1911: 32.
Neotorresita Obenberger, 1923a: 19.

Pseudanilara bicolor Carter

(Map 27)

Pseudanilara bicolor Carter, 1924b: 524.

Pseudanilara cupripes (Macleay)

(Map 28)

Anthaxia cupripes Macleay, 1872: 242.
Pseudanilara cupripes (Macleay) (Carter, 1924b: 526).
Neocuris dilaticollis Blackburn, 1892c: 42.
Melobasis laticeps Kerremans, 1898: 126.
Melanophila australasiae Kerremans, 1903b: 59.
Neotorresita achardi Obenberger, 1923a: 20.
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Stigmodera cruentata Murray, 1852: 253 (preocc.).

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Themognatha heros Gehin, 1855: 57.

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Stigmodera (Themognatha) queenslandica Obenberger, 1922b: 112 (misspelling).
- Themognatha pascoei** (Saunders)
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Stigmodera lateritia Thomson, 1879a: 30.
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- Themognatha sanguinipennis** (Gory and Laporte) (Map 99)
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- Themognatha tricolorata** (Waterhouse) (Map 102)
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- Castiarina abdominalis** (Saunders) (Map 106)
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Stigmodera laudabilis Kerremans, 1898: 146 (Barker, 1986: 28).
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Stigmodera (Castiarina) simplex Kerremans, 1903a: 210 (Barker, 1986: 28).
- Castiarina amplipennis** (Saunders) (Map 110)
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- Castiarina andersoni** (Gory and Laporte) (Map 111)
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Stigmodera verax Kerremans, 1898: 146 (Barker, 1986: 28).
Stigmodera (Castiarina) dicax Obenberger, 1922b: 119 (Barker, 1986: 28).
- Castiarina argillacea** (Carter) (Map 112)
Stigmodera (Castiarina) argillacea Carter, 1916: 126.
- Castiarina arida** (Barker)
Stigmodera (Castiarina) arida Barker, 1987: 135.
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Stigmodera melbournensis Thomson, 1879a: 34 (Barker, 1986: 28).
Stigmodera timida Kerremans, 1898: 147 (Barker, 1986: 28).
- Castiarina bella** (Saunders) (Map 116)
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Stigmodera cruentata Gory and Laporte, 1837: 29 (preocc.).
Stigmodera bella var. *dixonii* Carter, 1926: 57 (Barker, 1986: 28).
- Castiarina bifasciata** (Hope) (Map 117)
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Buprestis bicincta Boisduval, 1835: 89 (Barker, 1986: 28).
Stigmodera bicingulata Gory and Laporte, 1837: 30 (Barker, 1986: 28).
Stigmodera dejeani Gory, 1841, err. add (Barker, 1986: 28).
Stigmodera bicincta Gory, 1841: 131 (Barker, 1986: 28).
Stigmodera trispinosa Kerremans, 1890a: 43 (Barker, 1986: 28).
Stigmodera (Castiarina) bicincta var. *bina* Obenberger, 1922b: 115 (Barker, 1986: 28).
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- Castiarina brutella** (Thomson) (Map 119)
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Stigmodera uniformis Kerremans, 1898: 145 (Barker, 1986: 28).
Stigmodera (Castiarina) brutella spp. *victrix* Obenberger, 1922b: 119 (Barker, 1986: 28).
- Castiarina burnsi** (Barker) (Map 120)
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- Castiarina caillaina** (Barker) (Map 121)
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- Castiarina callubriensis** (Carter) (Map 122)
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Stigmodera felix Kerremans, 1898: 142 (Barker, 1986: 29).
- Castiarina castelnaudi** (Saunders) (Map 124)
Stigmodera castelnaudi Saunders, 1869: 9.
Stigmodera castelnaudii Thomson, 1878: 53.
Stigmodera thomsoniana Masters, 1886: 97 (Barker, 1986: 29).
Stigmodera laportei Kerremans, 1890a: 42 (Barker, 1986: 29).
Stigmodera (Castiarina) castelnaudi var. *naias* Obenberger, 1933: 104 (Barker, 1986: 29).
- Castiarina coeruleipes** (Saunders) (Map 125)
Stigmodera coeruleipes Saunders, 1869: 13.

- Stigmodera* (*Castiarina*) *coeruleipes* var. *montana* Carter, 1916: 106 (Barker, 1986: 29).
- Castiarina colorata** (Hope) (Map 126)
Stigmodera colorata Hope, 1847: 283.
- Castiarina commixta** (Carter)
Stigmodera commixta Carter, 1924a: 21.
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- Castiarina crenata** (Donovan) (Map 128)
Buprestis crenata Donovan, 1805, pl. 7, fig. 3.
Buprestis amphichroa Boisduval, 1835: 90 (Barker, 1986: 29).
Stigmodera sexspilota Gory and Laporte, 1837: 35 (Barker, 1986: 29).
Stigmodera (*Castiarina*) *amphichroa* var. *allecto* Obenberger, 1933: 105 (Barker, 1986: 29).
Stigmodera (*Castiarina*) *amphichroa* var. *euterpe* Obenberger, 1933: 105 (Barker, 1986: 29).
- Castiarina cruentata** (Kirby) (Map 129)
Buprestis cruentata Kirby, 1818b: 455.
Stigmodera neologa Thomson, 1879a: 35 (Barker, 1986: 29).
Stigmodera stillata Blackburn, 1890b: 148 (Barker, 1986: 29).
Stigmodera coelestis Kerremans, 1890a: 48 (Barker, 1986: 29).
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Stigmodera crux Saunders, 1868a: 473.
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Stigmodera (*Castiarina*) *magnetica* Carter, 1933: 161 (Barker, 1986: 29).
- Castiarina cyanipes** (Saunders) (Map 132)
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Stigmodera (*Castiarina*) *cyanipes* var. *lalage* Obenberger, 1933: 75 (Barker, 1986: 29).
- Castiarina decemmaculata** (Kirby) (Map 133)
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Stigmodera (*Castiarina*) *inaequalis* Kerremans, 1903a: 207 (Barker, 1986: 29).
Stigmodera (*Castiarina*) *decemmaculata* var. *bellula* Obenberger, 1933: 71 (Barker, 1986: 29).
- Castiarina delectabilis** (Hope) (Map 134)
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Stigmodera (*Castiarina*) *delectabilis* var. *cyaneiventris* Obenberger, 1933: 106 (Barker, 1986: 29).
Stigmodera (*Castiarina*) *delectabilis* var. *carneola* Obenberger, 1933: 106 (Barker, 1986: 29).
- Castiarina deyrollei** (Thomson) (Map 135)
Stigmodera deyrollei Thomson, 1879d: 125.
Stigmodera chobauti Théry, 1895: 328 (Barker, 1986: 30).
Stigmodera sancta Carter, 1913: 501 (Barker, 1986: 30).
- Castiarina dimidiata** (Carter) (Map 136)
Stigmodera dimidiata Carter, 1908: 422.
Stigmodera (*Castiarina*) *dorsalis* Obenberger, 1922b: 118 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *leai* var. *fasciosa* Obenberger, 1922b: 122 (Barker, 1986: 30).
- Castiarina distinguenda** (Saunders) (Map 137)
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Stigmodera distinguenda ssp. *differeus* Carter, 1931b: 364 (Barker, 1986: 30).
- Castiarina dugganensis** (Barker) (Map 138)
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- Castiarina erasma** (Carter) (Map 139)
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- Castiarina erythromelas** (Boisduval) (Map 140)
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Stigmodera longula Blackburn, 1892c: 54 (Barker, 1986: 30).
- Stigmodera* (*Castiarina*) *cicerini* Obenberger, 1928: 331 (Barker, 1986: 30).
- Castiarina erythroptera** (Boisduval) (Map 141)
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Stigmodera canaliculata Blackburn, 1892c: 51 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *erythroptera* ssp. *nigroterminata* Carter, 1934: 257 (Barker, 1986: 30).
- Castiarina flava** (Saunders) (Map 142)
Stigmodera flava Saunders, 1869: 17.
Stigmodera flavescens Masters, 1886: 86 (Barker, 1986: 30).
Stigmodera flava-Thomson, 1878: 55.
Stigmodera flavidula Kerremans, 1890a: 47 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *flava* var. *notulata* Obenberger, 1922b: 115 (Barker, 1986: 30).
- Castiarina flavopicta** (Boisduval) (Map 143)
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Stigmodera flavopicta-Gory and Laporte, 1837: 44.
Stigmodera flavovaria Saunders, 1871: 74 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *flavopicta* var. *nausicaa* Obenberger, 1933: 110 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *avopicta* var. *erato* Obenberger, 1933: 110 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *flavopicta* var. *palaeno* Obenberger, 1933: 111 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *flavopicta* var. *iris* Obenberger, 1933: 111 (Barker, 1986: 30).
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- Castiarina flavopurpurea** (Carter) (Map 144)
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- Castiarina flavosignata** (Macleay) (Map 145)
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Stigmodera (*Castiarina*) *flavosignata* var. *rufosignata* Carter, 1931b: 349 (Barker, 1986: 30).
- Castiarina fossoria** (Carter) (Map 146)
Stigmodera (*Castiarina*) *fossoria* Carter, 1927: 226.
- Castiarina fulviventris** (Macleay) (Map 147)
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Stigmodera guttigera Blackburn, 1901, p. 24 (Barker, 1986: 30).
Stigmodera (*Castiarina*) *mackayana* Carter, 1930b: 536 (Barker, 1986: 30).
- Castiarina gardnerae** (Barker) (Map 148)
Stigmodera (*Castiarina*) *gardnerae* Barker, 1987: 144.
- Castiarina gibbicollis** (Saunders) (Map 149)
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- Castiarina gordonburnsi** (Barker) (Map 150)
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- Castiarina goudiana** (Barker) (Map 151)
Stigmodera (*Castiarina*) *goudiana* Barker, 1987: 139.
- Castiarina hateleyi** (Barker) (Map 152)
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- Castiarina helmsi** (Carter) (Map 153)
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- Castiarina hilaris** (Hope) (Map 154)
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Stigmodera (*Castiarina*) *hilaris* var. *infasciata* Carter, 1933: 163 (Barker, 1986: 31).
- Castiarina hoffmanseggii** (Hope) (Map 155)
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- Castiarina hoffmanseggii* auctorum.
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Stigmodera (Castiarina) indistincta var. *acclivis* Obenberger, 1933: 76 (Barker, 1986: 31).
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Stigmodera (Castiarina) septemnotata Carter, 1916: 86 (Barker, 1986: 32).
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Stigmodera acutipennis Thomson, 1879a: 38 (Barker, 1986: 33).
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- Castiarina pulchripes** (Blackburn) (Map 197)
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- Castiarina punctiventris** (Saunders) (Map 199)
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- Castiarina recta** (Saunders) (Map 200)
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Stigmodera (*Castiarina*) *robusta* var. *unifasciatella* Obenberger, 1933: 75 (Barker, 1986: 33).
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Stigmodera (*Castiarina*) *rufipennis* var. *quadrioveolata* Obenberger, 1933: 69 (Barker, 1986: 33).
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Calochroa crucigera Hope, 1838: 162 (Barker, 1986: 33).
Stigmodera transversepicta Thomson, 1879a: 35 (Barker, 1986: 33).
Stigmodera macleayi Blackburn, 1892c: 48 (Barker, 1986: 33).
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Stigmodera (*Castiarina*) *crucioides* Obenberger, 1922b: 118 (Barker, 1986: 33).
Stigmodera (*Castiarina*) *scalaris* var. *acte* Obenberger, 1933: 108 (Barker, 1986: 33).
Stigmodera (*Castiarina*) *scalaris* var. *archianassa* Obenberger, 1933: 108 (Barker, 1986: 33).
- Castiarina scintillata** (Barker) (Map 206)
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Castiarina hopei Boheman, 1858: 61 (Barker, 1986: 34).
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Stigmodera (*Castiarina*) *variata* Kerremans, 1903a: 209 (Barker, 1986: 34).
- Castiarina simulata** (Gory and Laporte) (Map 212)
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Stigmodera (*Castiarina*) *ravilla* Obenberger, 1922b: 117 (Barker, 1986: 34).
Stigmodera (*Castiarina*) *acutangula* Obenberger, 1928: 333 (Barker, 1986: 34).
Stigmodera (*Castiarina*) *yorkensis* Obenberger, 1928: 335 (Barker, 1986: 34).
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- Castiarina skusei** (Blackburn) (Map 213)
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Stigmodera johannae Théry, 1911: 55 (Barker, 1986: 34).
- Castiarina subpura** (Blackburn) (Map 216)
Stigmodera subpura Blackburn, 1903: 307.
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Stigmodera (*Castiarina*) *subtinctoria* Carter, 1933: 159.
- Castiarina subvicina** (Barker) (Map 218)
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- Castiarina terminalis** (Kerremans) (Map 220)
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Stigmodera cordifer Kerremans, 1890a: 44 (Barker, 1986: 34).
Stigmodera colorata Kerremans, 1898: 141 (Barker, 1986: 34).
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- Castiarina triramosa** (Thomson) (Map 223)
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- Castiarina uncalata** (Barker) (Map 224)
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- Castiarina undulata** (Donovan) (Map 225)
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Castiarina laportei Boheman, 1858: 61 (Barker, 1986: 35).
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- Castiarina variegata** (Barker) (Map 226)
Stigmodera (*Castiarina*) *variegata* Barker, 1983: 162.
- Castiarina variopicta** (Thomson) (Map 227)
Stigmodera variopicta Thomson, 1878: 54.
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- Stigmodera coeruleiventris* Saunders, 1869: 20 (Barker, 1986: 35).
Stigmodera (Castiarina) vegeta var. *hopeana* Obenberger, 1933: 104 (Barker, 1986: 35).
- Castiarina vicina** (Saunders) (Map 229)
Stigmodera vicina Saunders, 1868b: 43 (replacement name).
Stigmodera bicincta Gory and Laporte, 1837: 31 (preocc.).
- Castiarina victoriensis** (Blackburn) (Map 230)
Stigmodera victoriensis Blackburn, 1890b: 152.
Stigmodera sensitiva Kerremans, 1898: 148 (Barker, 1986: 35).
- Castiarina viridolinea** (Barker) (Map 231)
Stigmodera (Castiarina) viridolinea Barker, 1986: 25.
- Castiarina vittata** (Saunders) (Map 232)
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- Castiarina wellsae** (Barker) (Map 233)
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Stigmodera sigma Kerremans, 1890a: 43 (Barker, 1986: 35).
Stigmodera confinis Kerremans, 1898: 151 (Barker, 1986: 35).
Stigmodera (Castiarina) wilsoni ssp. *septentrionis* Obenberger, 1922b: 16 (Barker, 1986: 35).
- Castiarina xanthopilosa** (Hope) (Map 235)
Stigmodera xanthopilosa Hope, 1847: 283.
Castiarina splendida Gehin, 1855: 64 (Barker, 1986: 35).
Stigmodera (Castiarina) xanthopilosa var. *subfascigera* Obenberger, 1933: 70 (misspelling).
Stigmodera (Castiarina) xanthopilosa var. *dichroptera* Obenberger, 1933: 70 (misspelling).
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- Tribe **CHRYSOBOTHRIINI**
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- CHRYSOBOTHRIIS** Eschscholtz
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Amblis Gistel, 1834: 10.
Odontomus Kirby, 1837: 156.
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- Chrysobothris mastersii** Macleay (Map 236)
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Chrysobothris blackburni Obenberger, 1923b: 77 (Carter, 1940: 384).
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- Tribe **ACTENODINI** Kerremans
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Merimna corporaali Obenberger, 1924a: 69.
- Subfamily **AGRILINAE** Gory and Laporte
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 Agrili Carter and Théry, 1929: 268.
- Tribe **COROEBINI** Bedel
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- Synechocera tasmanica** Théry (Map 238)
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Paracephala cylindrica Kerremans, 1898: 177 (Carter, 1940: 388).
- Paracephala pistacina** (Hope) (Map 240)
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Paracephala pistacina (Hope) (Saunders, 1868b: 63).
Aphanisticus canaliculatus Germar, 1848: 180 (Carter, 1940: 388).
Paracephala minuta Kerremans, 1898: 177 (Carter, 1940: 388).
- MELIBOEITHON** Obenberger
Meliboeithon Obenberger, 1920b: 170.
- Meliboeithon intermedium** (Kerremans) (Map 241)
Paracephala intermedia Kerremans, 1898: 177.
Meliboeithon intermedium (Kerremans) (Bellamy, 1988: 434).
Meliboeithon fissus Obenberger, 1920b: 171 (Bellamy, 1988: 434).
- DINOCEPHALIA** Obenberger
Dinocephalia Obenberger, 1923a: 39.
Pseudosynechocera Obenberger, 1943: 33.
- Dinocephalia burnsi** Bellamy (Map 242)
Dinocephalia burnsi Bellamy, 1988: 449.
- Dinocephalia cyaneipennis** (Blackburn) (Map 243)
Paracephala cyaneipennis Blackburn, 1893: 130.
Dinocephalia cyaneipennis (Blackburn) (Bellamy, 1988: 445).
- Dinocephalia thoracica** (Kerremans) (Map 244)
Paracephala thoracica Kerremans, 1900: 343.
Dinocephalia thoracica (Kerremans) (Bellamy, 1988: 442).
Dinocephalia gigantea Obenberger, 1923a: 40 (Bellamy, 1988: 442).
- Dinocephalia transsecta** (Carter) (Map 245)
Paracephala transsecta Carter, 1921: 306.
Dinocephalia transsecta (Carter) (Bellamy, 1988: 446).
Paracephala impressicollis Obenberger, 1924 a: 155. Bellamy, 1988: 447).
- ALCINOUS** Deyrolle
Alcinous Deyrolle, 1864: 115.
Cisseis (part) Gory and Laporte, 1839: 1.
- Alcinous fossicollis** (Kerremans) (Map 246)
Cisseis fossicollis Kerremans, 1903a: 229.
Alcinous fossicollis (Kerremans) (Carter and Théry, 1929: 272, 276).
- Alcinous nodosus** Kerremans (Map 247)
Alcinous nodosus Kerremans, 1898: 175.
Alcinous minor Kerremans, 1898: 176.
- CISSEIS** Gory and Laporte
Cisseis Gory and Laporte, 1839: 1.
Diphucrania (part) Dejean, 1833 (preocc.).
Cinyra auctorum.
Coraeus auctorum.
- Cisseis acuducta** (Kirby) (Map 248)
Trachys acuducta Kirby, 1837: 162.
Cisseis acuducta (Kirby) (Saunders, 1868b: 60).
Ethon marmoreum Gory and Laporte, 1839: 3.
Cisseis aenea Gemminger and Harold, 1869: 1429.
Cisseis cuprifrons Kerremans, 1898: 157.

- Cisseis laeta* Kerremans, 1903a: 227.
Cisseis constricta Blackburn (Map 249)
Cisseis constricta Blackburn, 1887: 254.
Cisseis lindi Blackburn, 1887: 254.
Cisseis cupreicollis (Hope) (Map 250)
Ethon cupreicollis Hope, 1846: 219.
Cisseis cupreicollis (Hope) (Saunders, 1868b: 58).
Ethon aeneicollis Hope, 1846: 220.
Cisseis morosa Kerremans, 1898: 158.
Cisseis cupripennis (Guérin) (Map 251)
Buprestis cupripennis Guérin, 1830: 65.
Cisseis cupripennis (Guérin) (Gory and Laporte, 1839: 5).
Cisseis duodecimmaculata (Fabricius) (Map 252)
Buprestis duodecimmaculata Fabricius, 1801: 191.
Cisseis duodecimmaculata (Fabricius) (Saunders, 1871: 103).
Buprestis duodecimguttata Boisduval, 1835: 93.
Cisseis quatuordecimnotata Hope, 1846: 218.
Cisseis xanthosticta Saunders, 1871: 103.
Cisseis pustulata Thomson, 1879a: 51.
Cisseis duodecimmaculata var. *fallaciosula* Obenberger, 1935a: 36.
Cisseis fulgidicollis Macleay (Map 253)
Cisseis fulgidicollis Macleay, 1888: 1231.
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Cisseis leucosticta (Kirby) (Saunders, 1868b: 57).
Cisseis stellulata Dalman, 1823: 54.
Cisseis stellatula Kerremans, 1892: 225 (lapsus).
Cisseis fulgidifrons Kerremans, 1898: 161.
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Cisseis marmorata Gory and Laporte, 1839: 4.
Cisseis similis Saunders, 1868b: 59.
Cisseis viridicollis Thomson, 1879a: 50.
Cisseis aenea Kerremans, 1898: 158.
Cisseis marmorata var. *prasina* Carter, 1923a: 168.
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Cisseis nitidicollis Kerremans (Map 257)
Cisseis nitidicollis Kerremans, 1898: 162.
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Cisseis notulata (Germar) (Saunders, 1871: 103).
Cisseis notula Kerremans, 1892: 226 (Obenberger, 1935b: 850).
Cisseis inops Kerremans, 1898: 168.
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Cisseis parva Blackburn, 1887: 253.
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Cisseis rubicunda Kerremans, 1898: 169.
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Cisseis subbifascialis Carter, 1927: 229.
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Cisseis uniformis Thomson, 1879a: 53.
Cisseis coraeboides Kerremans, 1898: 166.
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Pachycisseis Théry, 1929: 268 (to genus: Bellamy, 1988: 417).
Pachycisseis bicolor (Gory and Laporte) (Map 272)
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Neospades Blackburn, 1887: 251 (to genus: Bellamy, 1988: 417).
Neospades chrysopygia (Germar) (Map 273)
Coraeus chrysopygius Germar, 1848: 178.
Cisseis dimidiata Macleay, 1872: 248.
Cisseis semirugosa Thomson, 1879a: 51.
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Cisseis apicalis Macleay, 1888: 1232.
Cisseis purpureotincta Macleay, 1888: 1232.
Neospades simplex Blackburn
Neospades simplex Blackburn, 1888: 861.
Cisseis nigripennis Macleay, 1888: 1231.
Cisseis bella Blackburn, 1891: 298.
Cisseis ignicollis Kerremans, 1898: 164.

HYPOCISSEIS Thomson
Hypocisseis Thomson, 1879a: 49.
Maschaliz Waterhouse, 1887b: 293.
Cisseoides Kerremans, 1893: 118.
Brachycisseis (part) Théry, 1931: 26.
Coroebus auctorum.
Hypocisseis minuta Carter (Map 274)
Hypocisseis minuta Carter, 1923a: 175.
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Hypocisseis ornata Carter, 1923a: 175.
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Cisseoides albopicta Kerremans, 1898: 171.
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Ethon Gory and Laporte, 1839: 1.
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Ethon affine Gory and Laporte, 1839: 4.
Diphucrania reichei Chevrolat, 1838: 82.
Ethon proximum Boheman, 1858: 63.

Ethon aurifluum Saunders, 1868b: 56.

Ethon purpurascens Saunders, 1868b: 56.

The publication date of Gory and Laporte's monograph is uncertain (Bellamy, 1985; Gardner, 1990) but Chevrolat's paper, often cited as being published in 1837, bears the year 1838 on the title page (von Hayek, pers. comm.). *Ethon affine* is used here for the sake of stability although *reichei* is the senior epithet.

Ethon corpulentum Boheman (Map 278)

Ethon corpulentum Boheman, 1858: 62 (replacement name).

Buprestis fissiceps Laporte and Gory, 1839: 4 (preocc.).

Ethon fissiceps (Kirby) (Map 279)

Buprestis fissiceps Kirby, 1818b: 458.

Ethon fissiceps (Kirby) (Gory and Laporte, 1839: 4).

Ethon viride Gory and Laporte, 1839: 6.

Ethon diversum Kerremans, 1898: 156.

Ethon leai Carter (Map 280)

Ethon leai Carter, 1924a: 26.

Ethon maculatum Blackburn (Map 281)

Ethon maculatus Blackburn, 1887: 250.

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AGRILUS Curtis

Agrilus Curtis, 1825, No. 67.

Anambus Thomson, 1864: 38.

Engyaulus Waterhouse, 1889: 50.

Teres Harris, 1829: 2.

Uragrilus Semenov-Tian-Shianskij, 1935: 276.

Paradormorphus Waterhouse, 1887a: 183.

Agrilus australasiae Gory and Laporte (Map 282)

Agrilus australasiae Gory and Laporte, 1839: 21.

Agrilus hypoleucus Gory and Laporte, 1839: 37.

Agrilus assimilis Hope, 1846: 217.

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Agrilus flavo-taeniatus Thomson, 1879a: 73.

Agrilus hypoleucus ssp. *cooki* Obenberger, 1923b: 77.

Agrilus hypoleucus ssp. *tasmanicus* Obenberger, 1923b: 77.

Agrilus danesi Obenberger, 1923b: 79.

Agrilus domini Obenberger, 1923b: 78.

Agrilus raphelisi Obenberger, 1923b: 78.

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Agrilus deauratus Macleay, 1872: 249.

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Agrilus frenchi Blackburn, 1891: 302.

Subfamily TRACHYINAE Gory and Laporte

Trachysites Gory and Laporte, 1840: 1.

Traches Carter and Théry, 1929: 269.

Tribe GERMARICINI Cobos

Germaricini Cobos, 1979: 428.

GERMARICA Blackburn

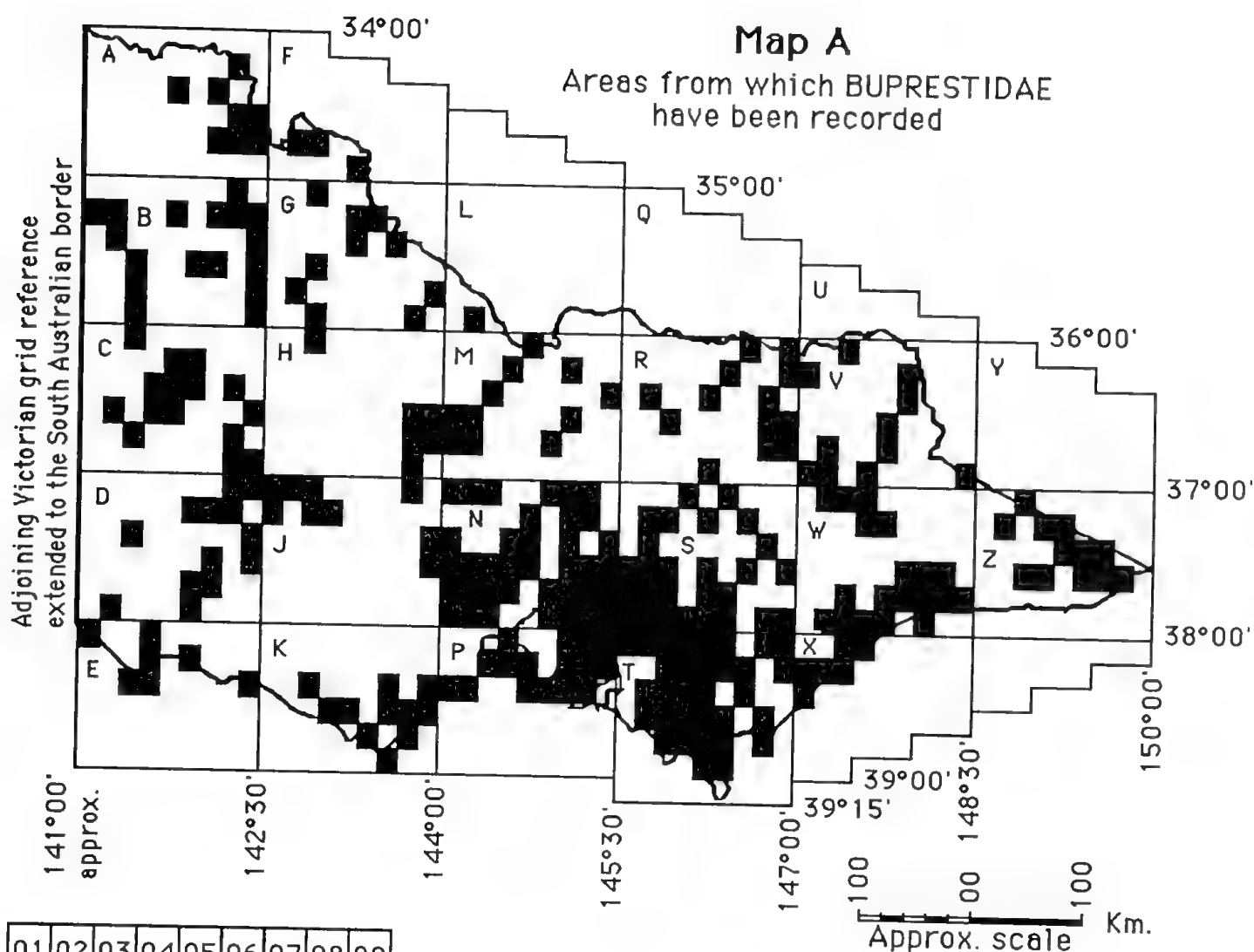
Germarica Blackburn, 1887: 257.

Germarica lilliputana (Thomson) (Map 284)

Aphanisticus lilliputanus Thomson, 1879a: 75.

Germarica lilliputana (Thomson) (Carter, 1924a: 30).

Germarica casuarinae Blackburn, 1887: 257.



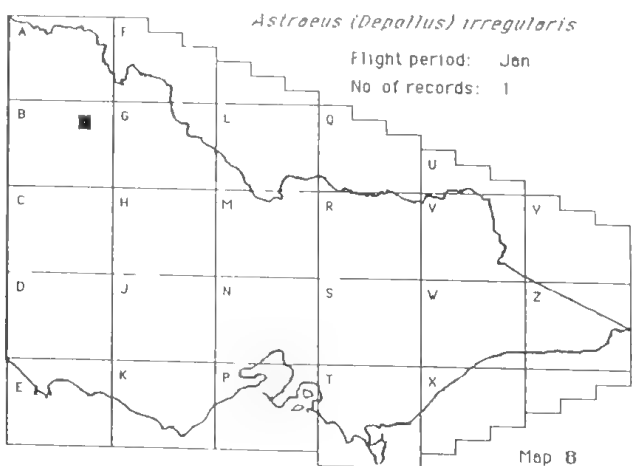
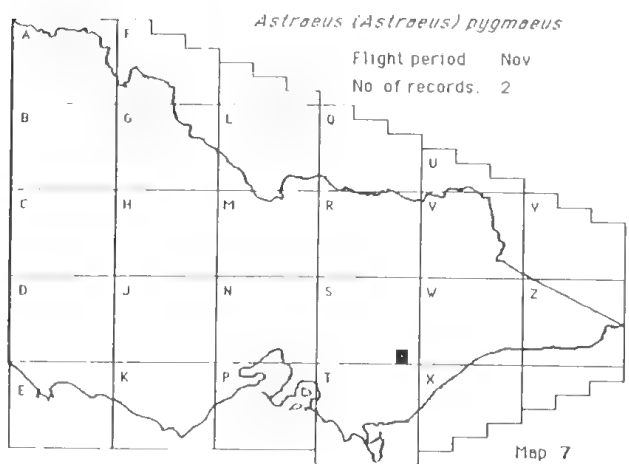
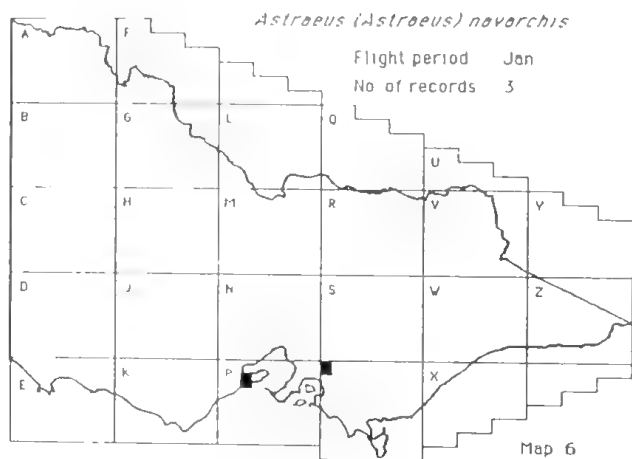
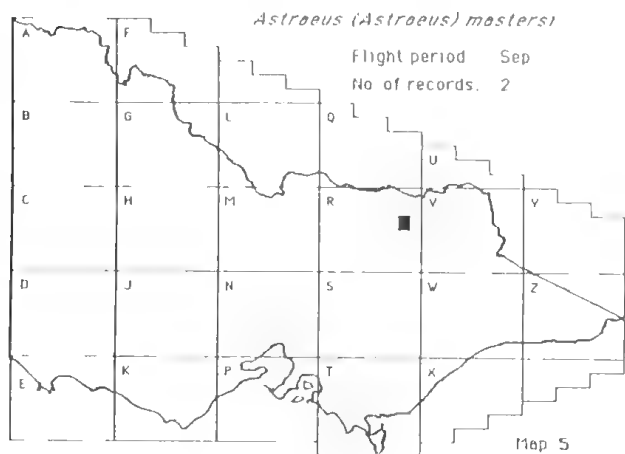
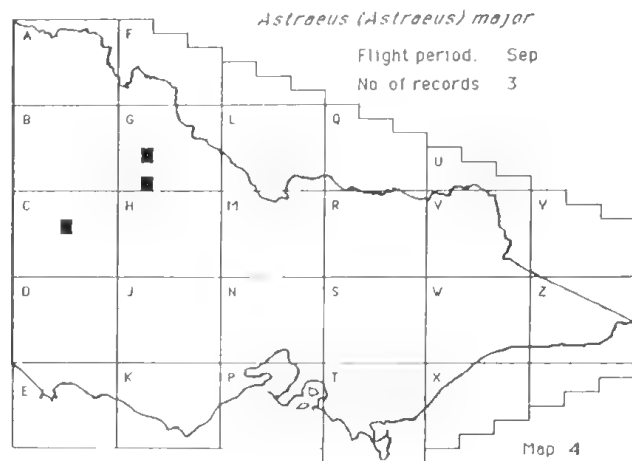
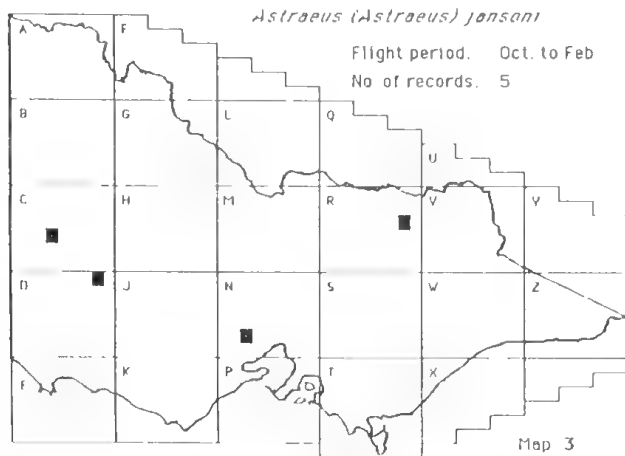
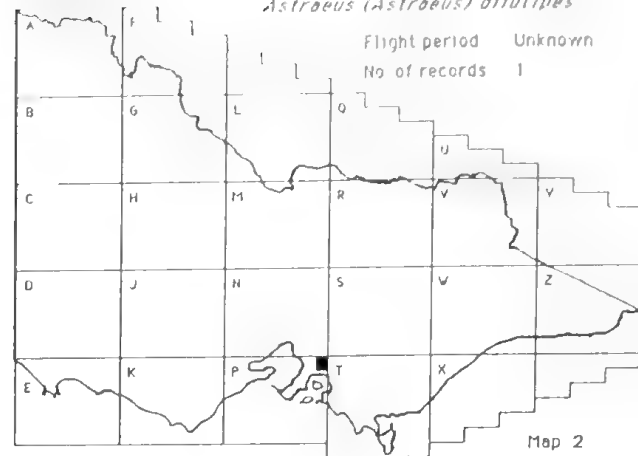
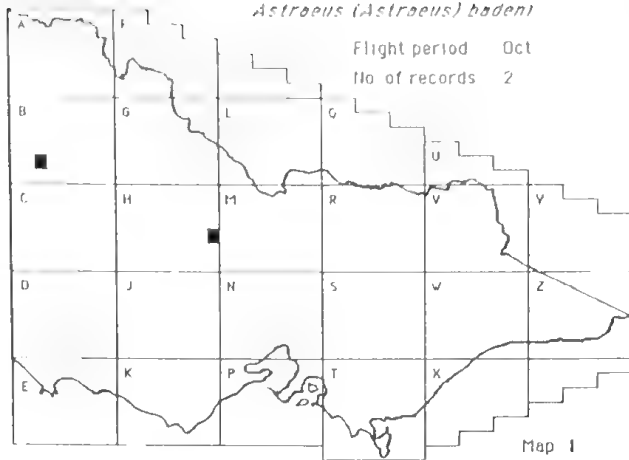
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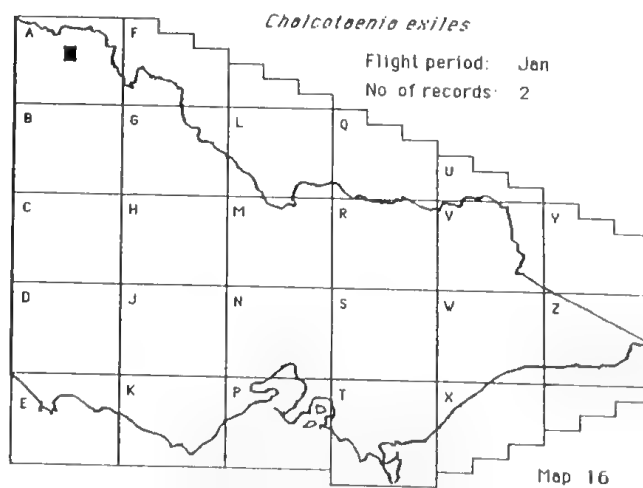
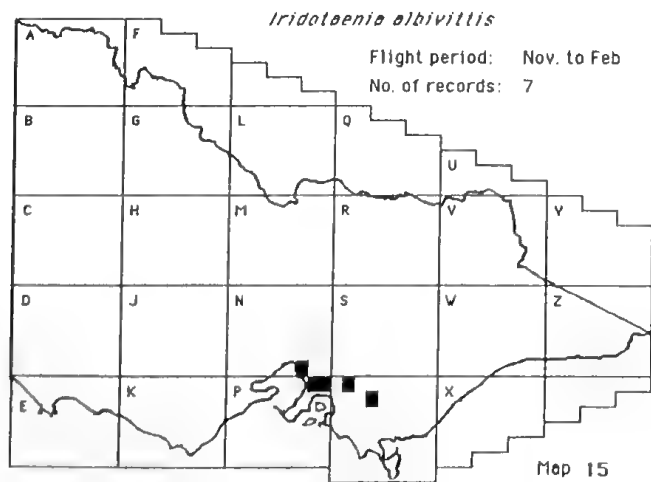
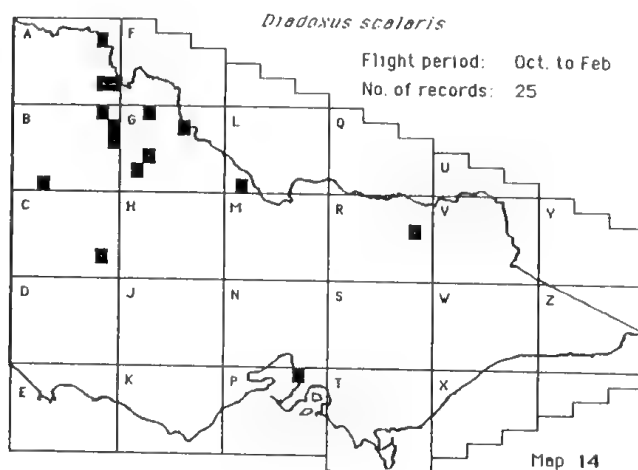
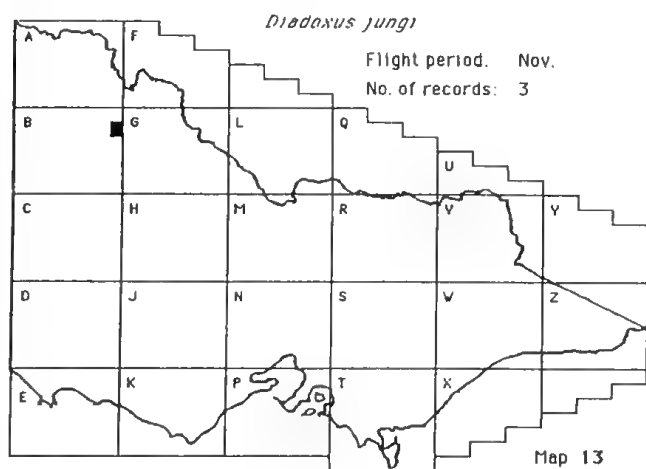
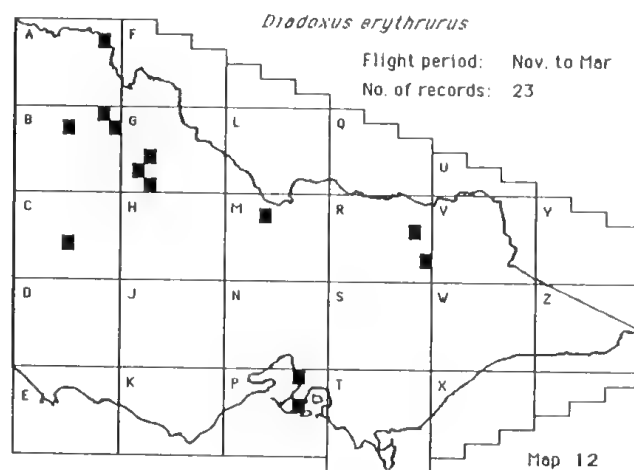
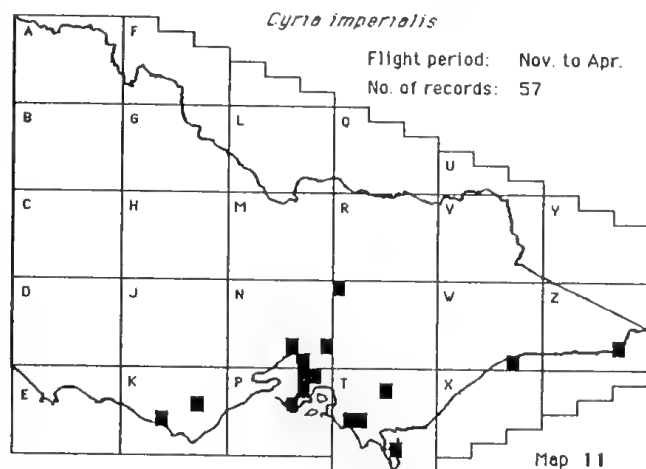
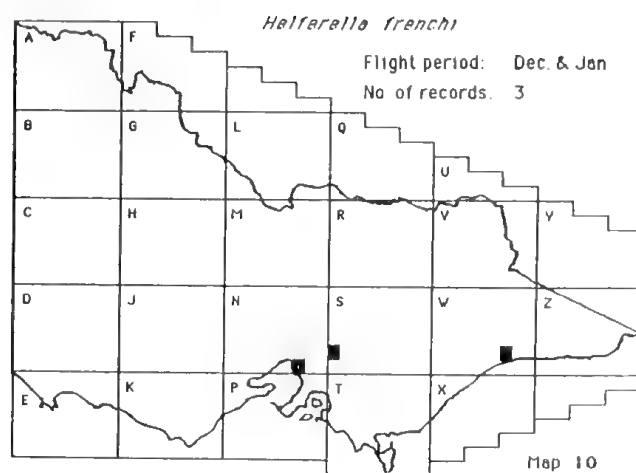
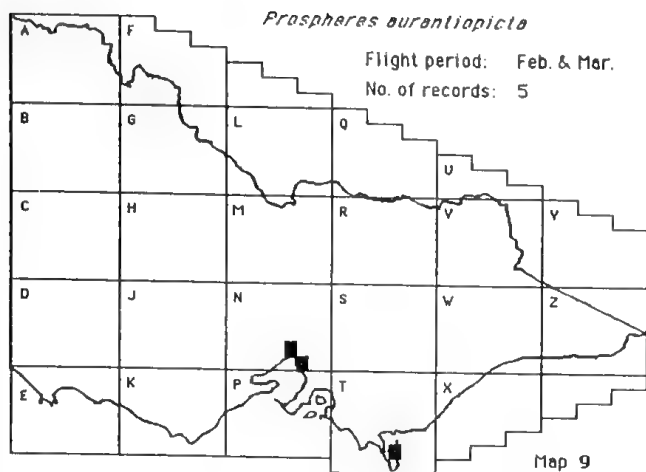
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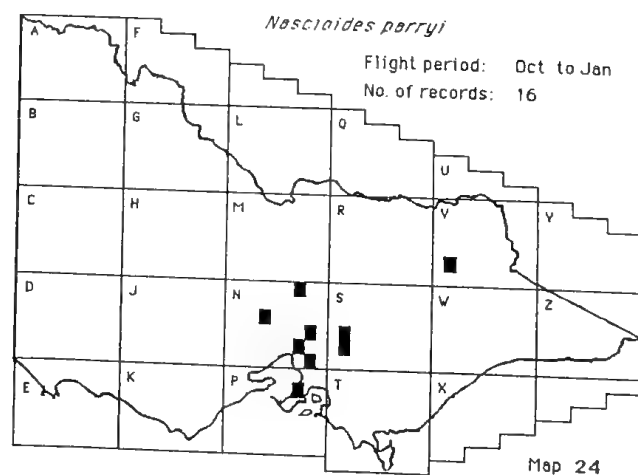
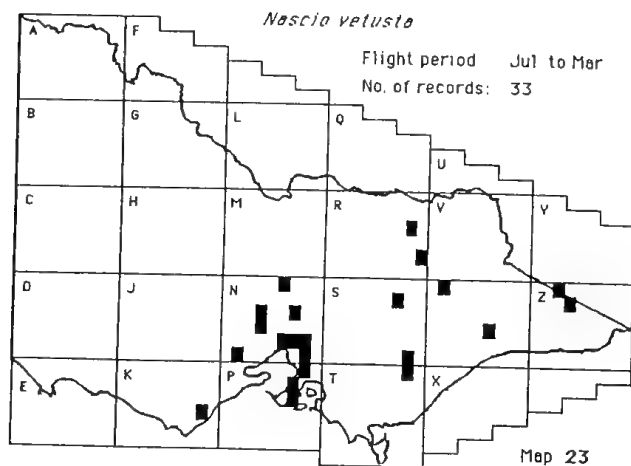
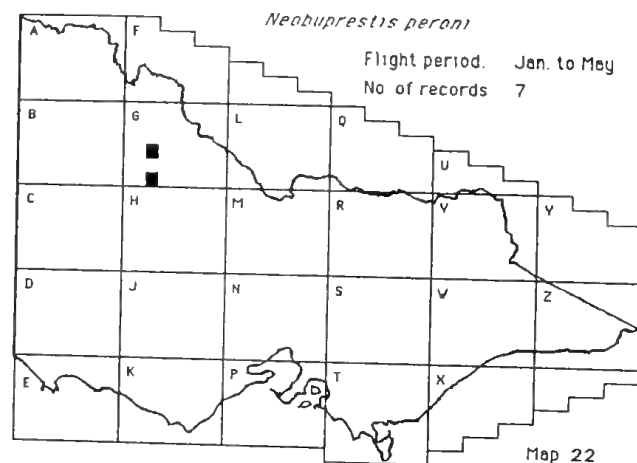
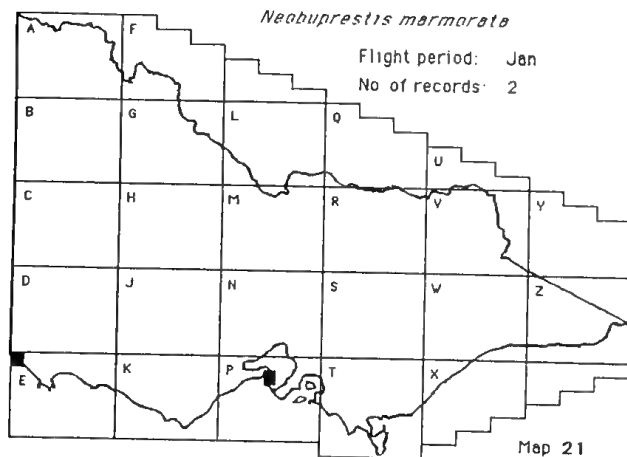
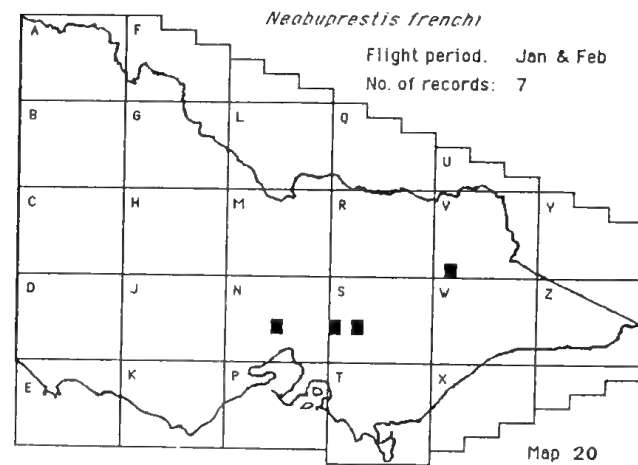
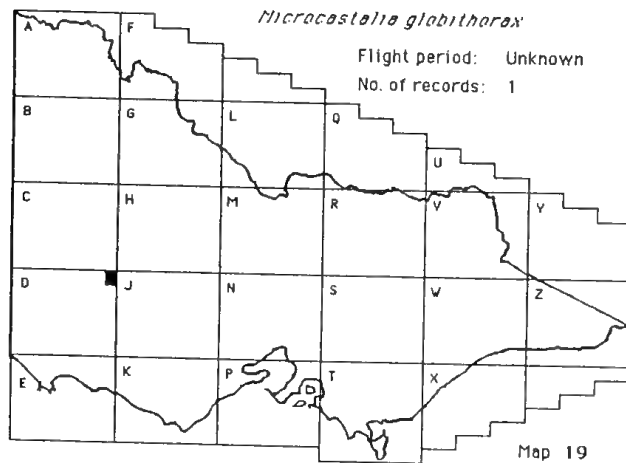
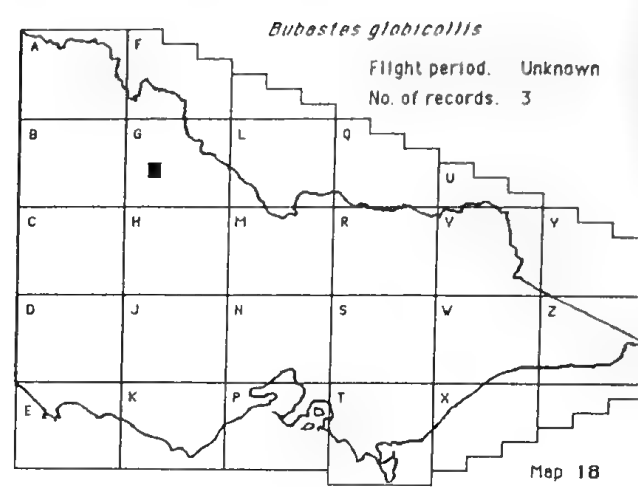
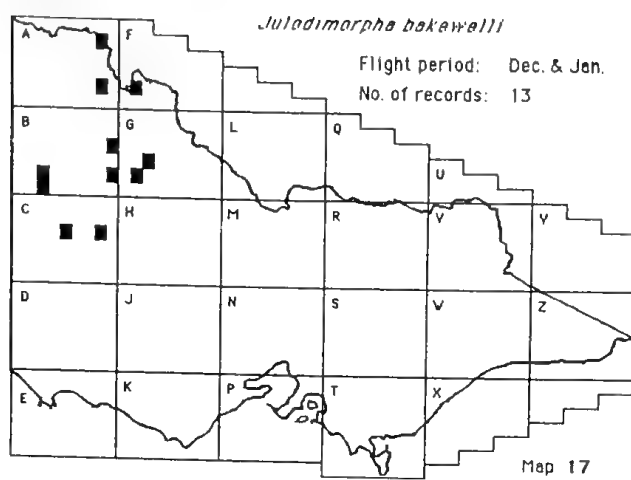
Map A. Areas from which Buprestidae have been recorded.

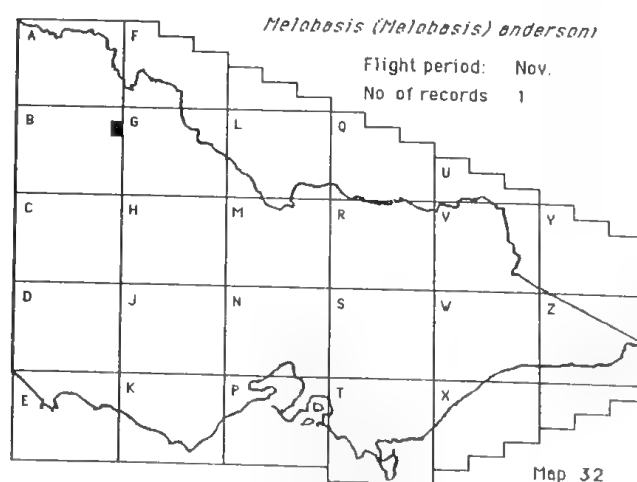
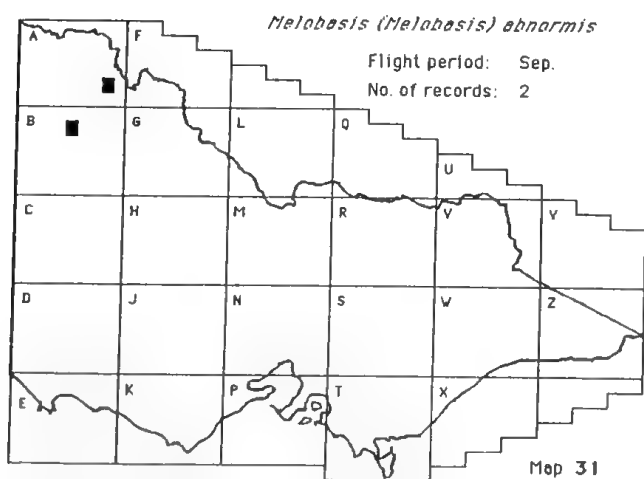
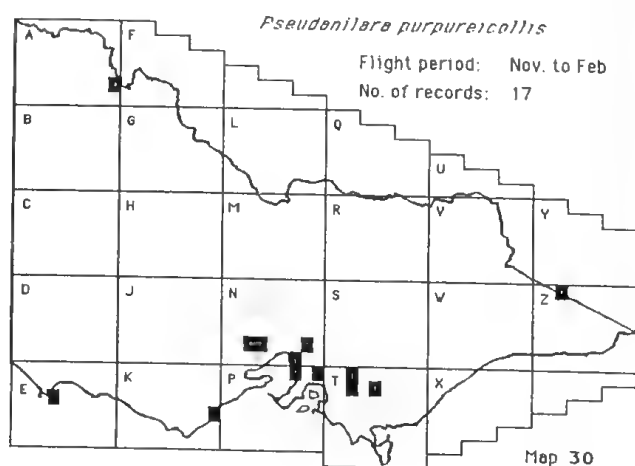
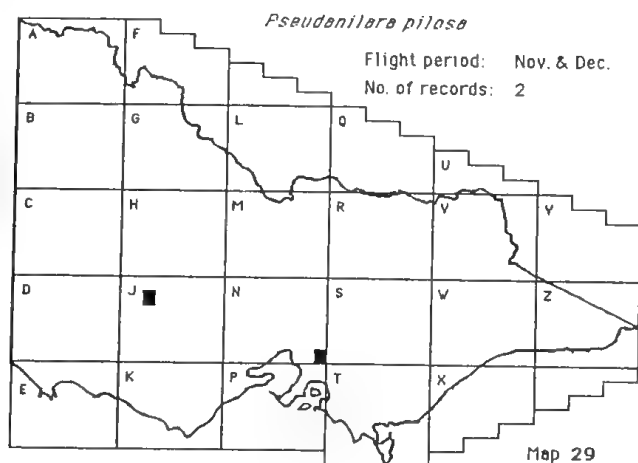
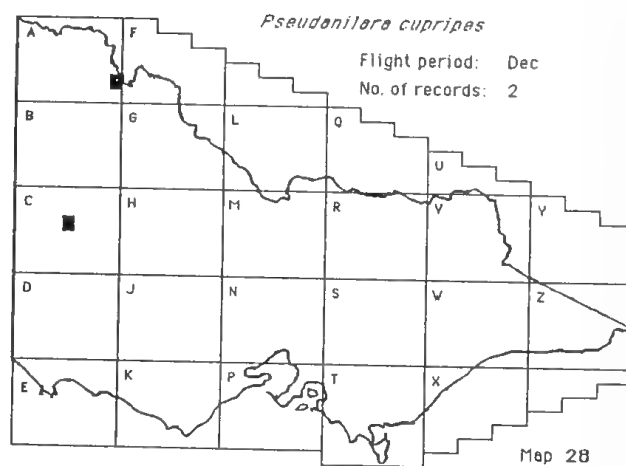
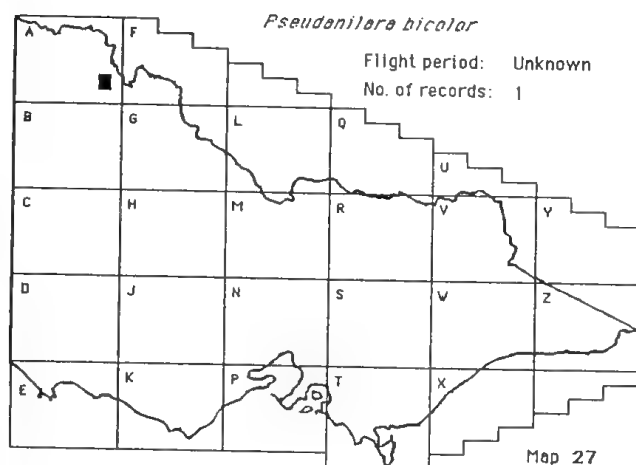
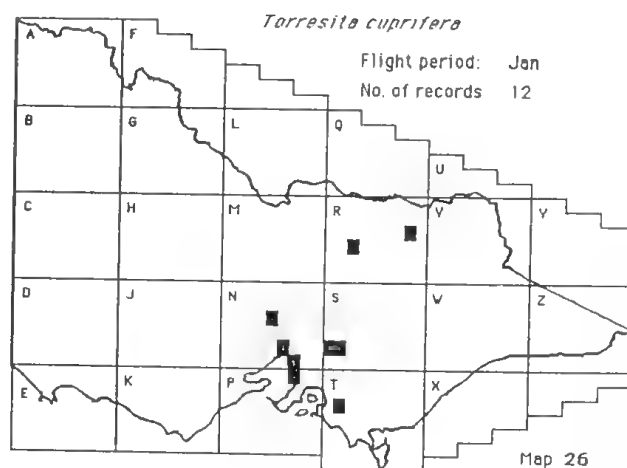
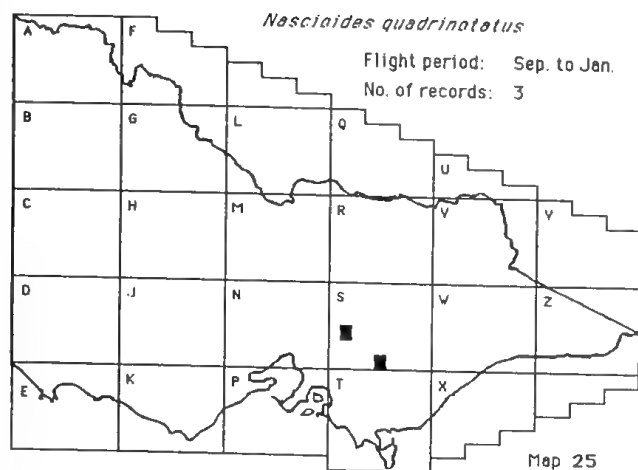
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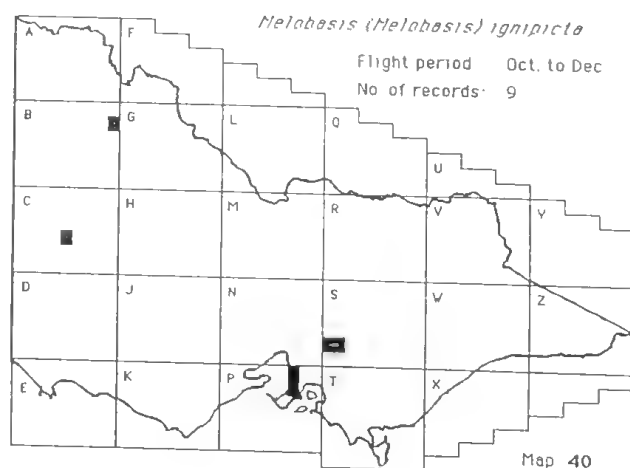
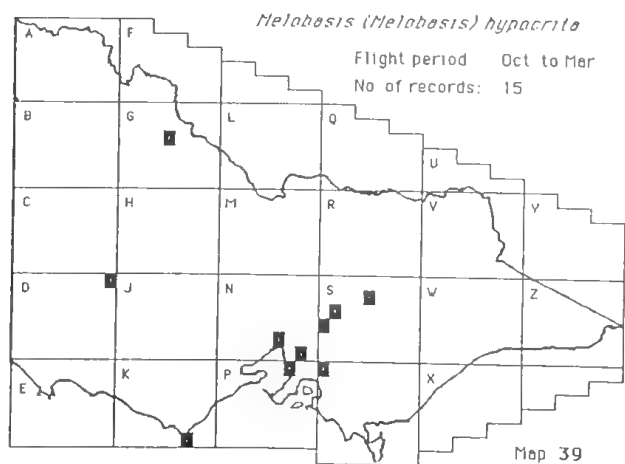
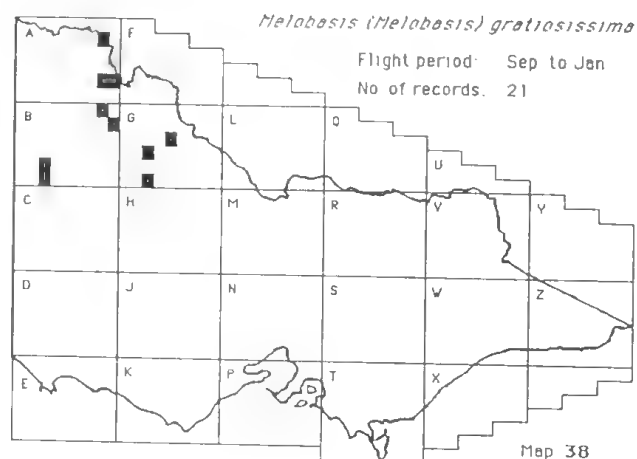
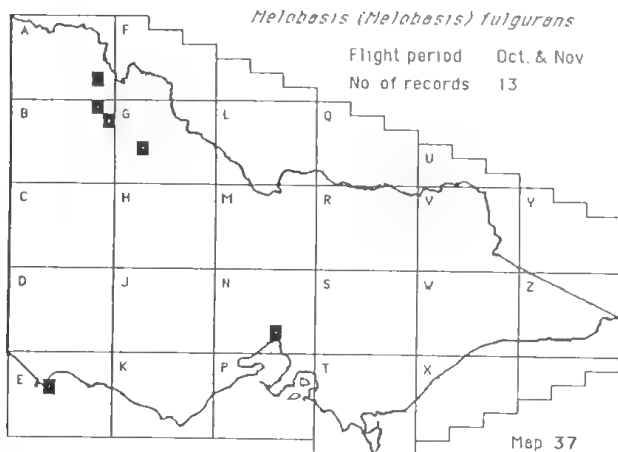
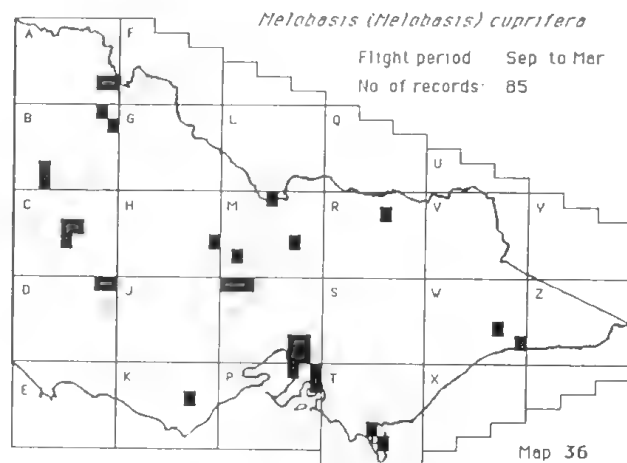
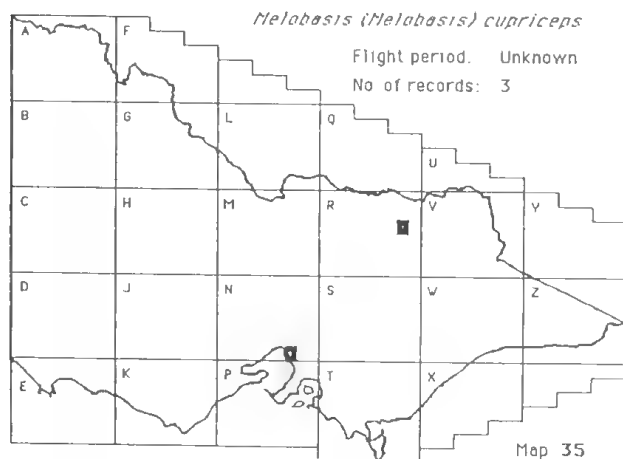
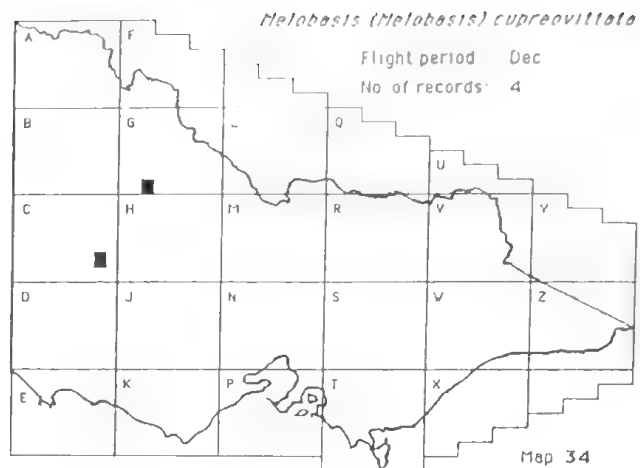
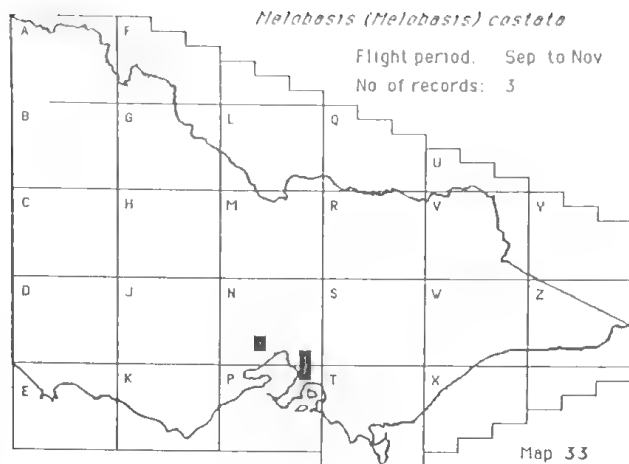
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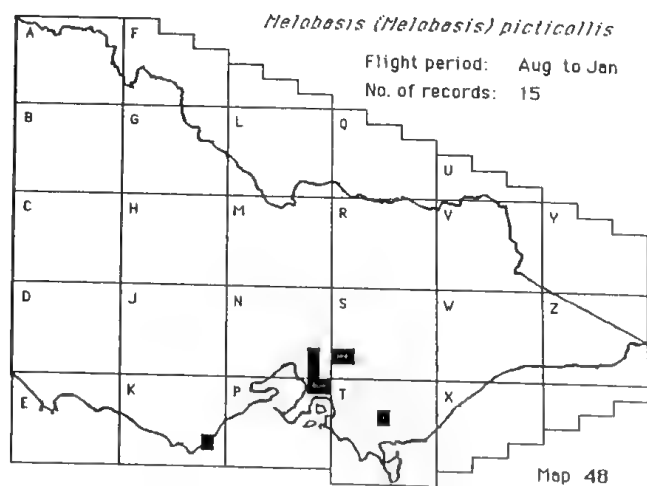
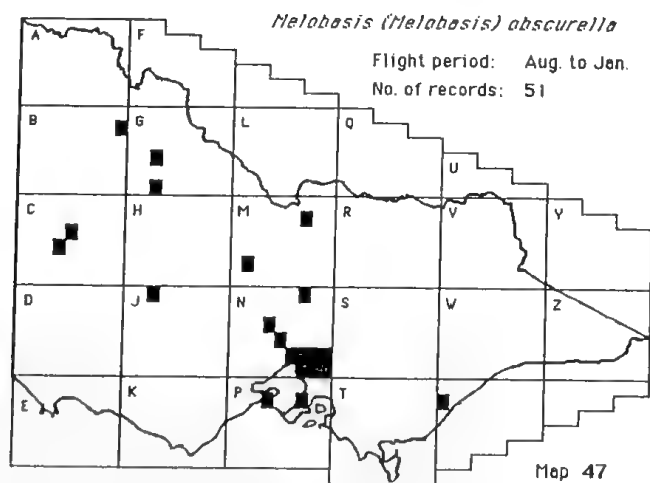
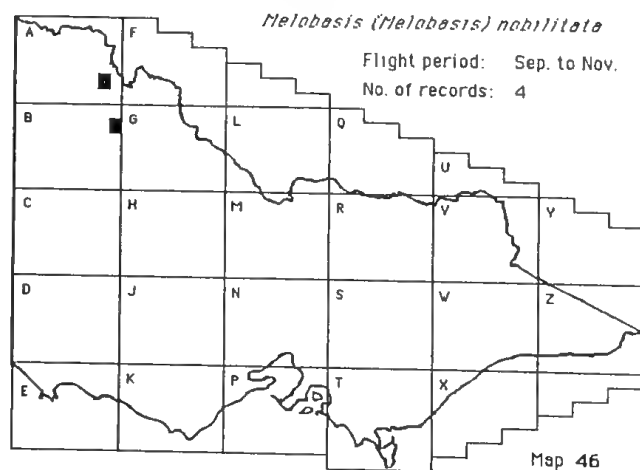
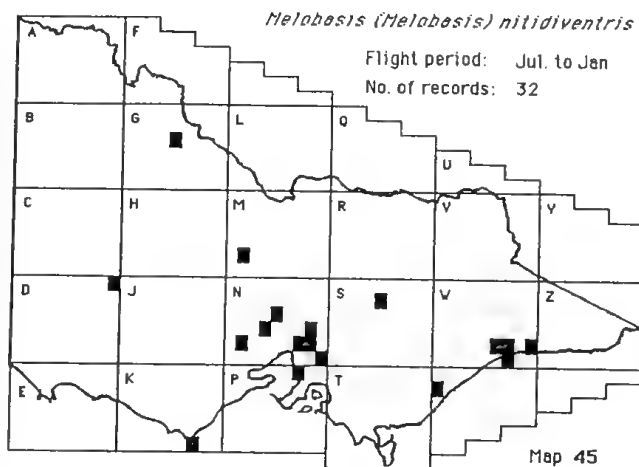
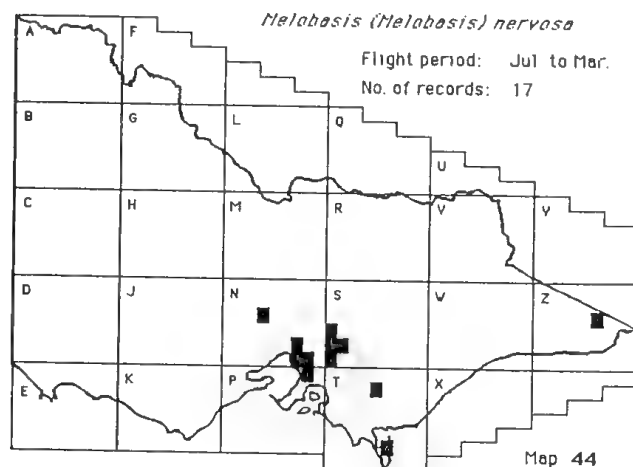
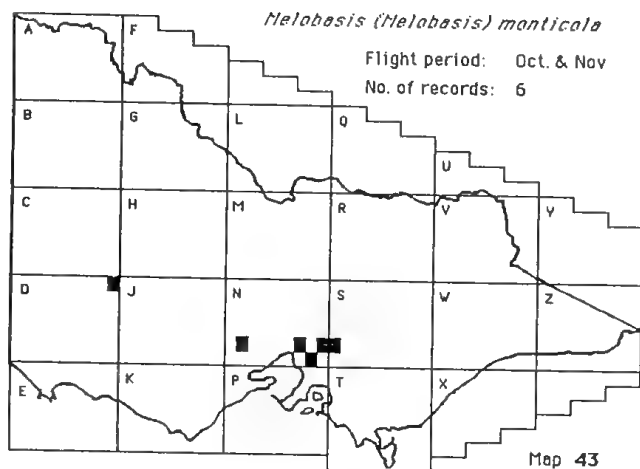
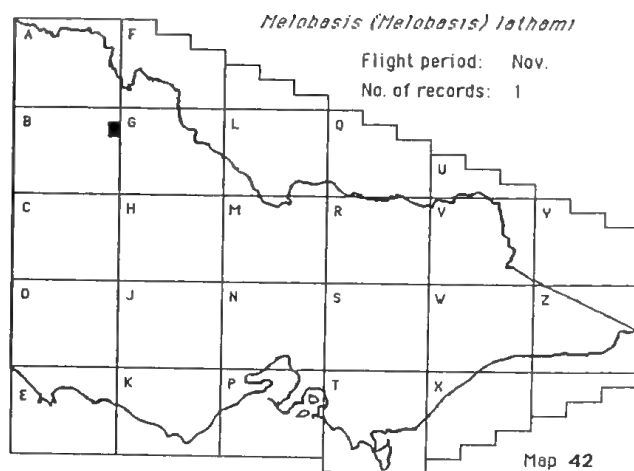
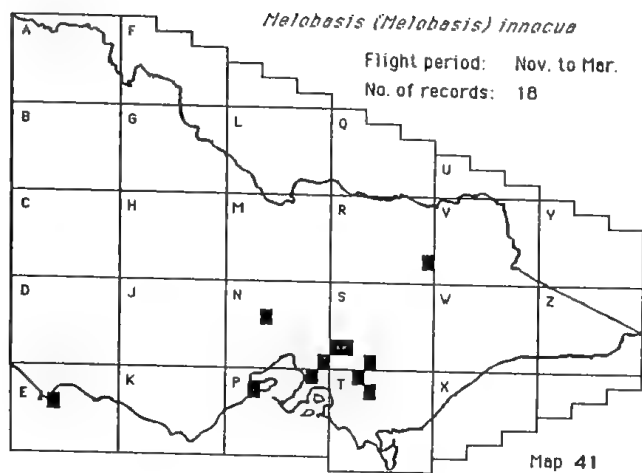


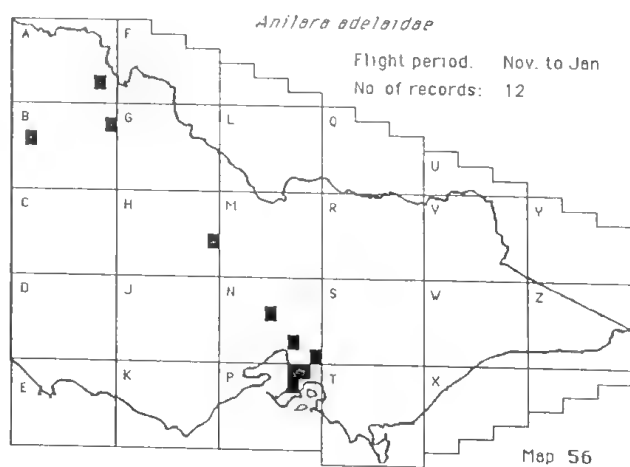
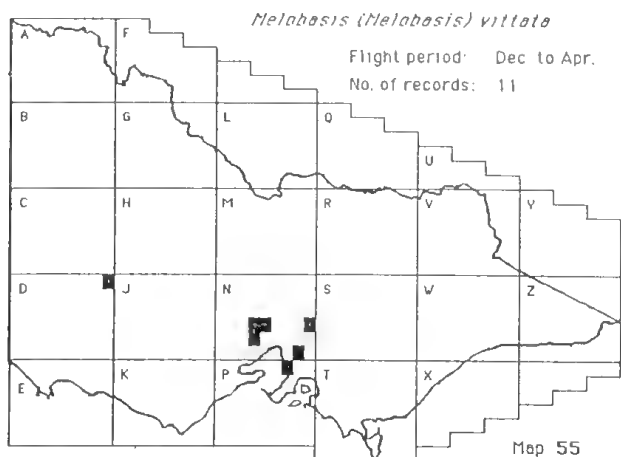
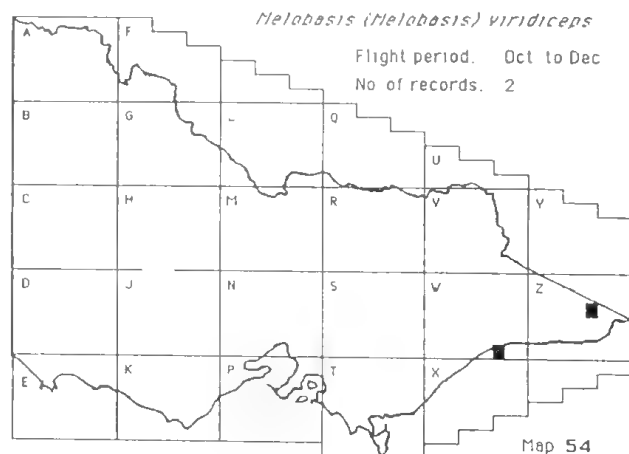
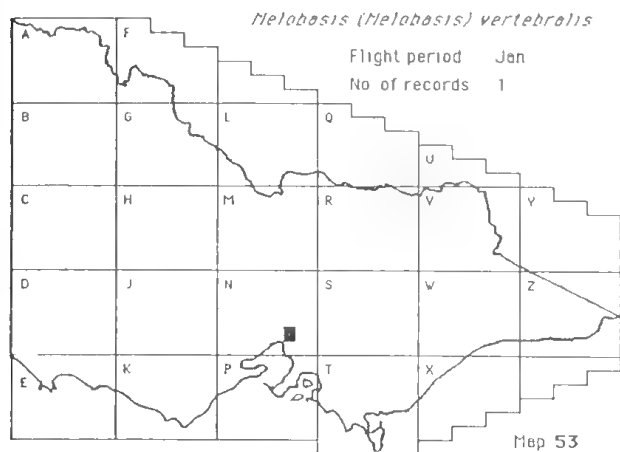
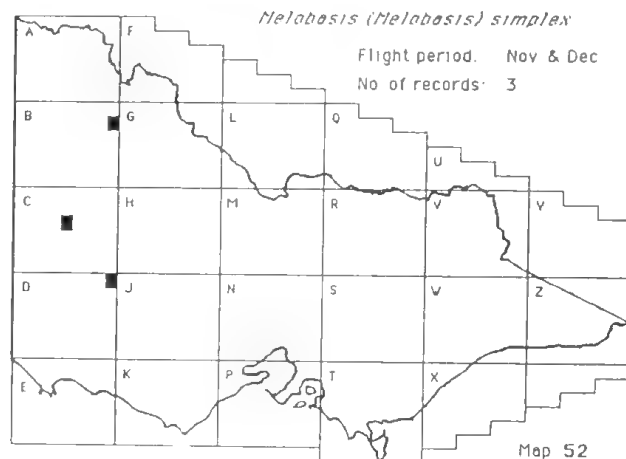
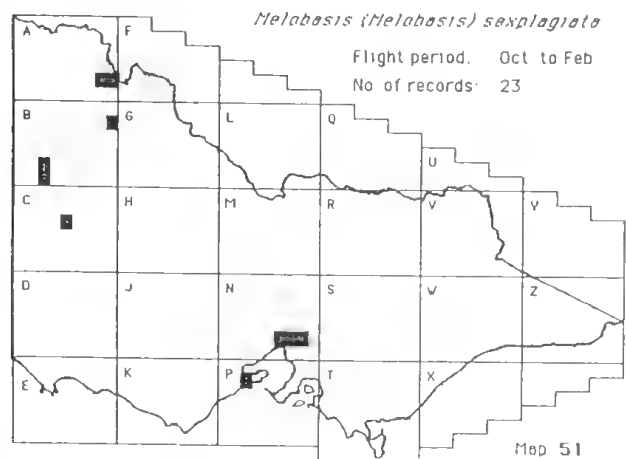
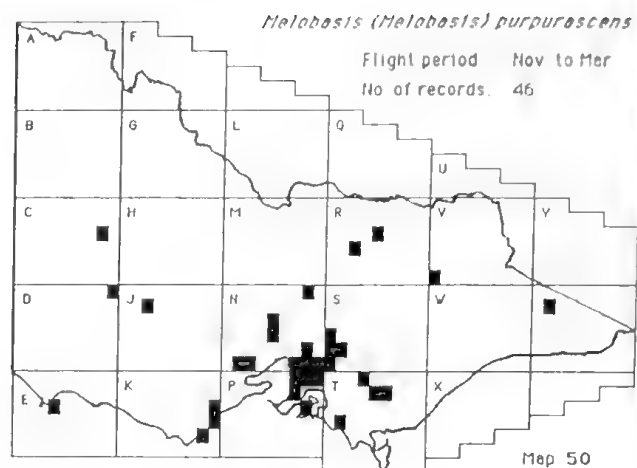
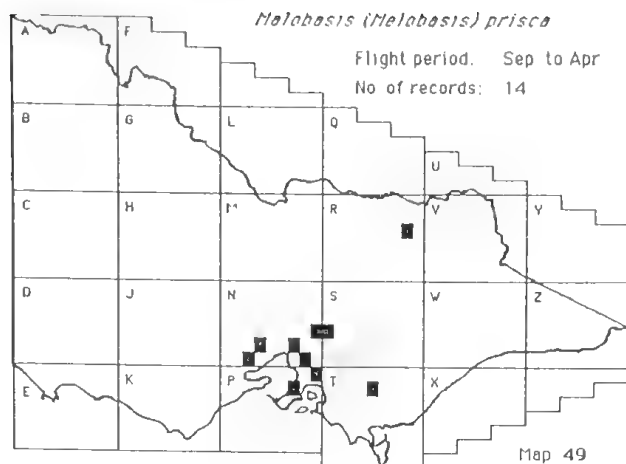


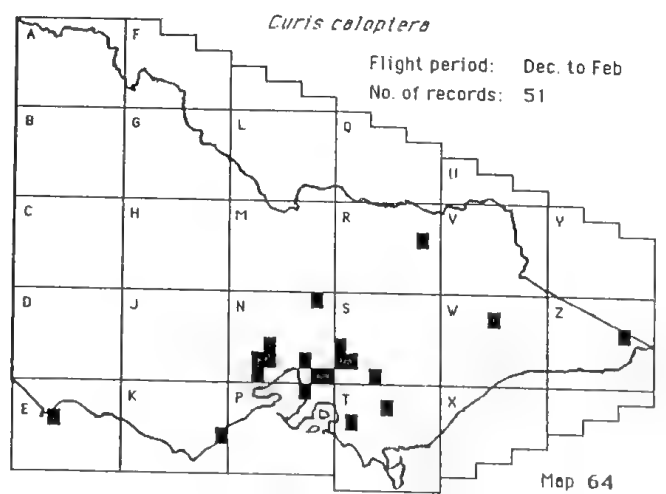
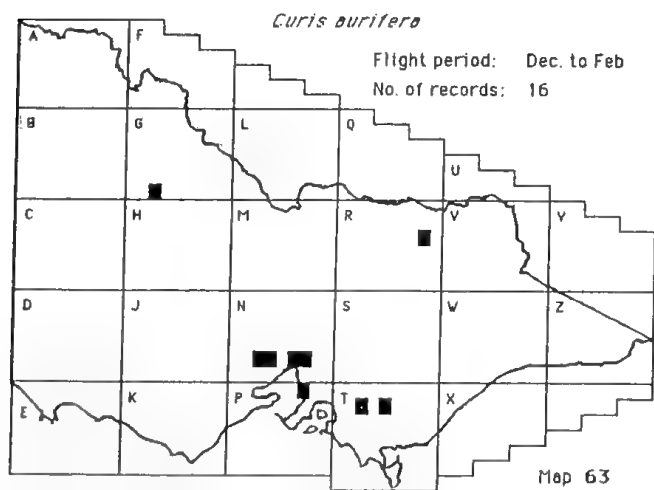
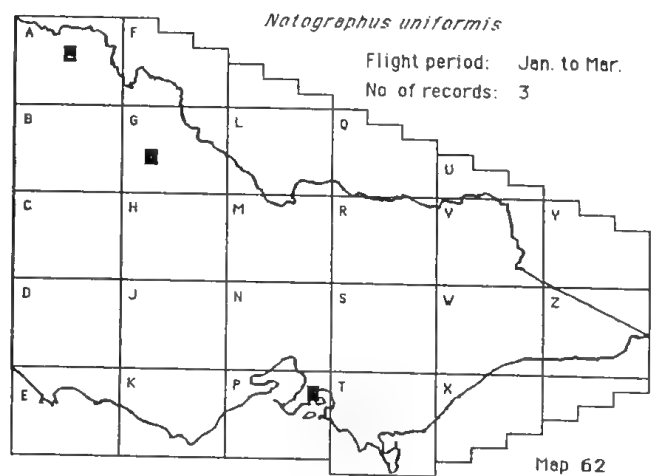
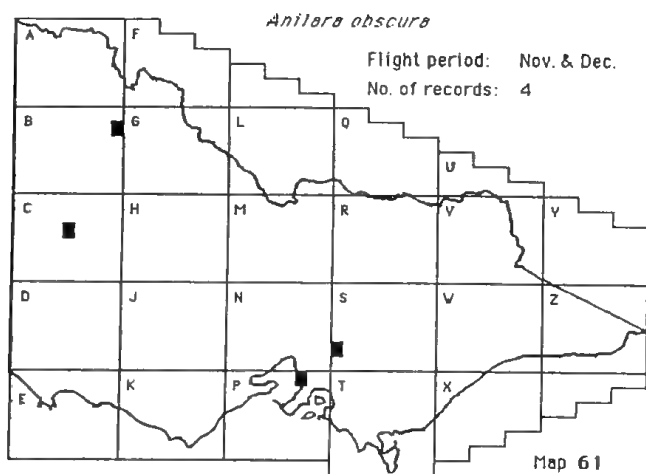
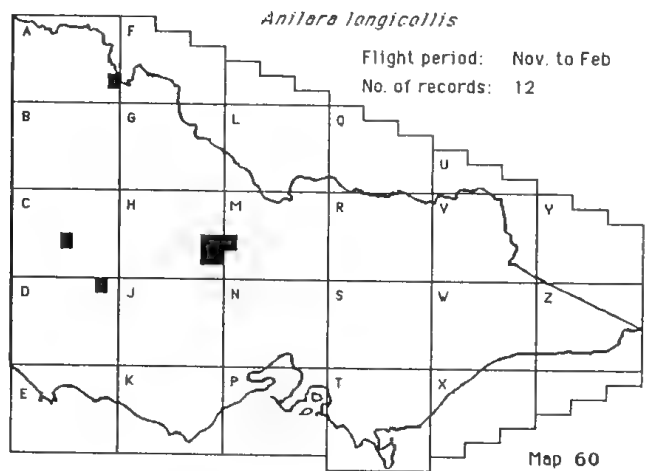
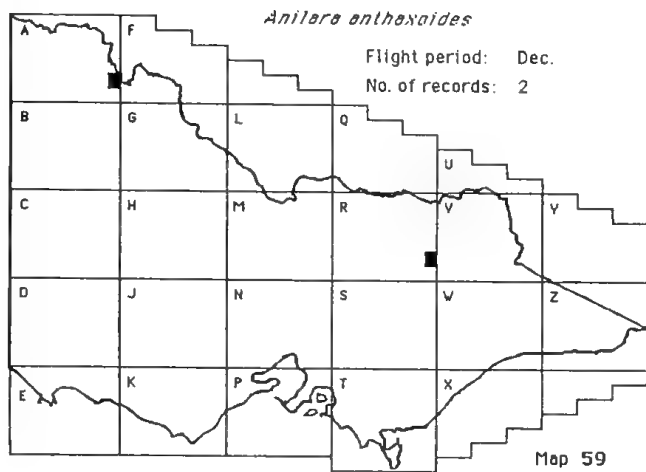
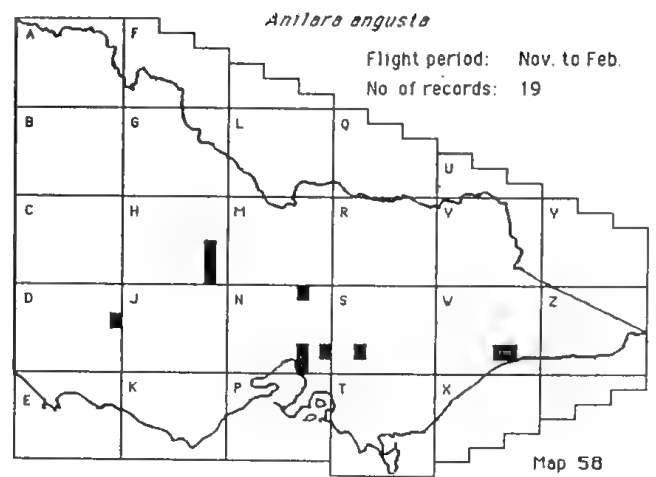
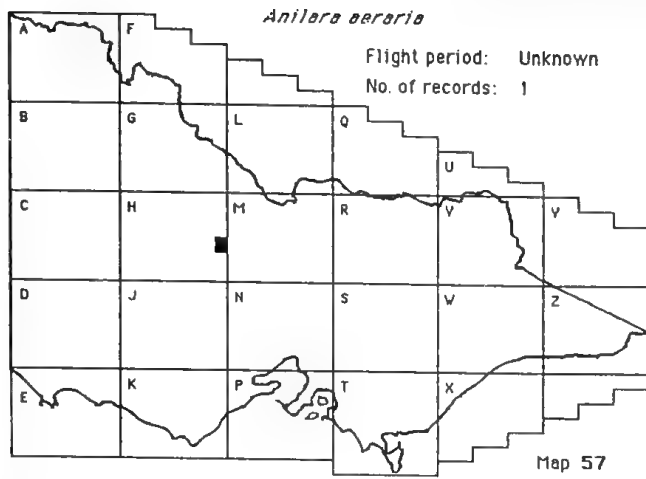


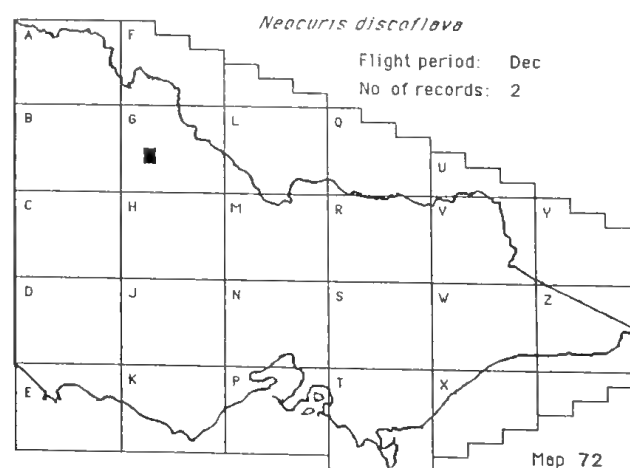
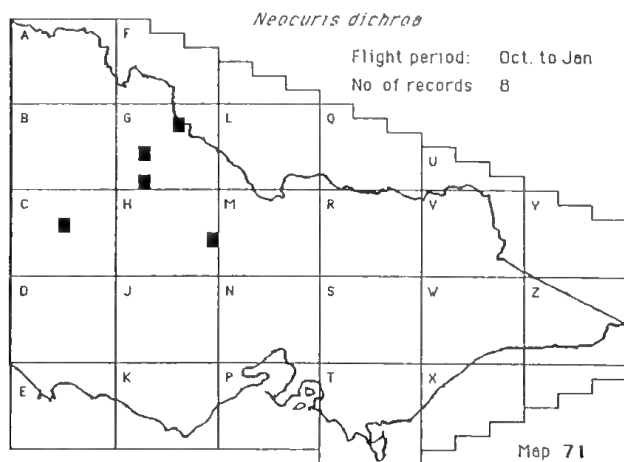
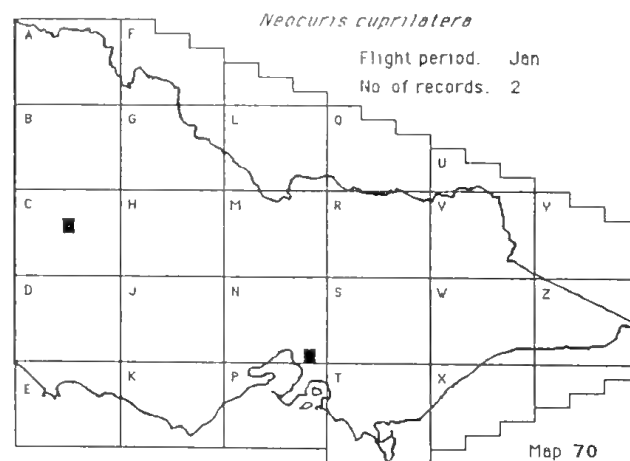
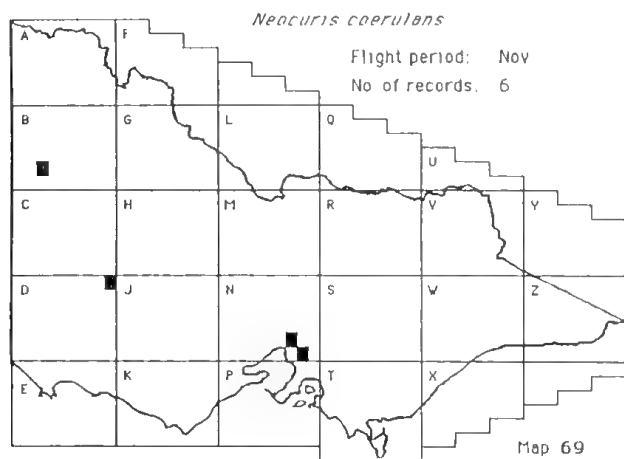
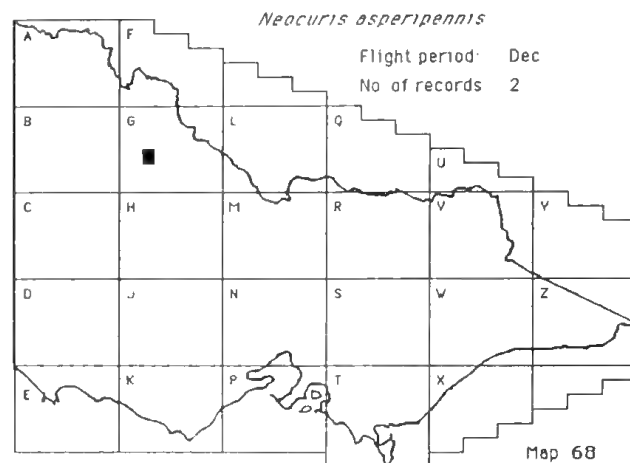
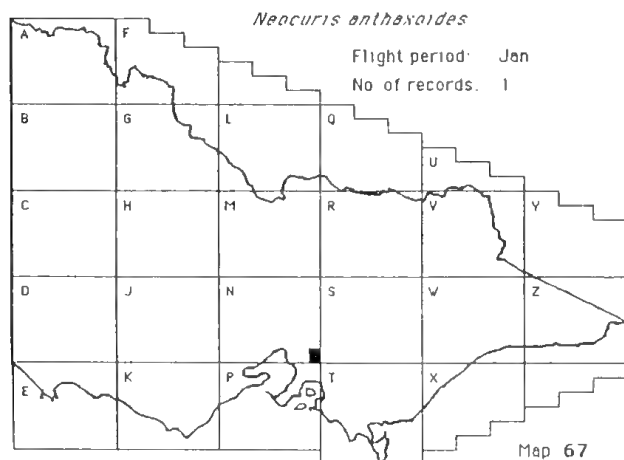
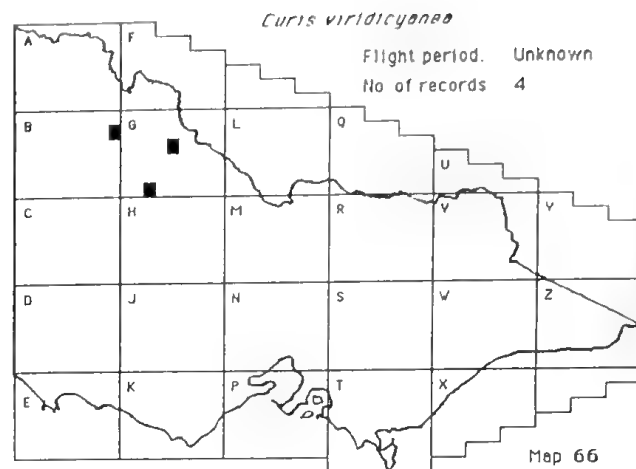
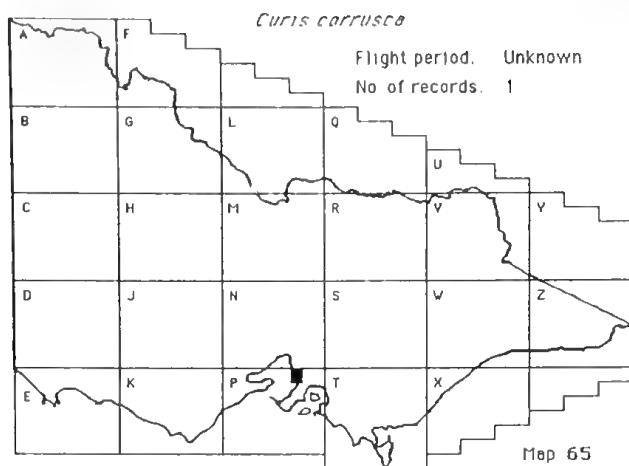


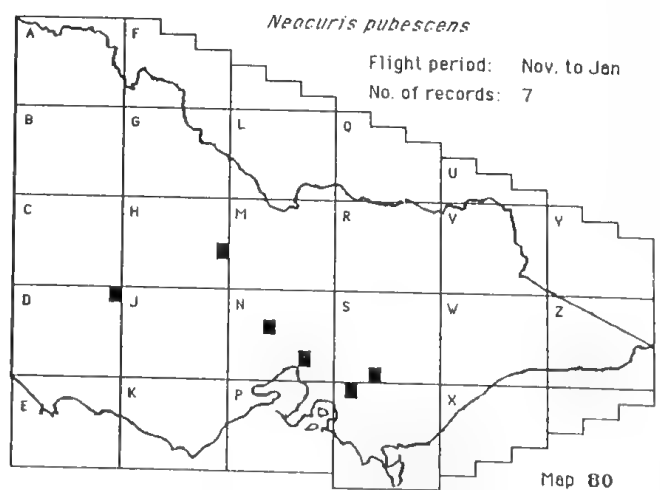
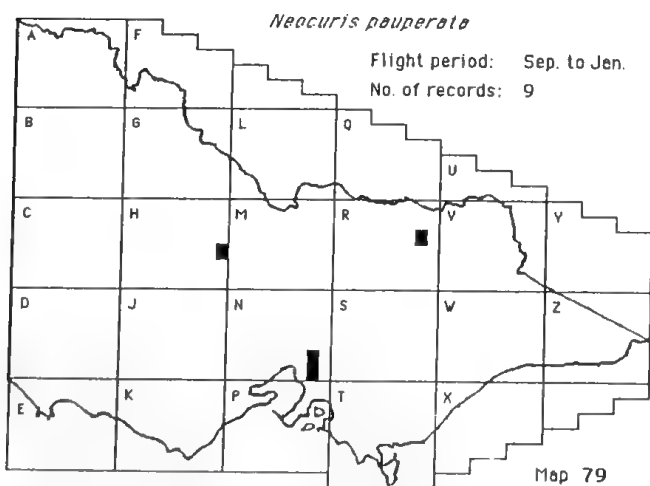
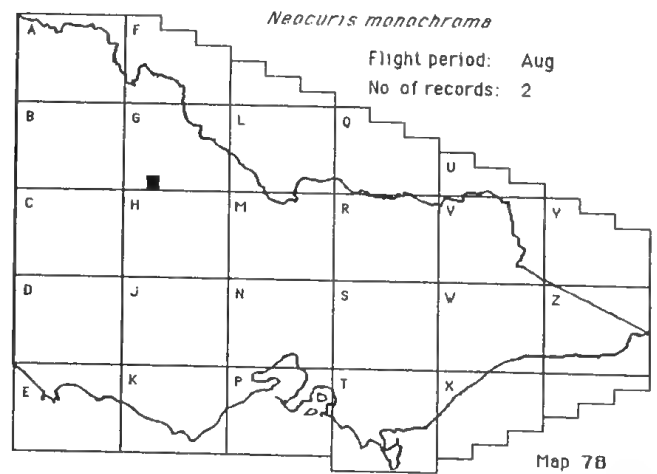
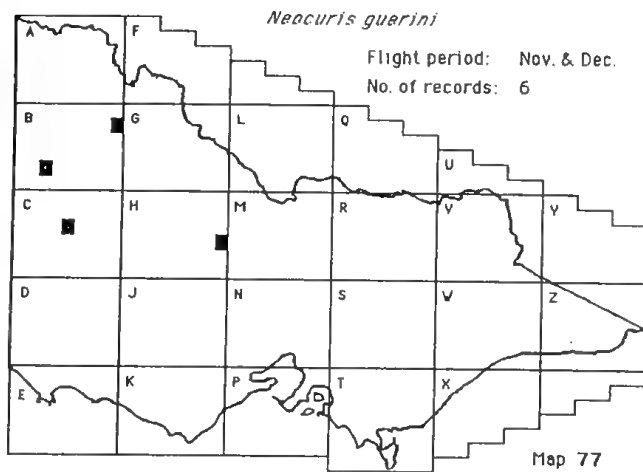
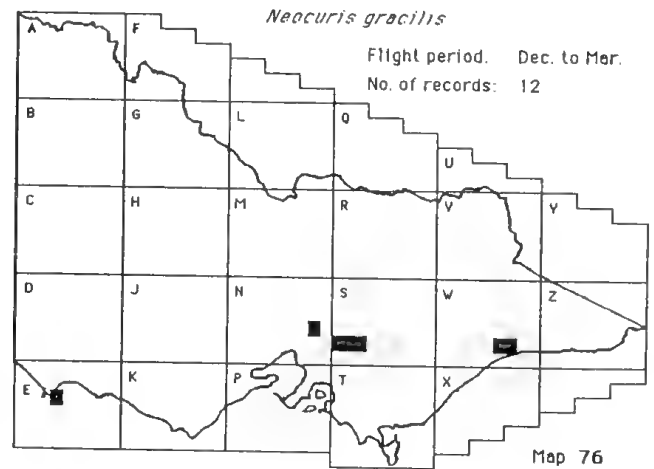
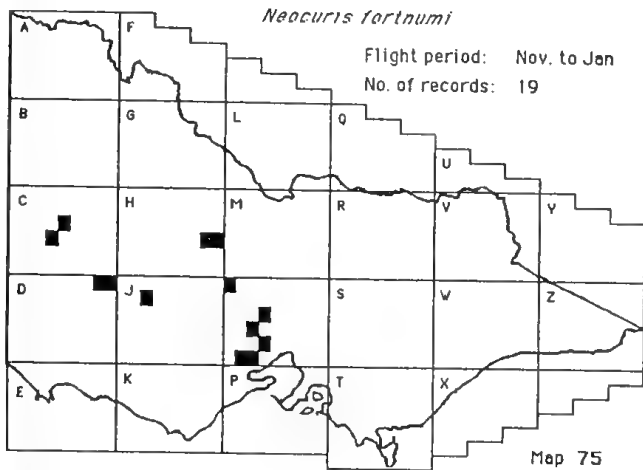
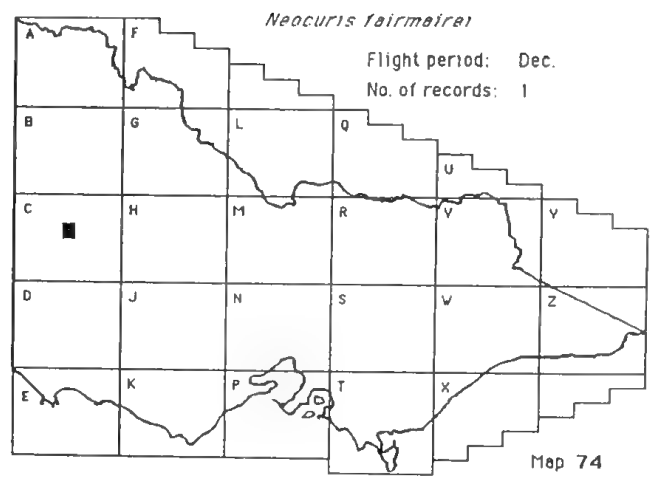
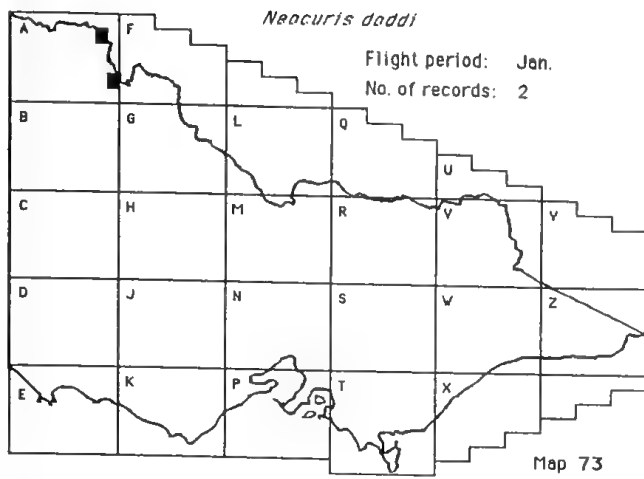


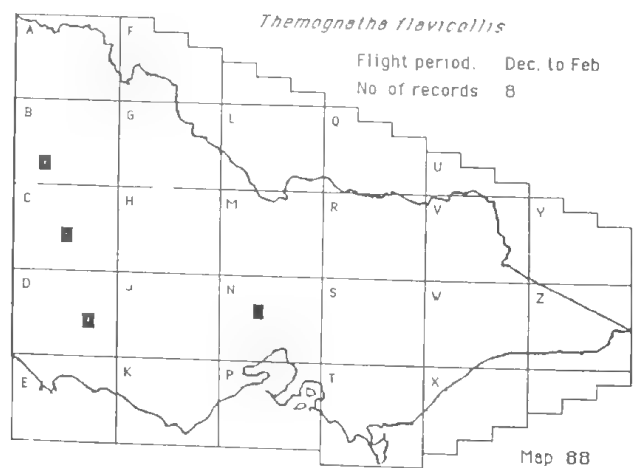
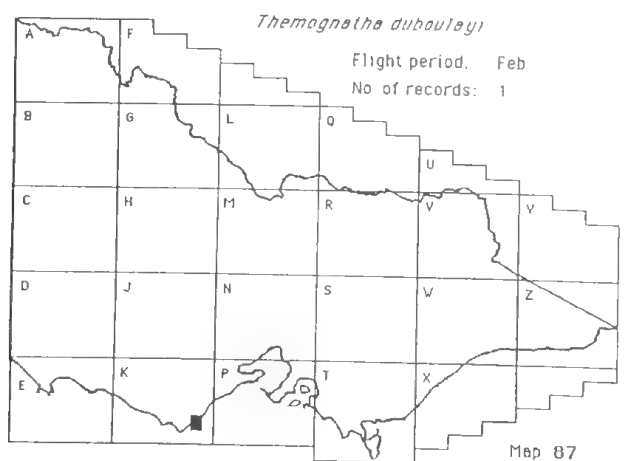
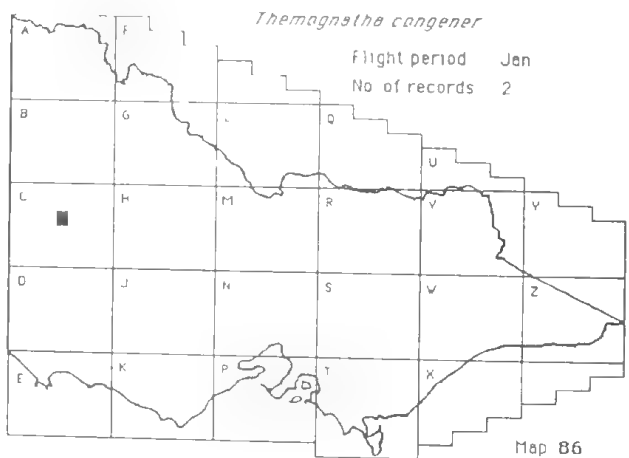
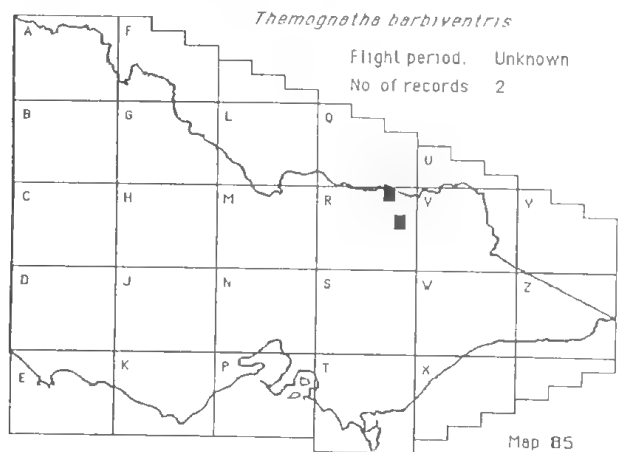
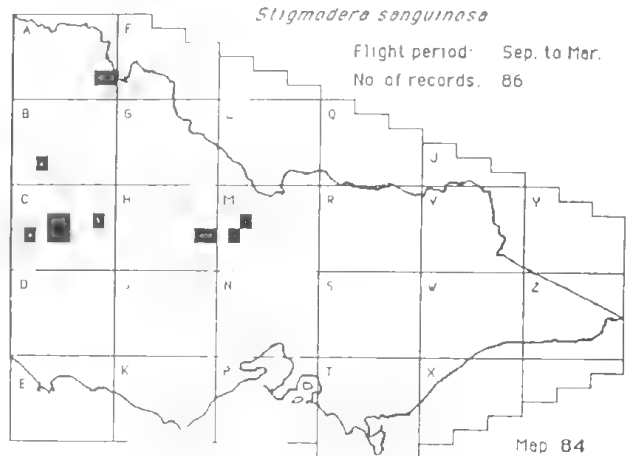
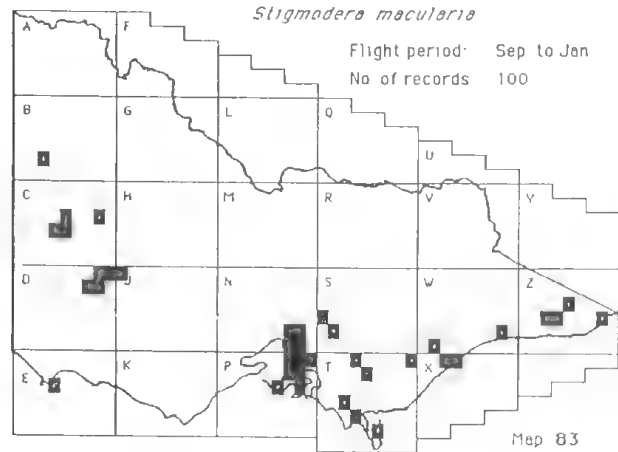
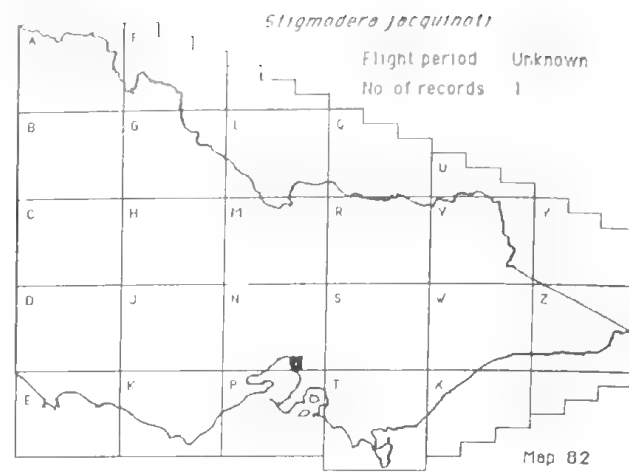
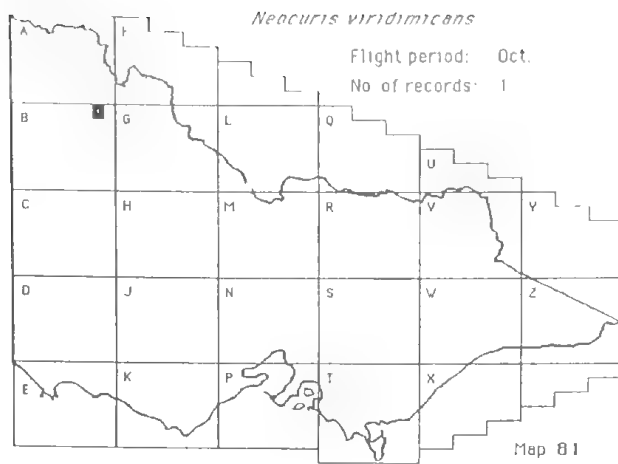


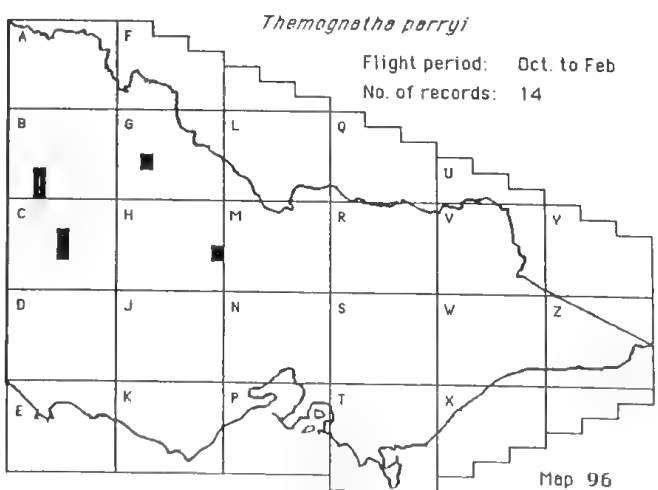
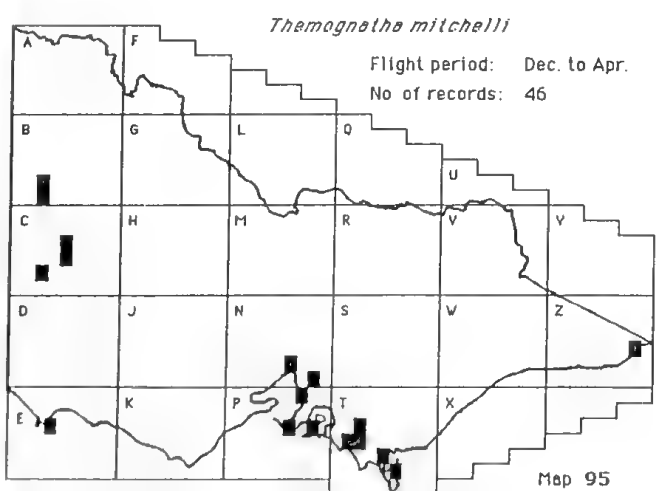
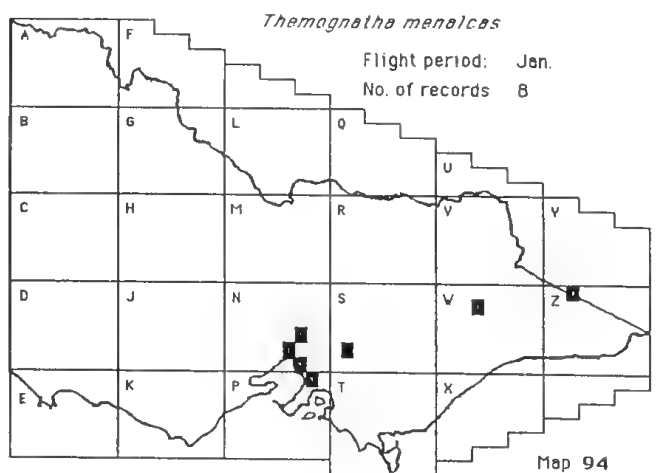
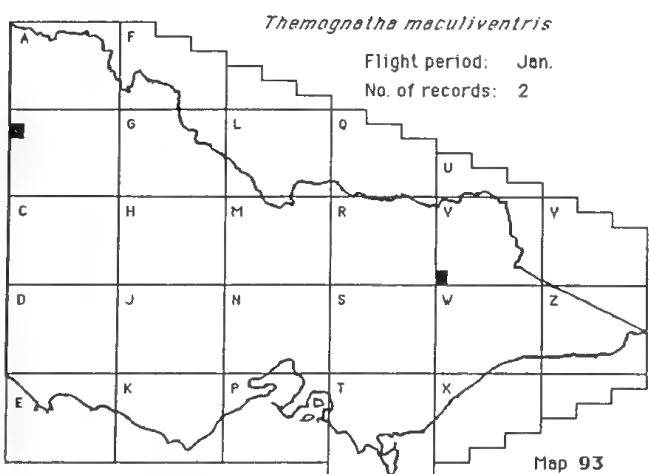
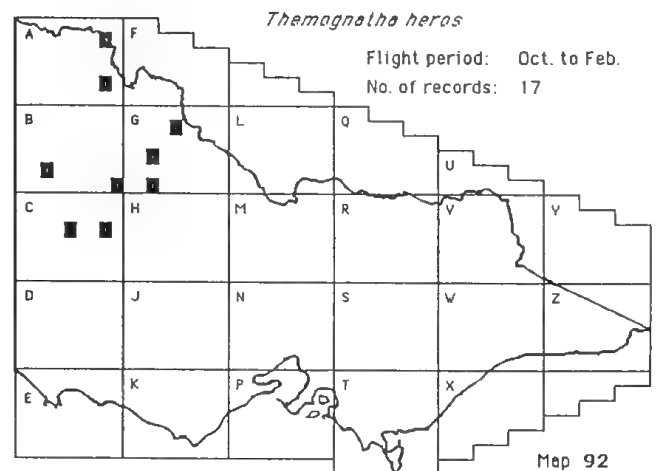
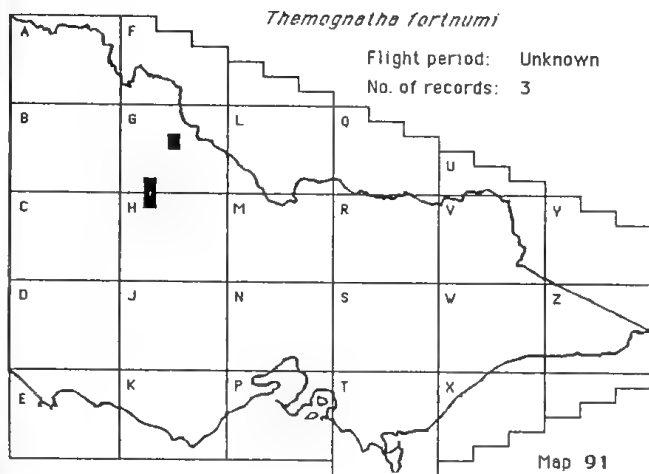
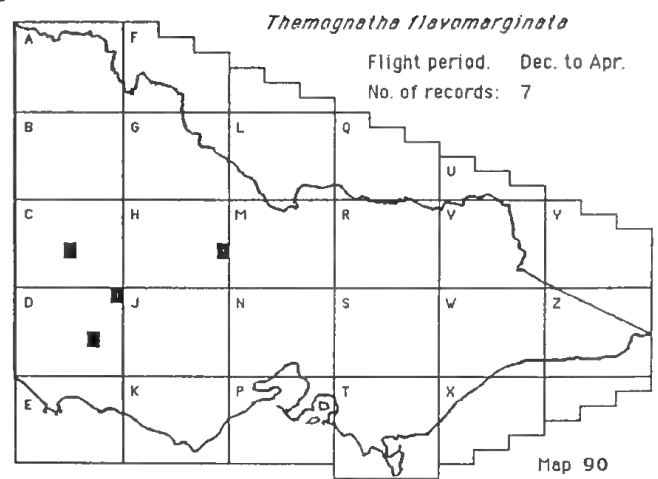
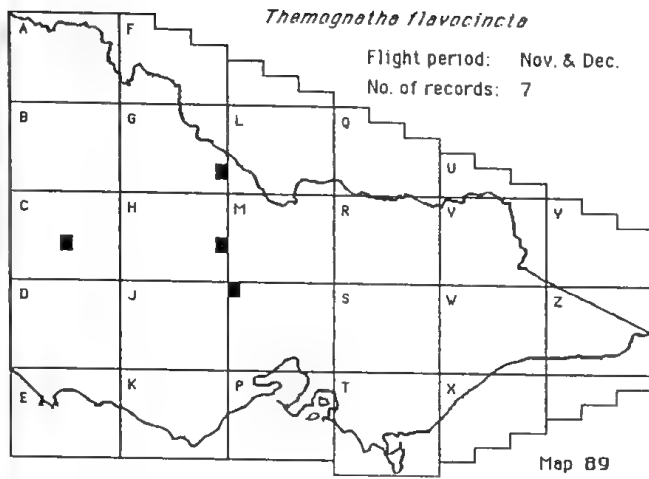


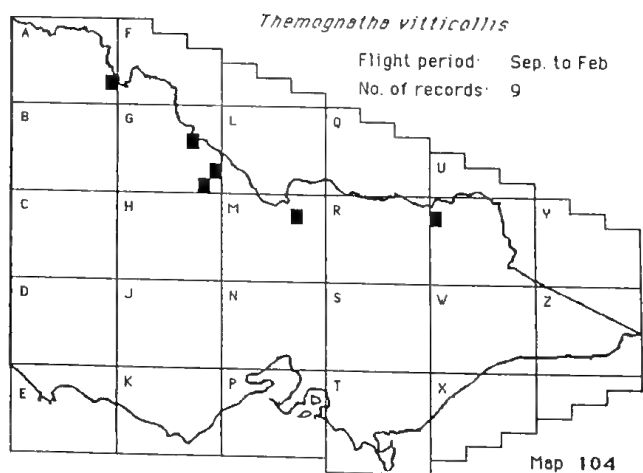
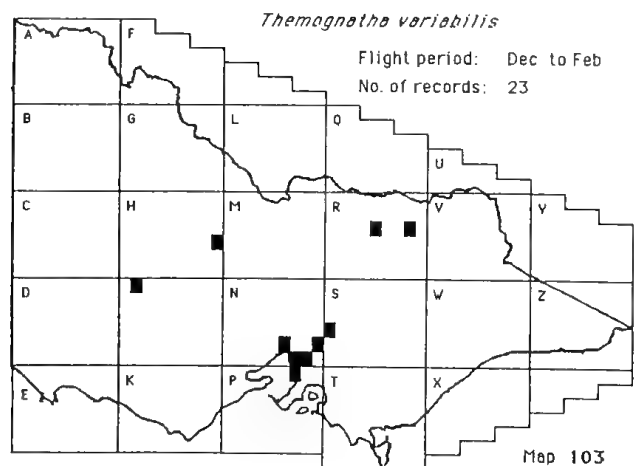
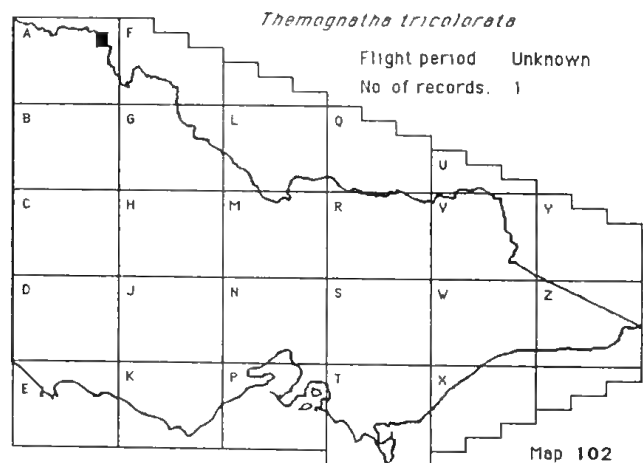
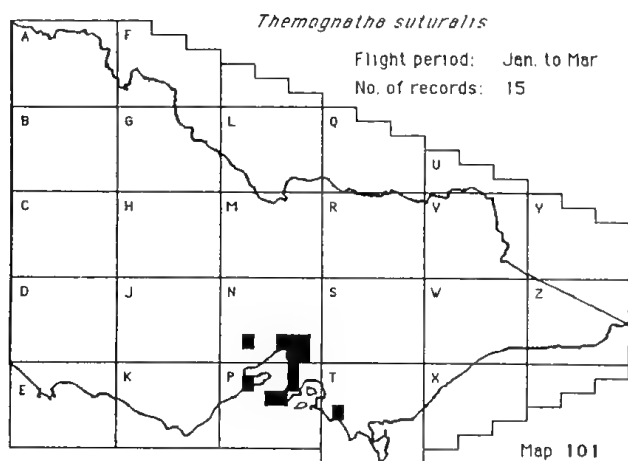
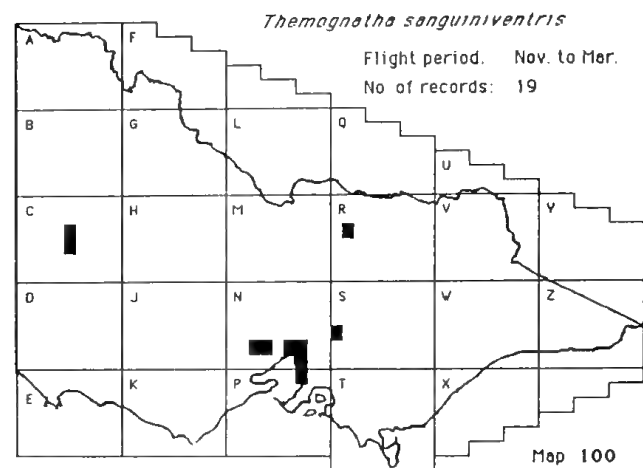
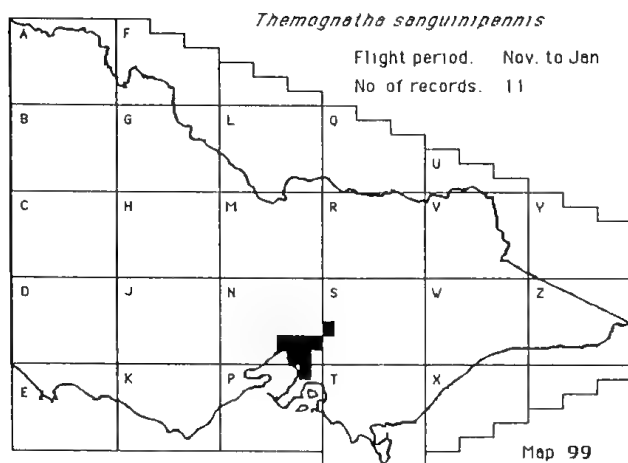
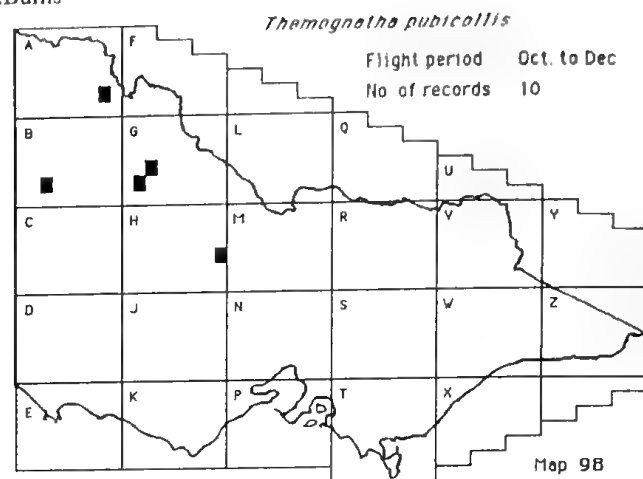
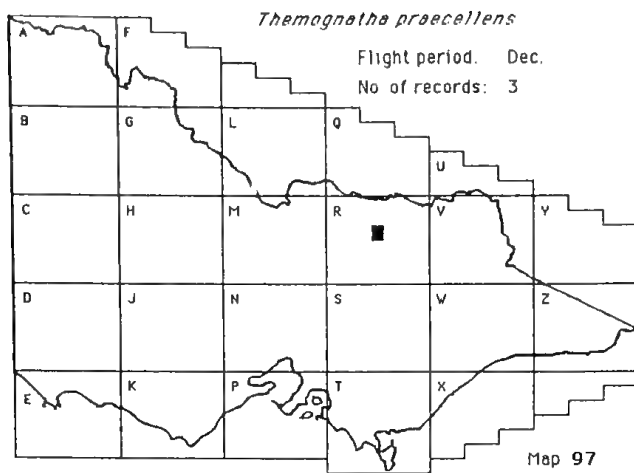


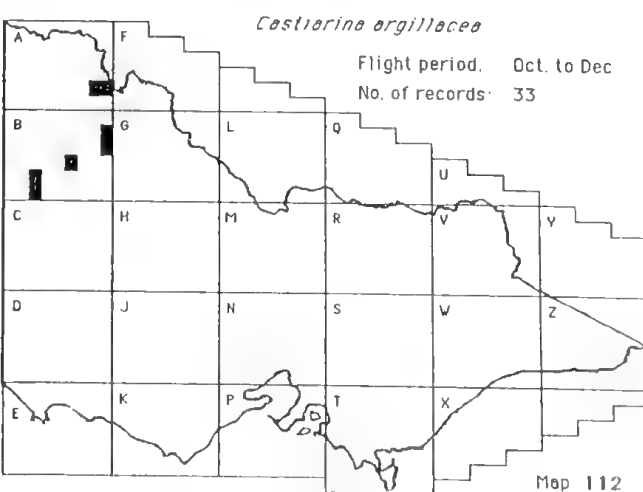
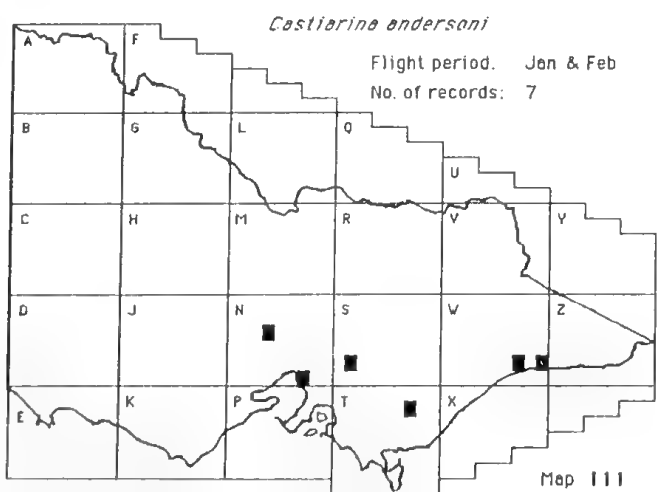
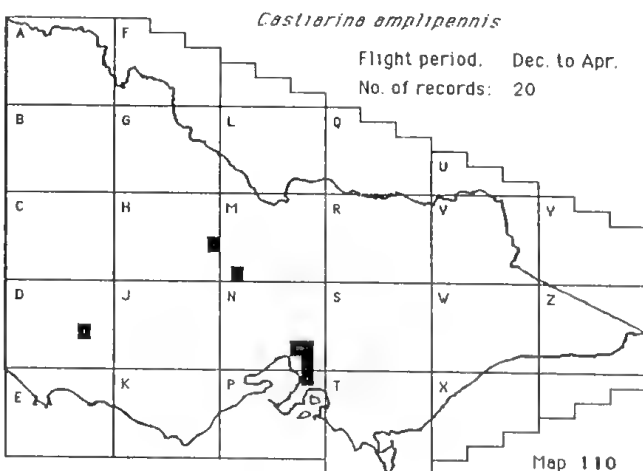
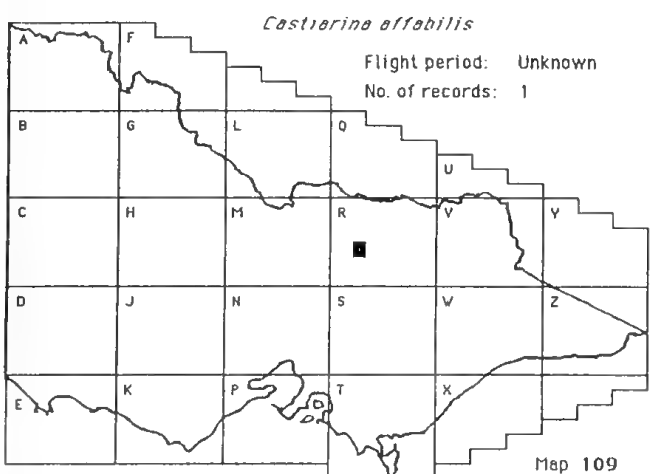
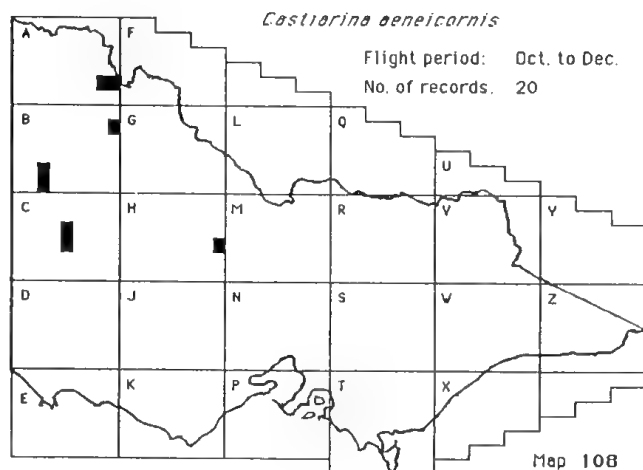
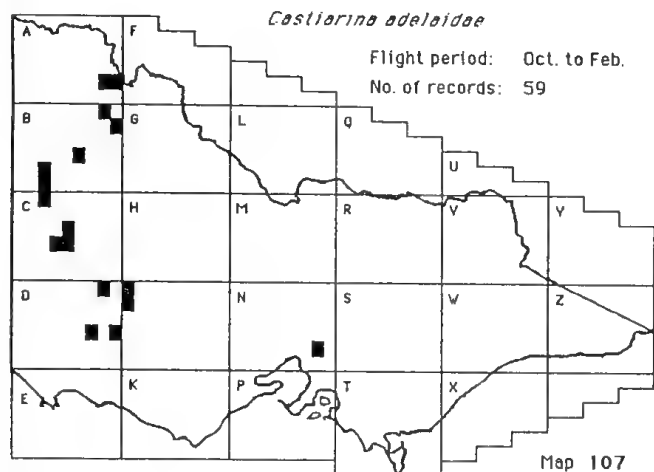
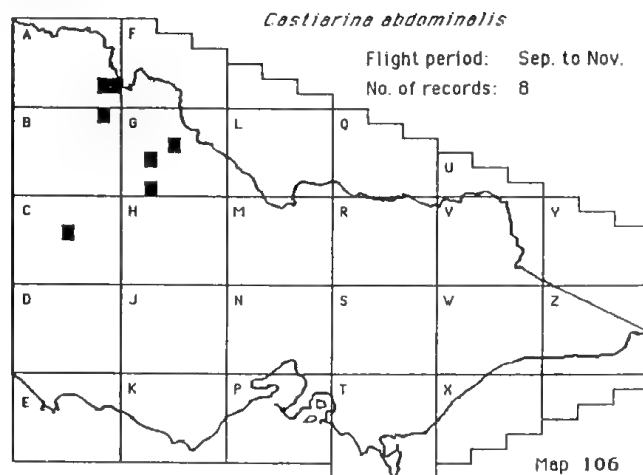
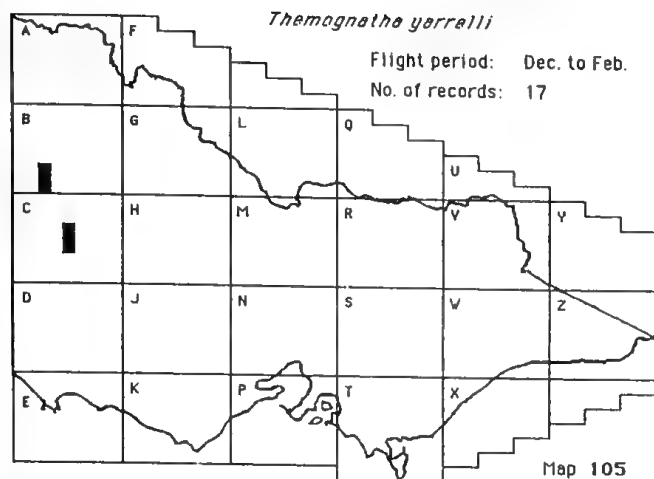


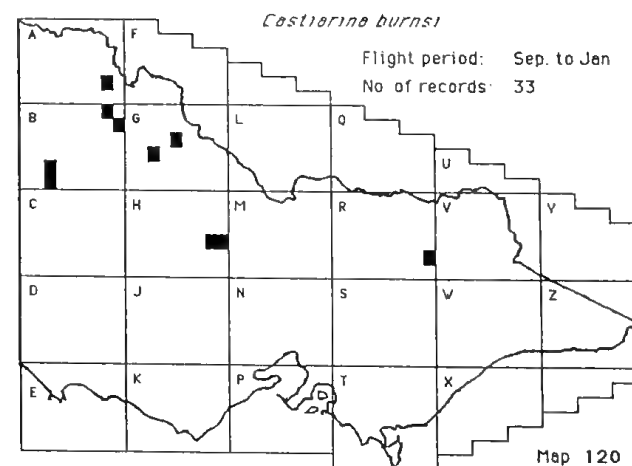
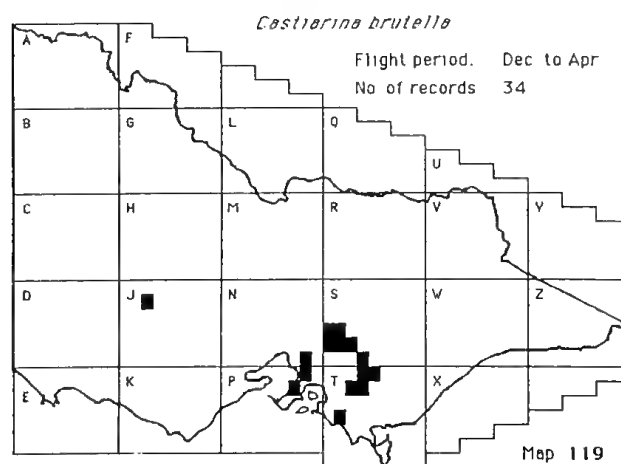
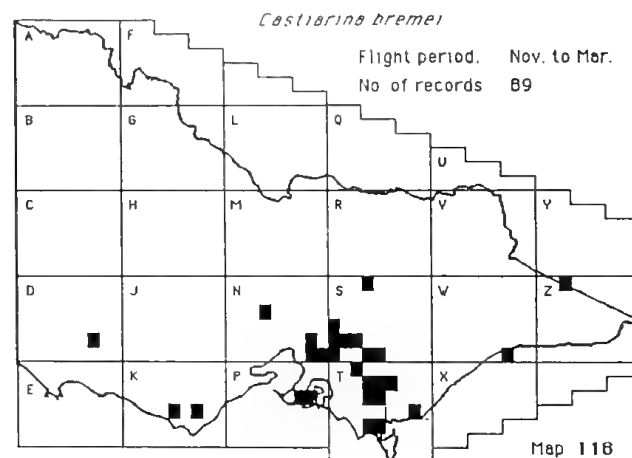
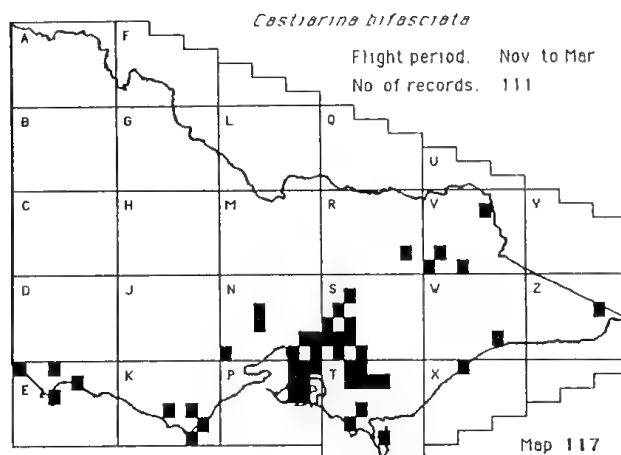
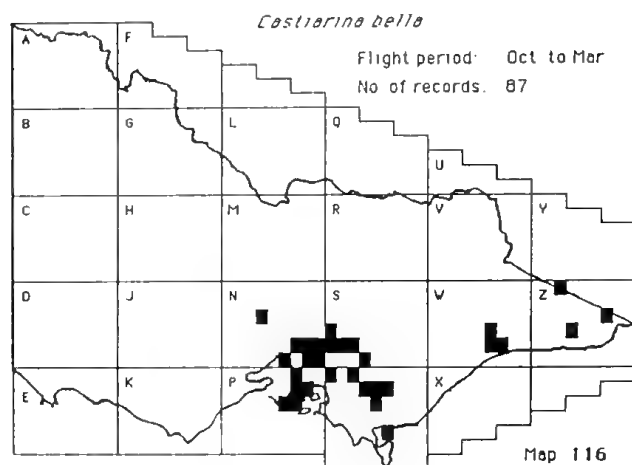
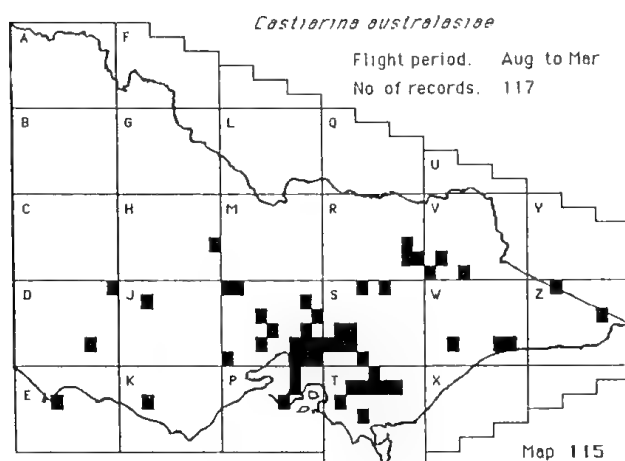
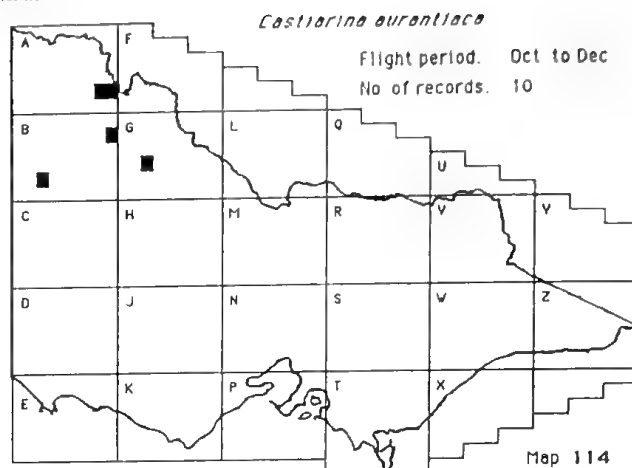
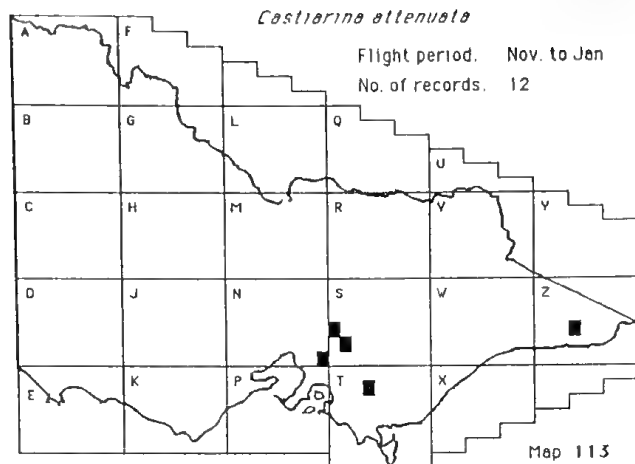


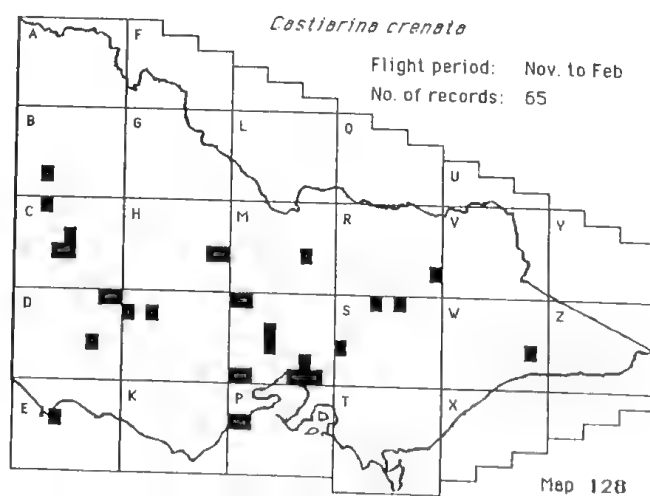
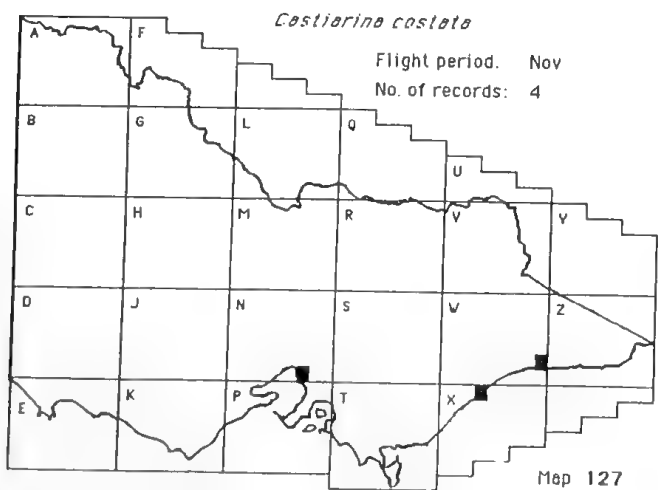
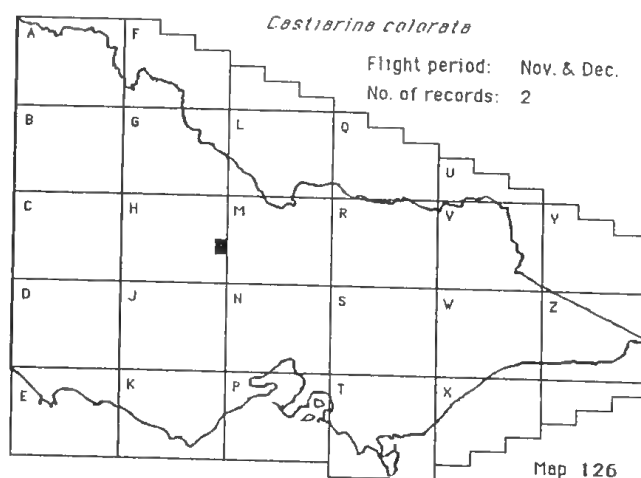
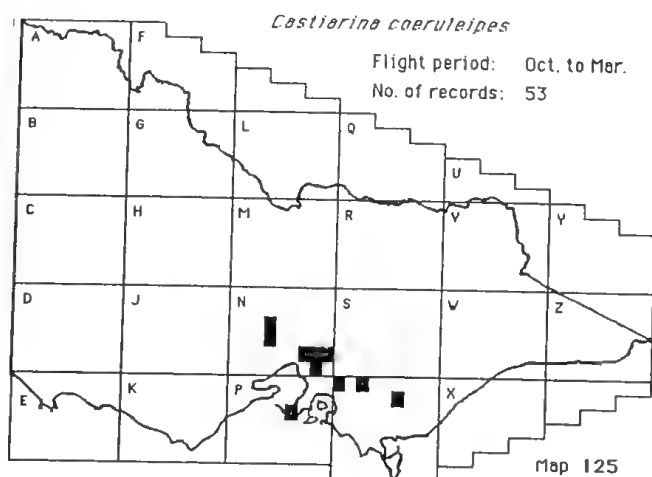
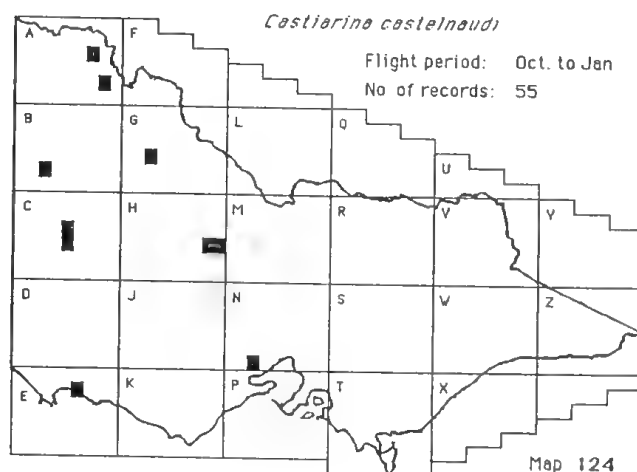
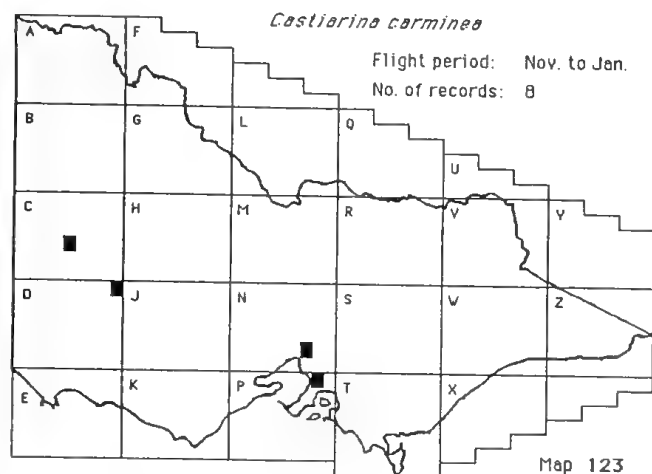
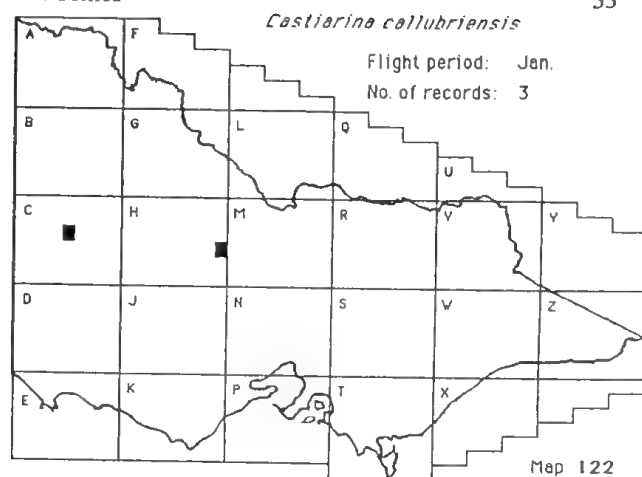
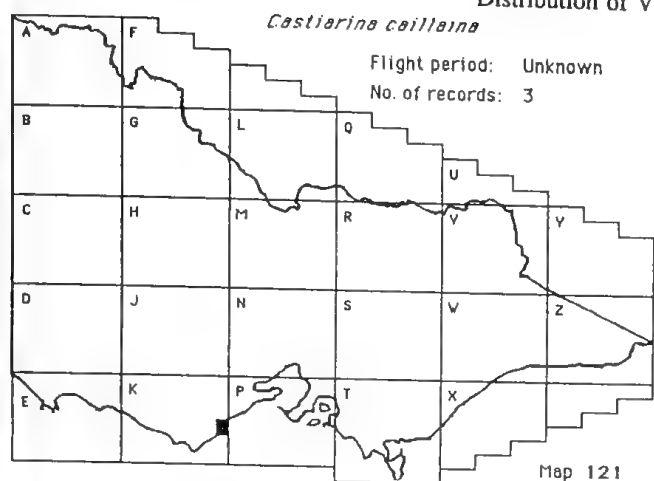


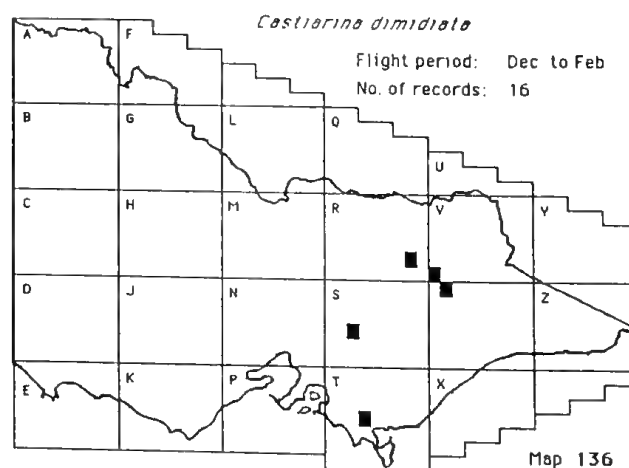
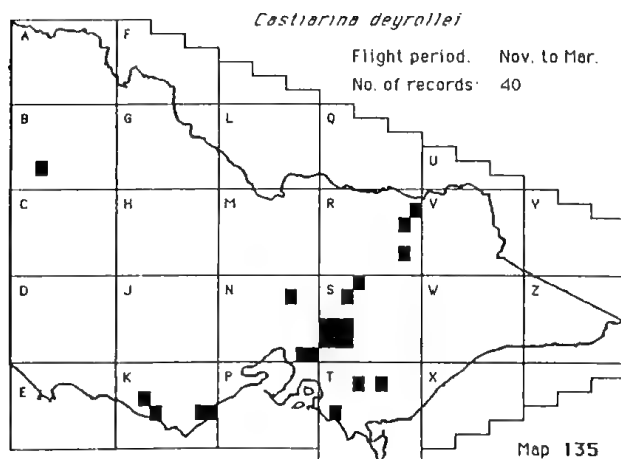
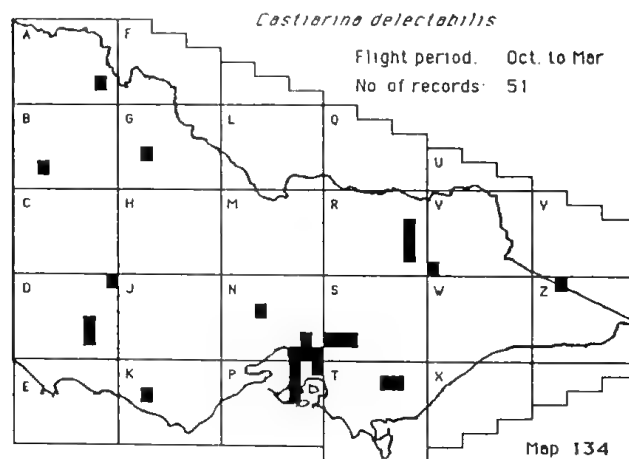
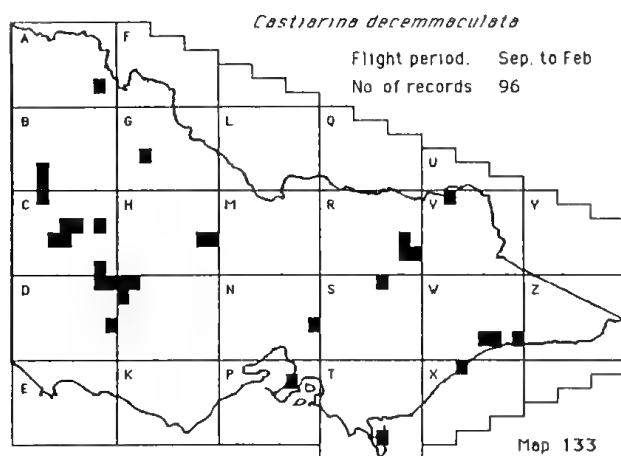
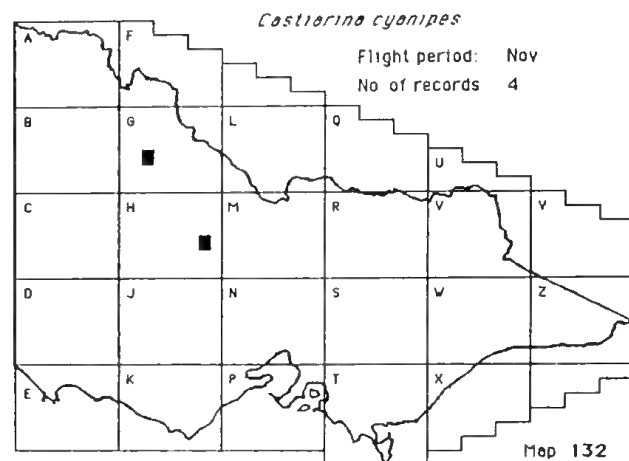
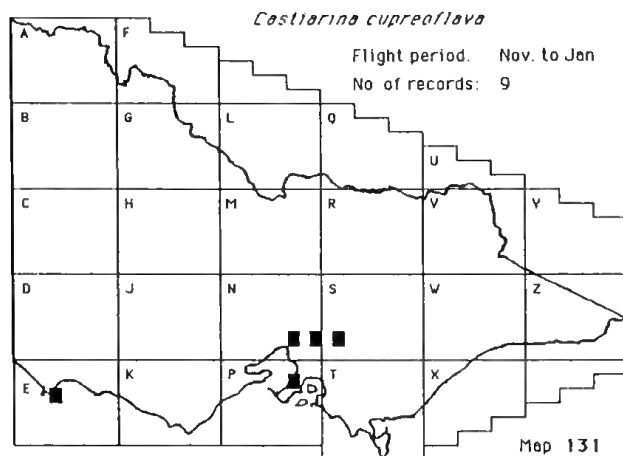
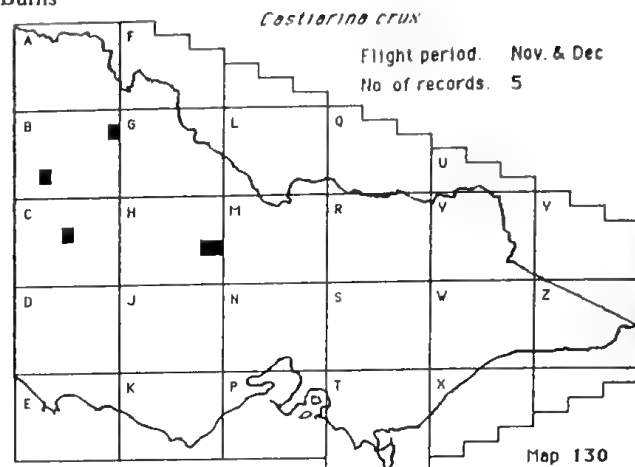
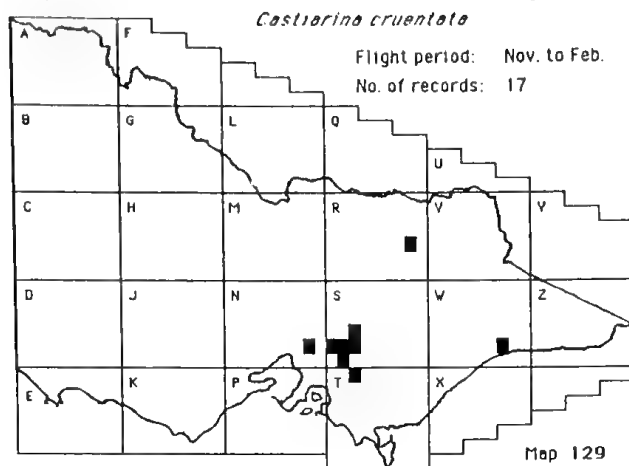


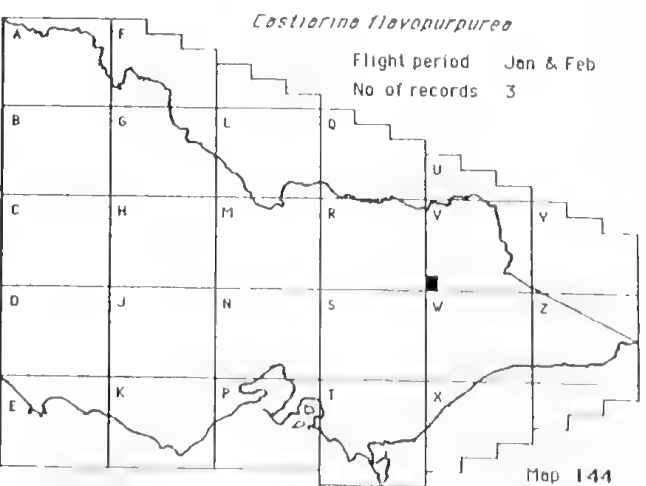
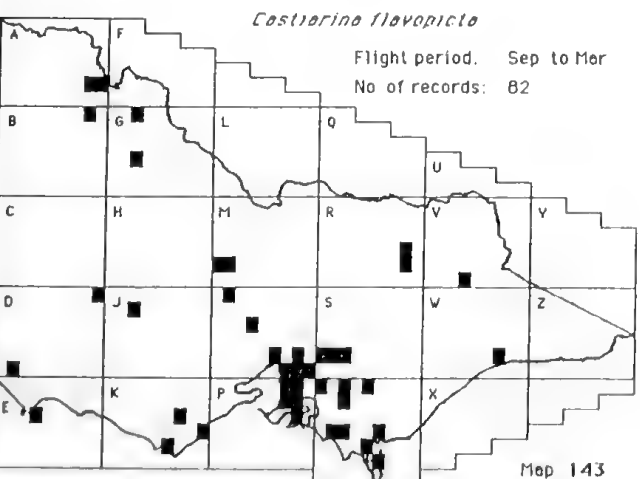
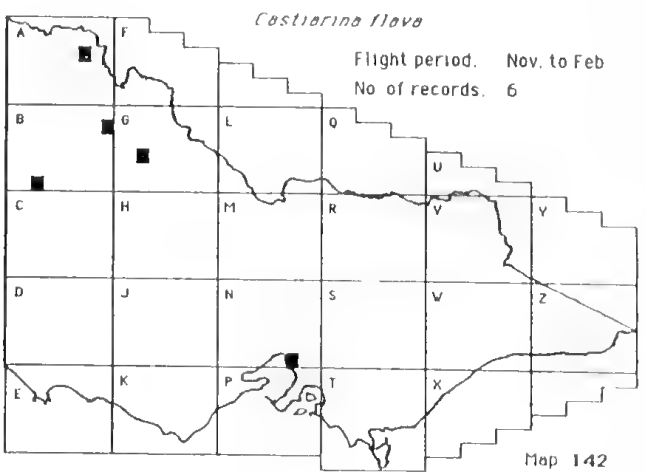
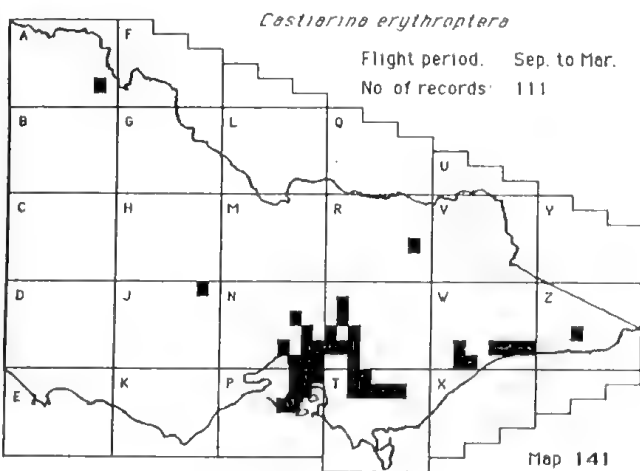
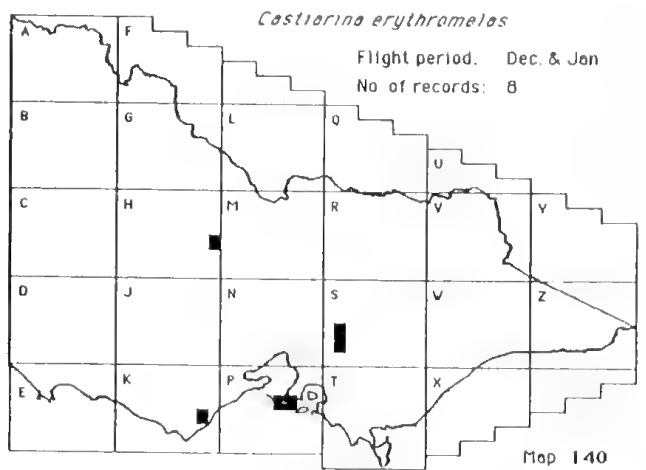
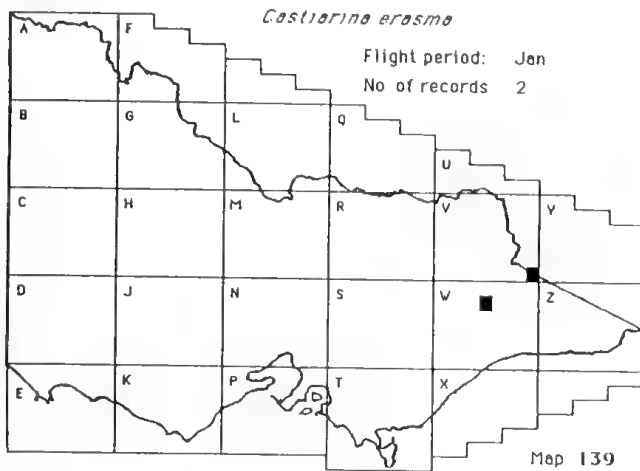
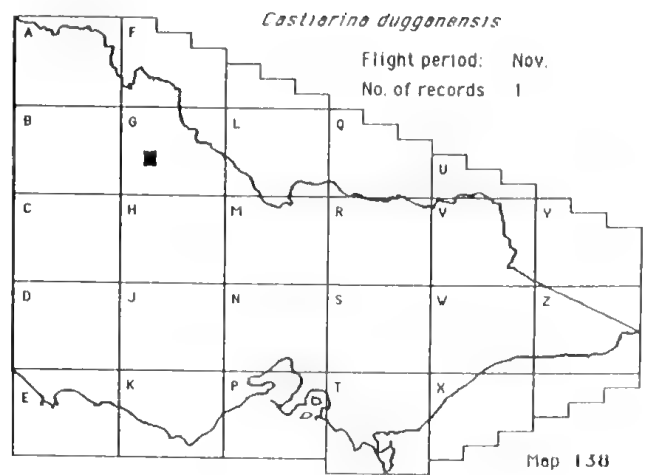
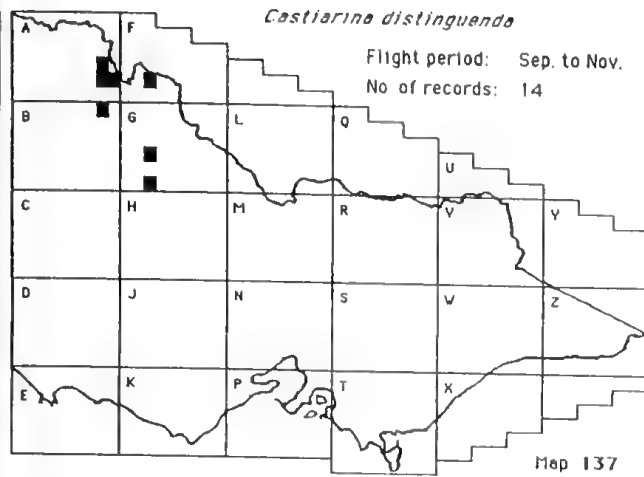


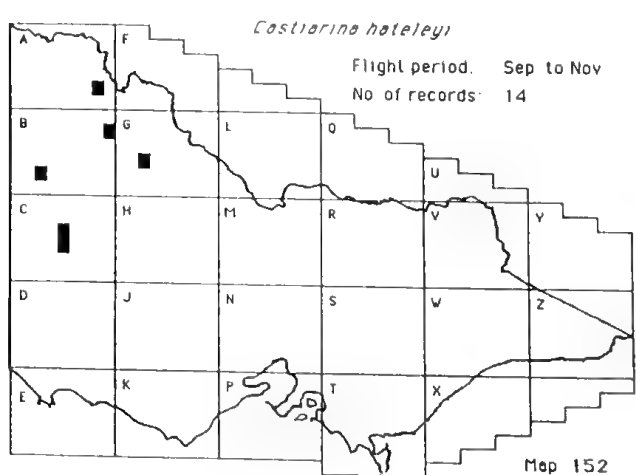
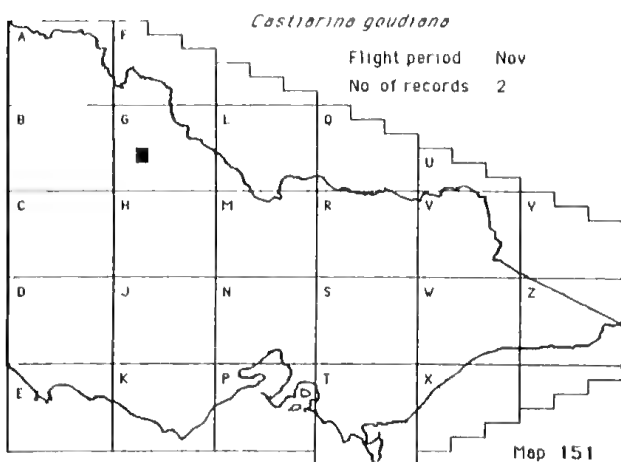
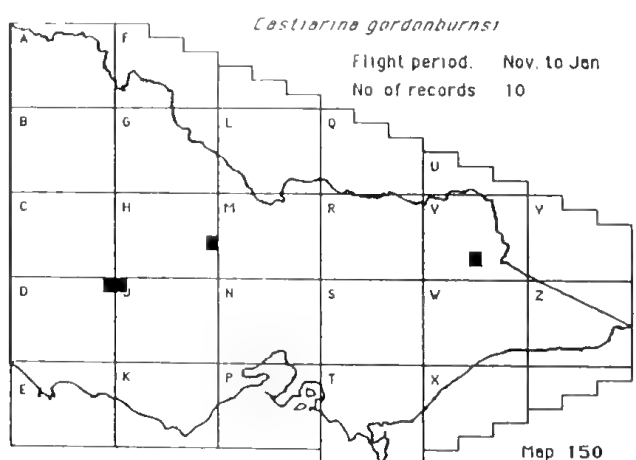
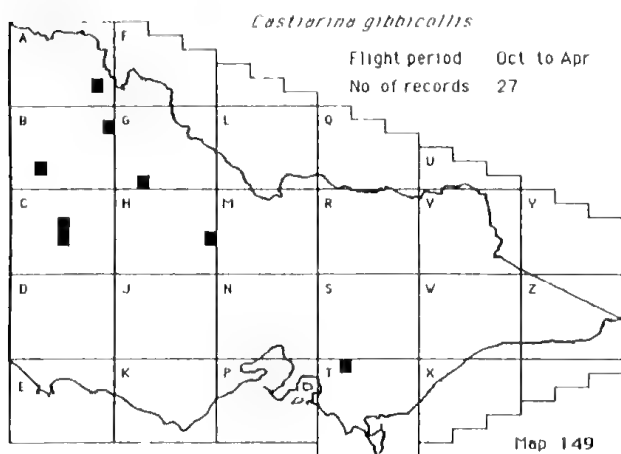
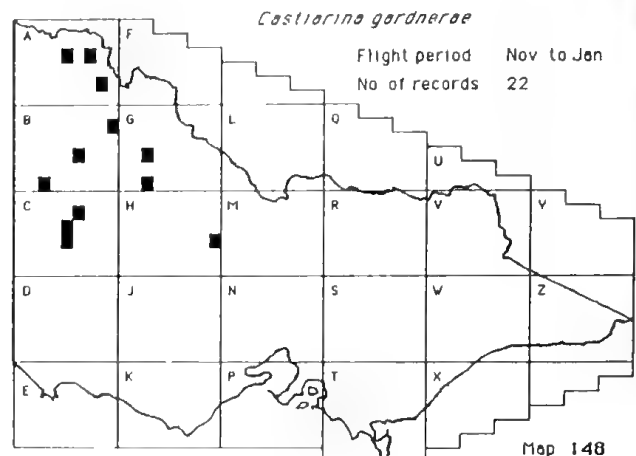
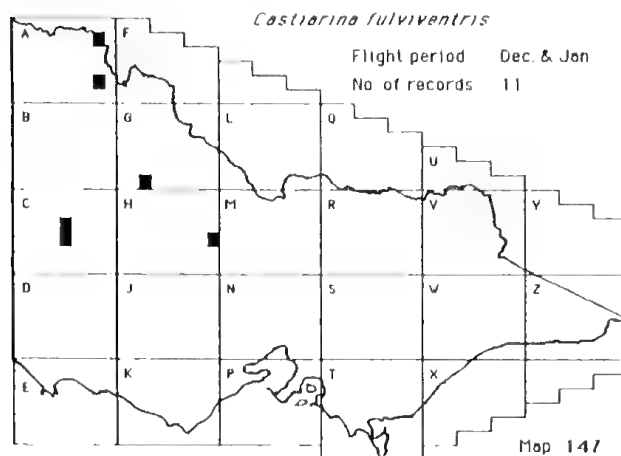
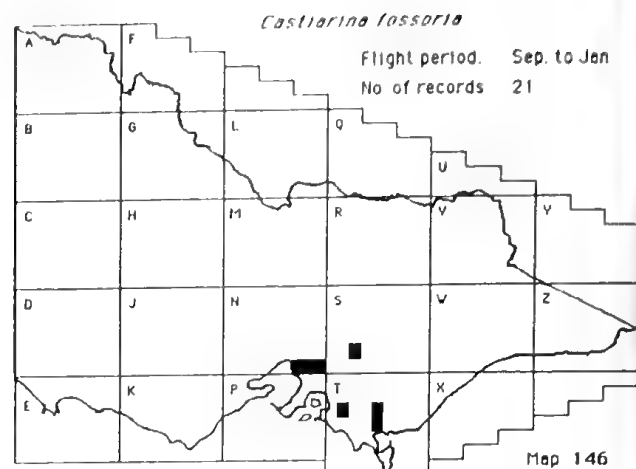
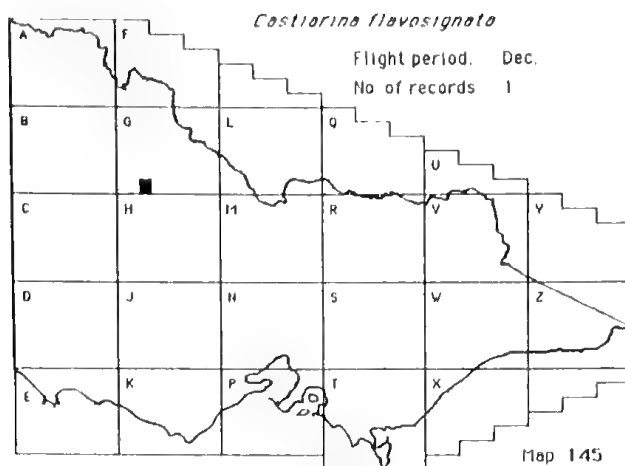


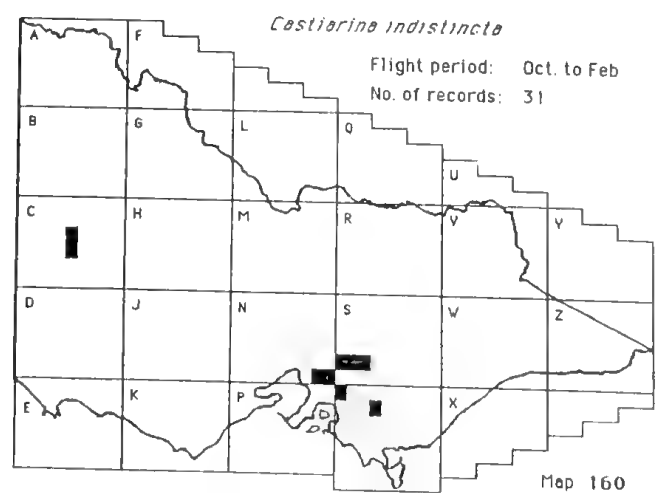
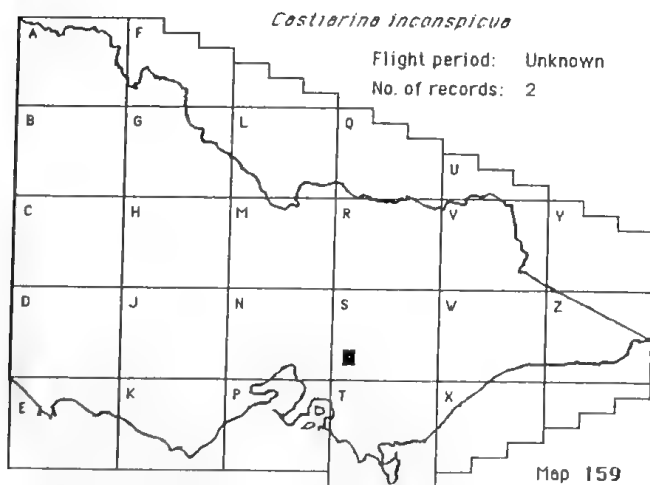
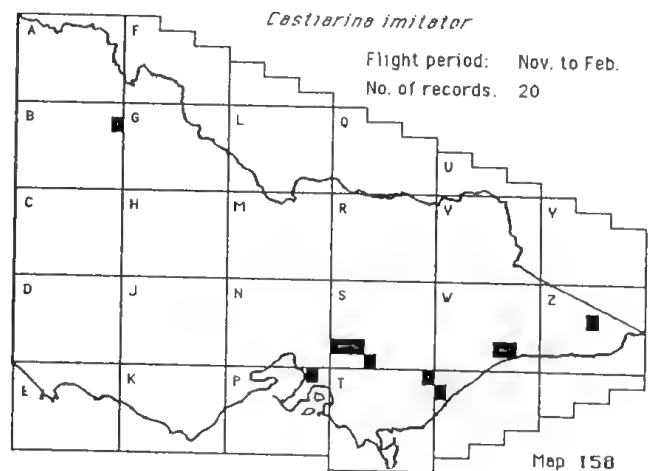
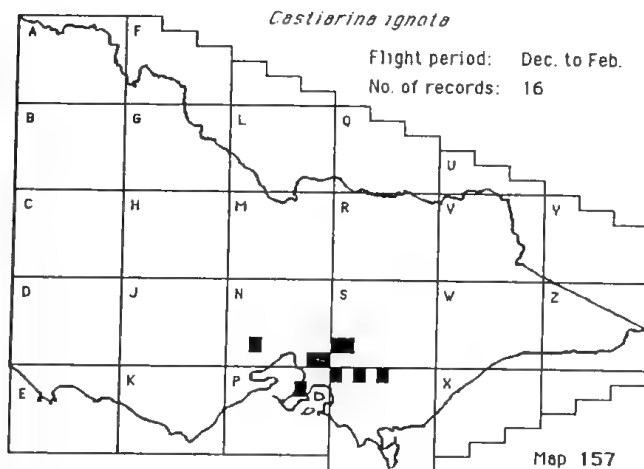
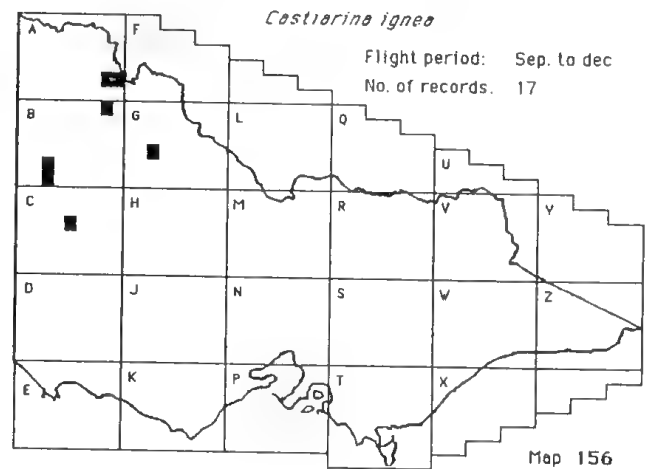
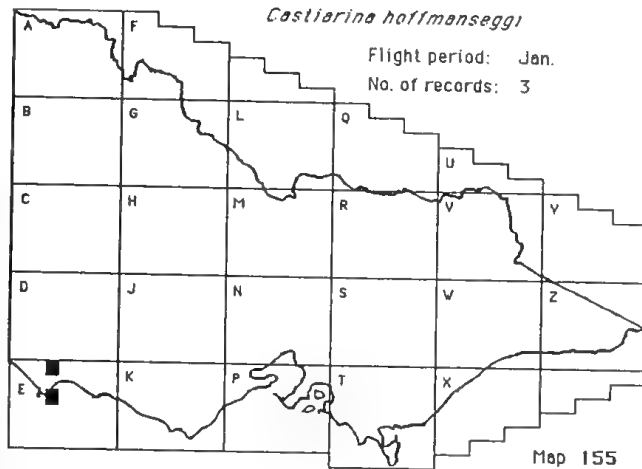
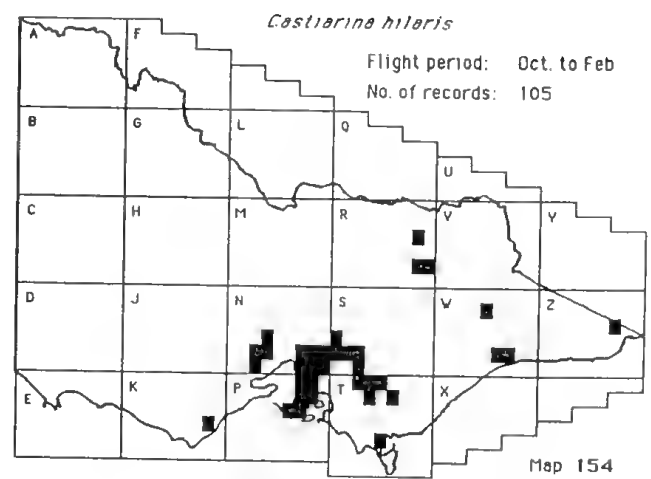
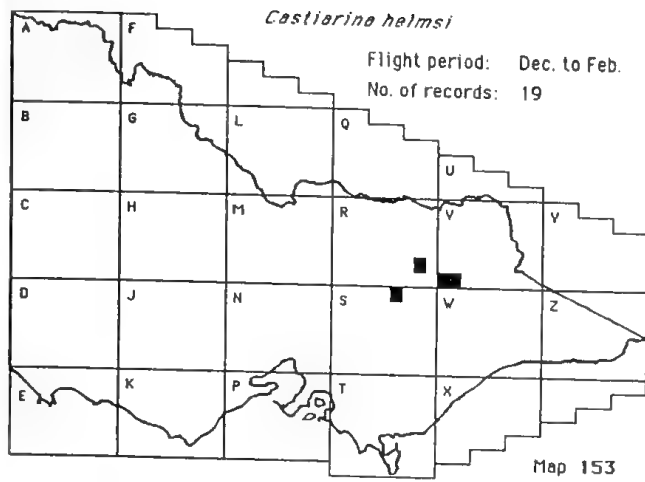


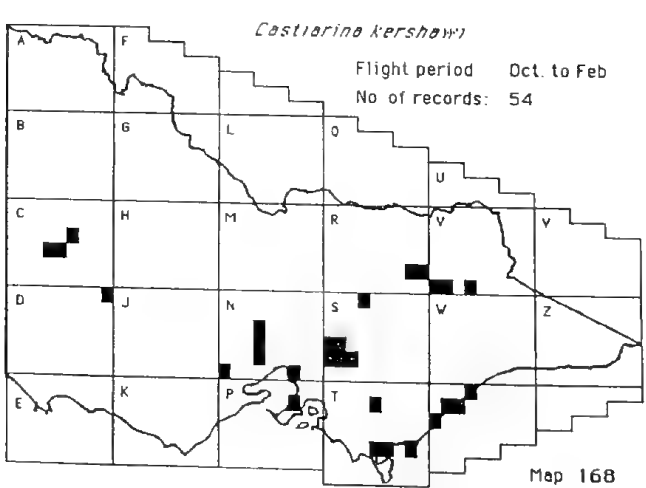
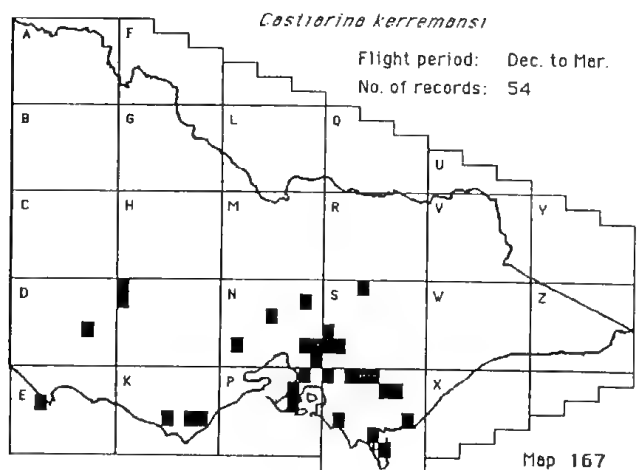
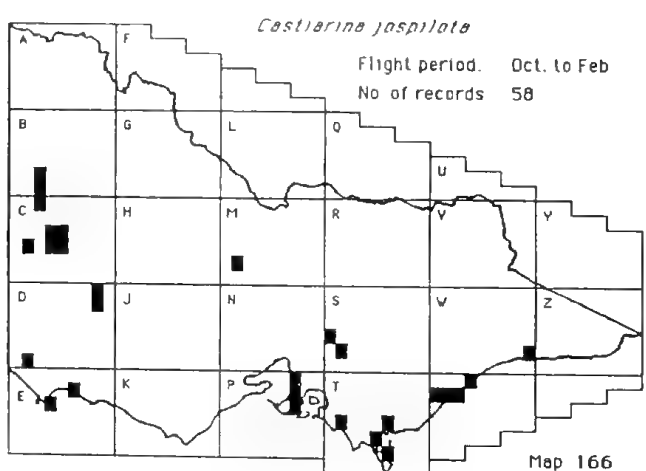
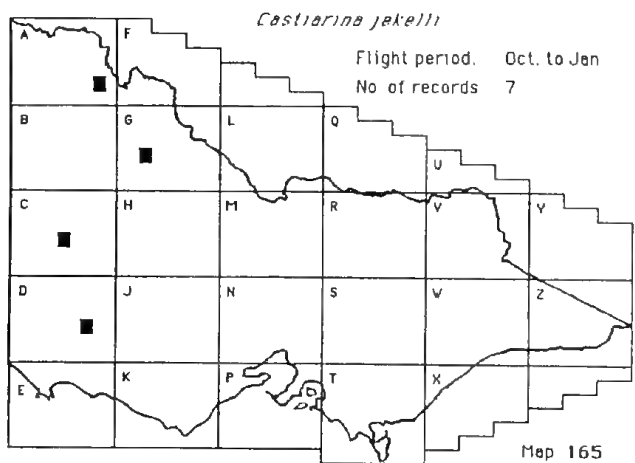
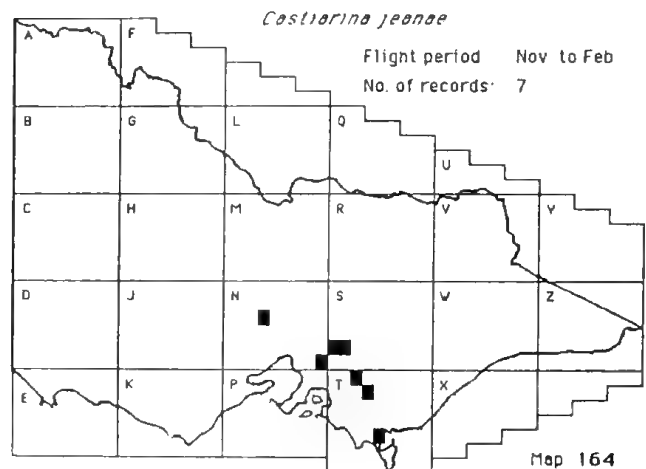
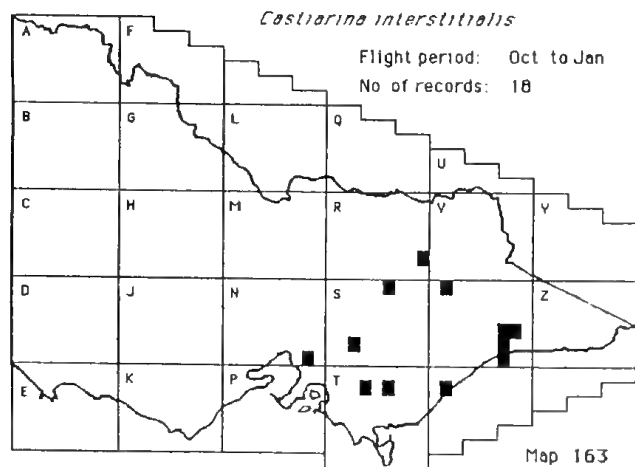
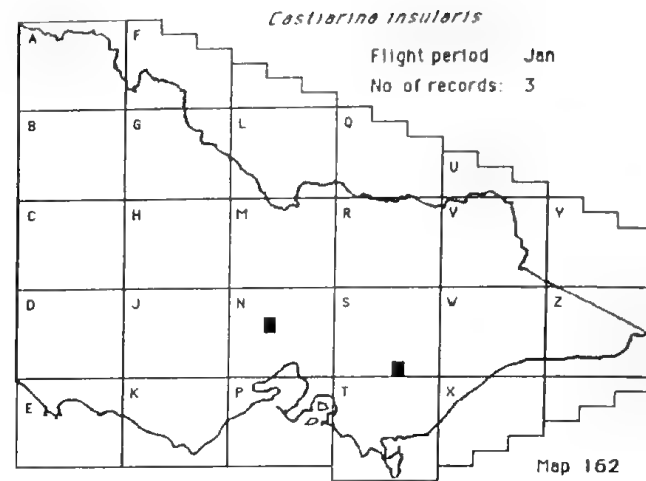
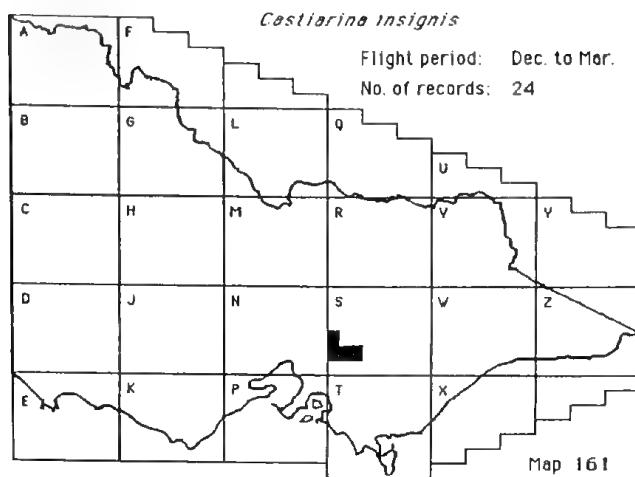


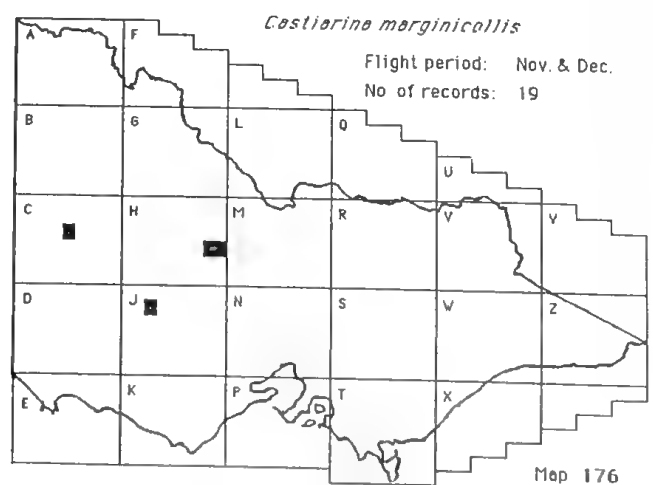
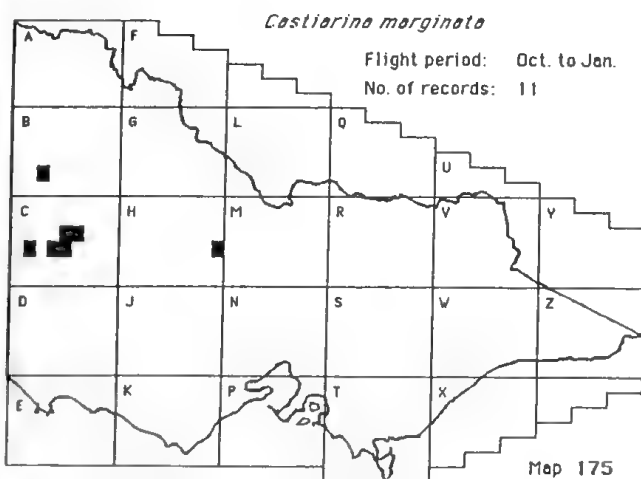
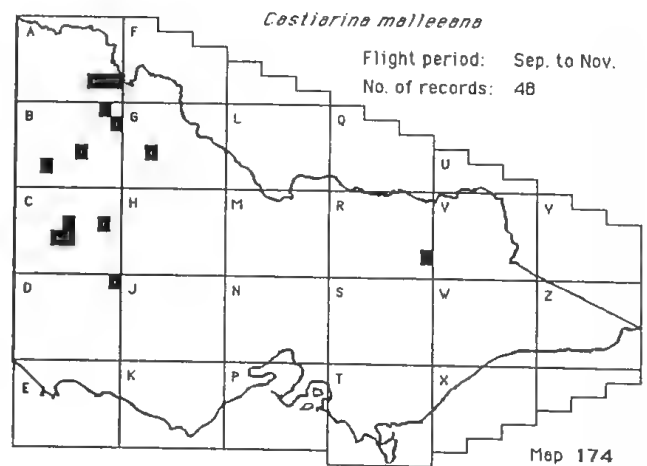
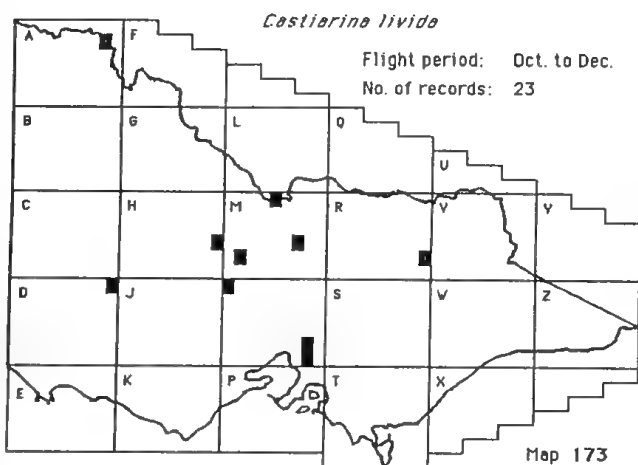
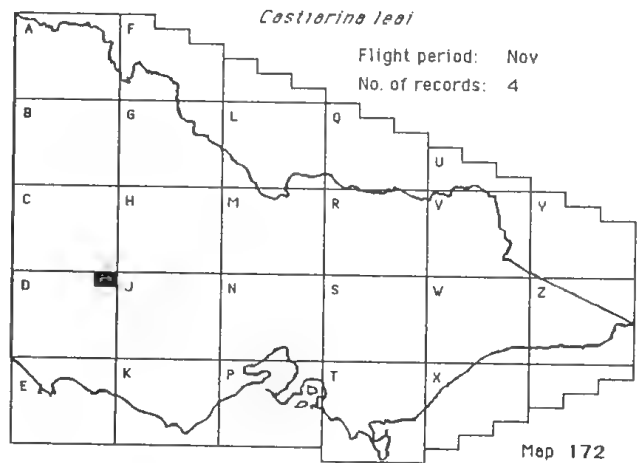
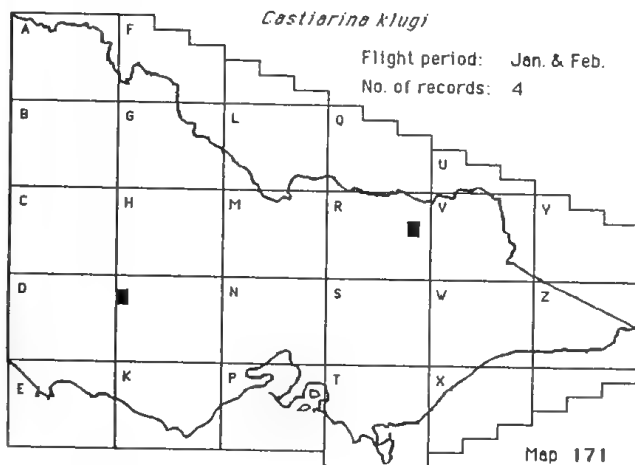
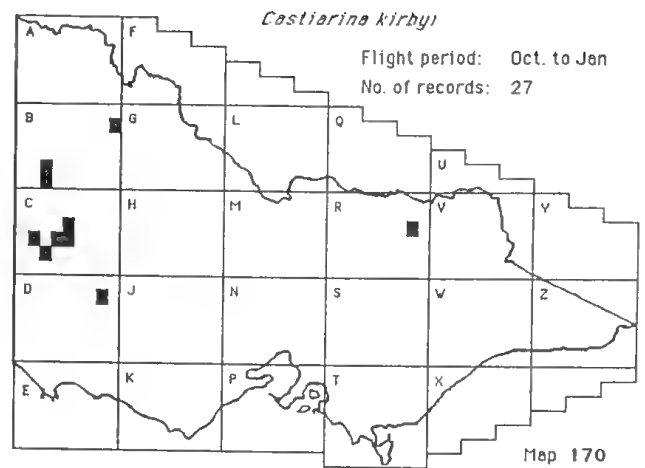
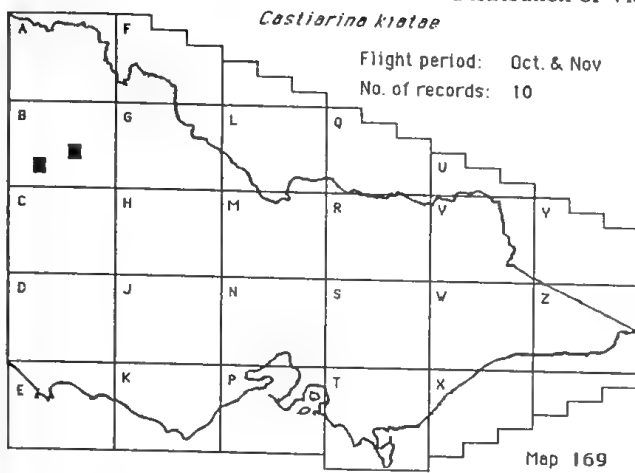


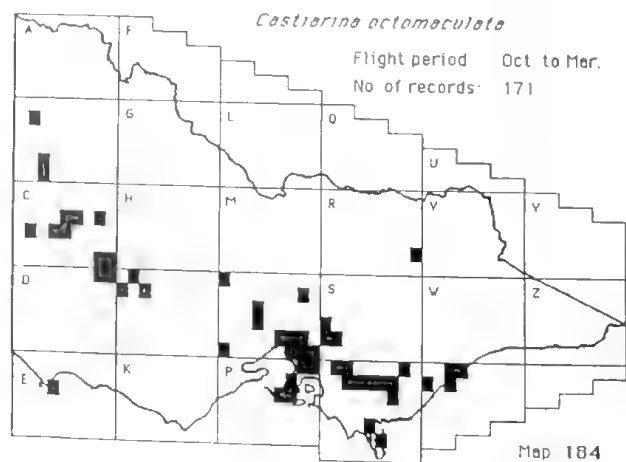
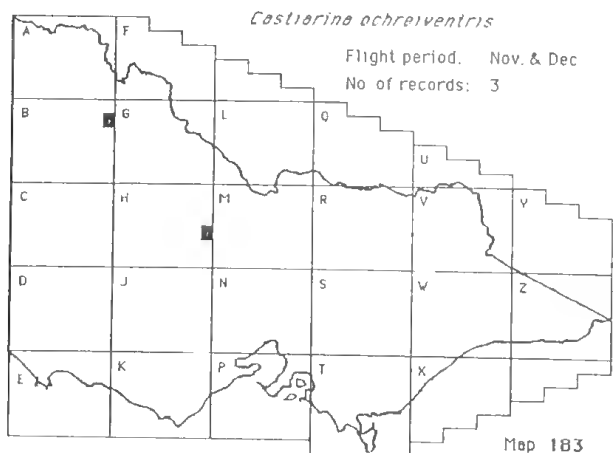
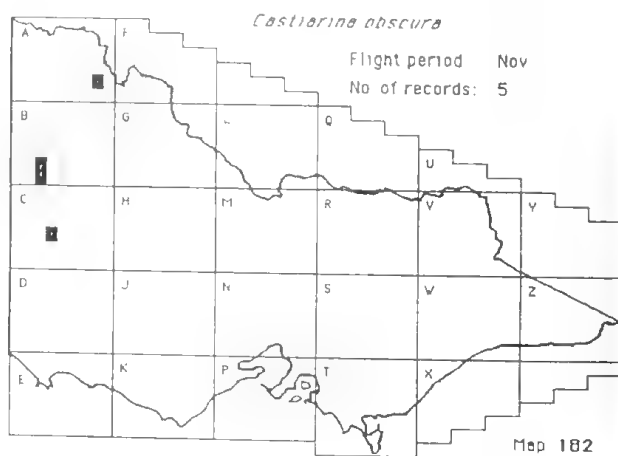
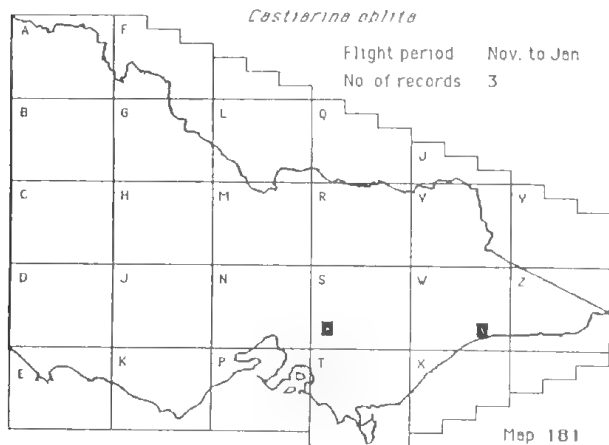
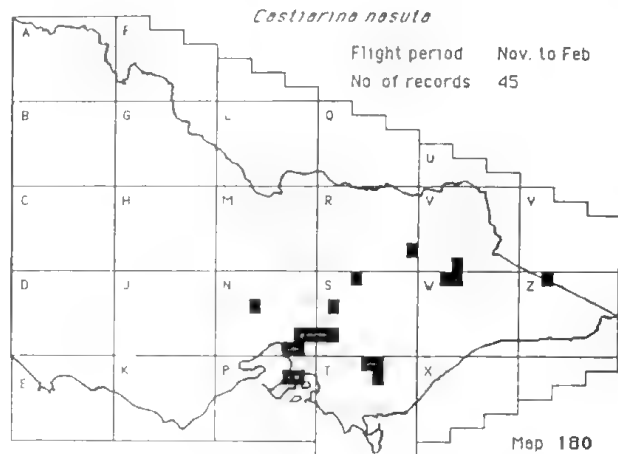
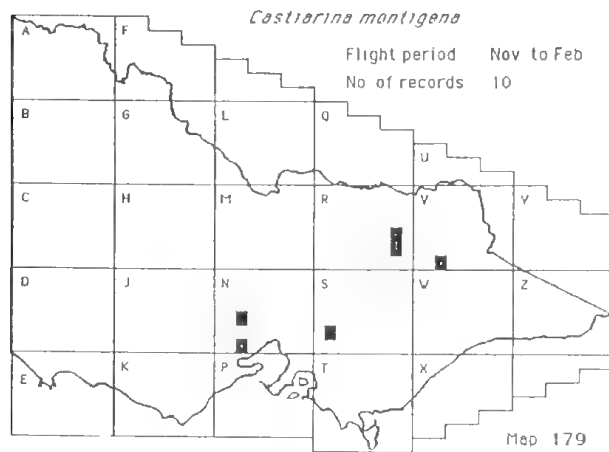
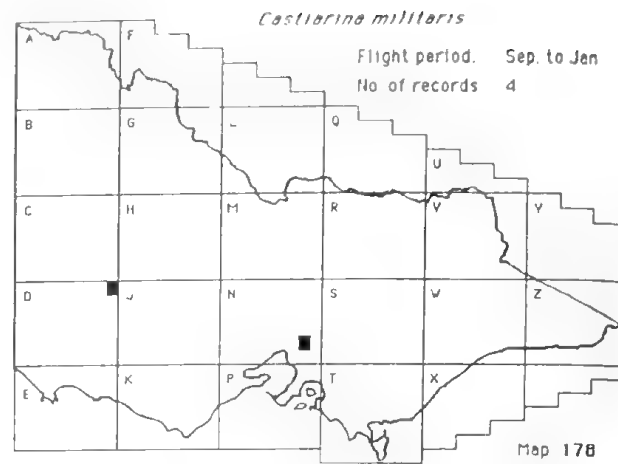
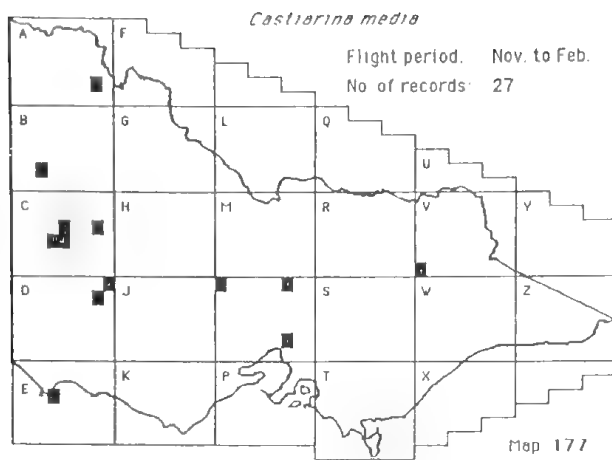






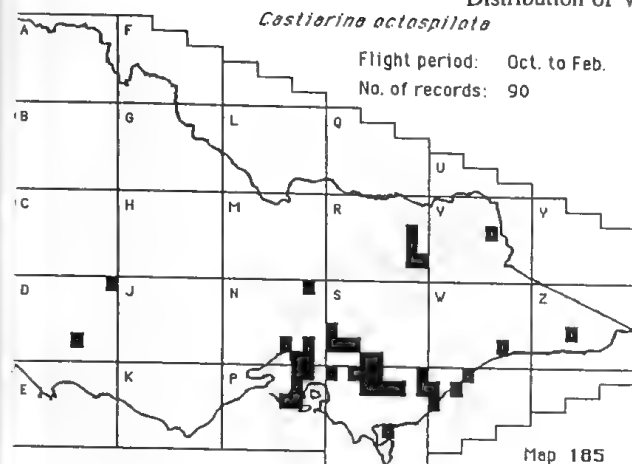






Castiarina octaspilata

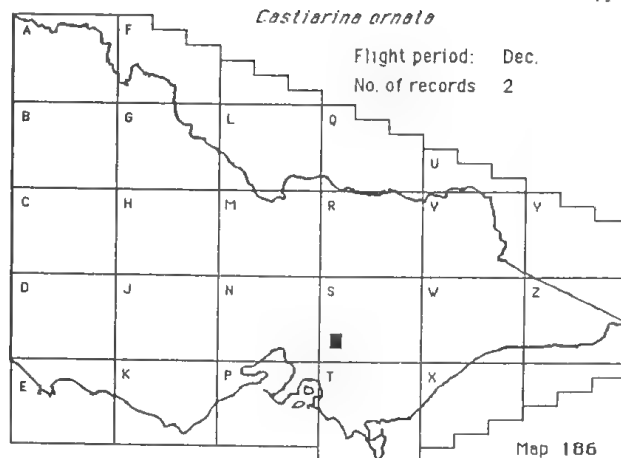
Flight period: Oct. to Feb.
No. of records: 90



Map 185

Castiarina ornata

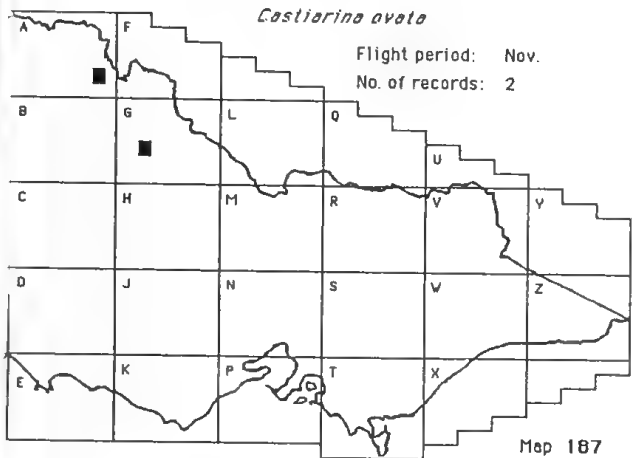
Flight period: Dec.
No. of records: 2



Map 186

Castiarina ovata

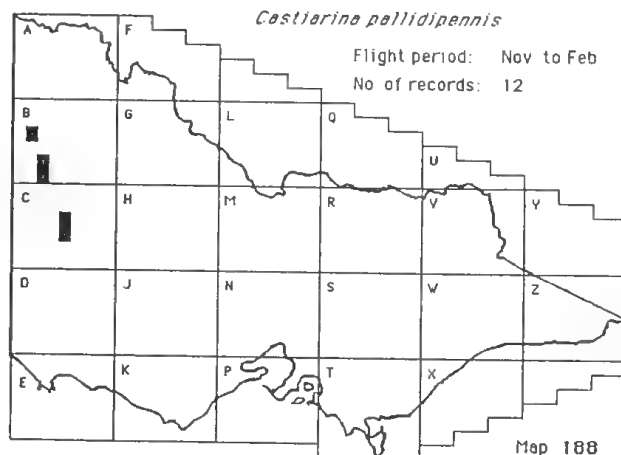
Flight period: Nov.
No. of records: 2



Map 187

Castiarina pallidipennis

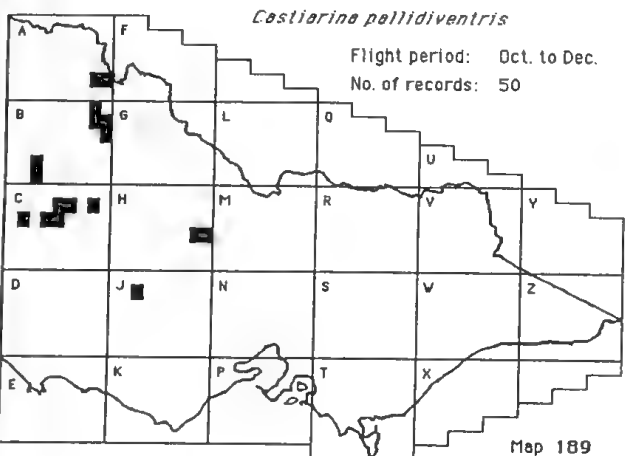
Flight period: Nov to Feb
No. of records: 12



Map 188

Castiarina pallidiventris

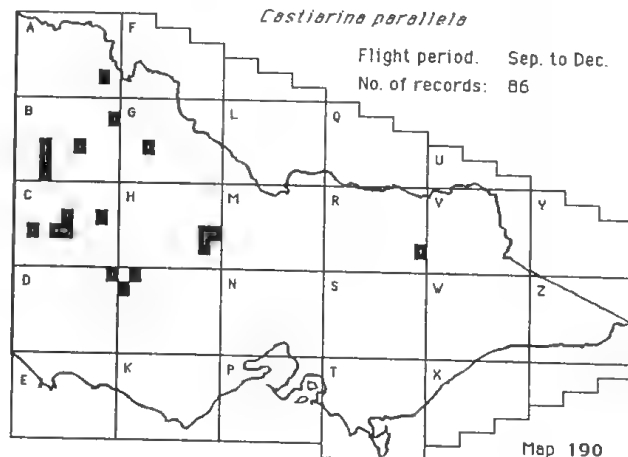
Flight period: Oct. to Dec.
No. of records: 50



Map 189

Castiarina parallela

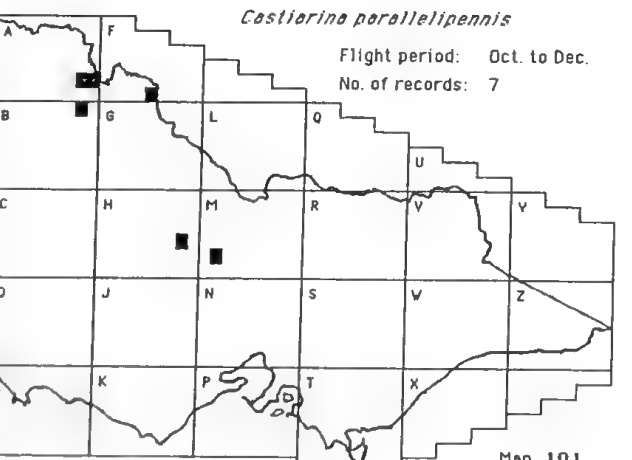
Flight period: Sep. to Dec.
No. of records: 86



Map 190

Castiarina parallelipennis

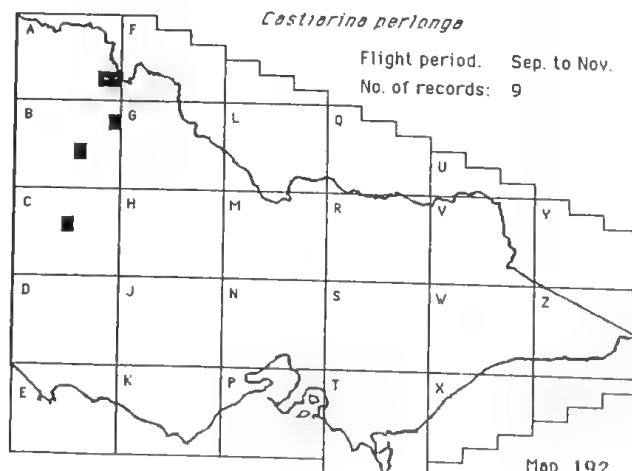
Flight period: Oct. to Dec.
No. of records: 7



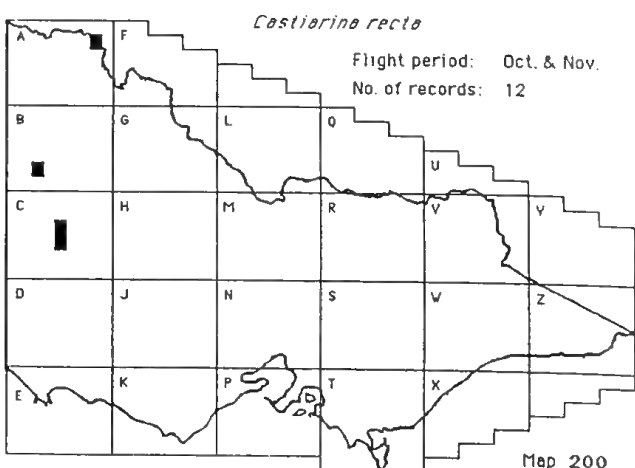
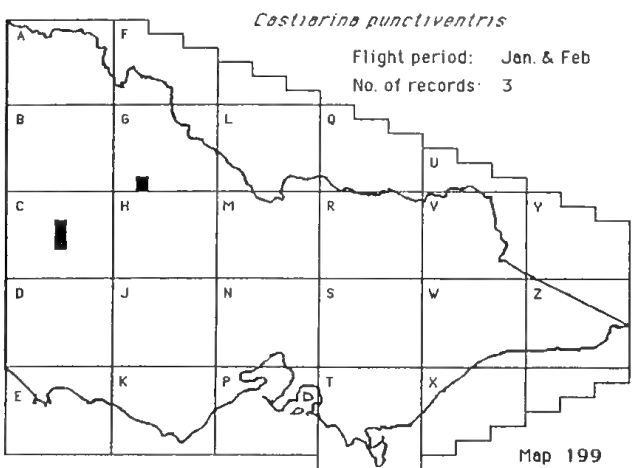
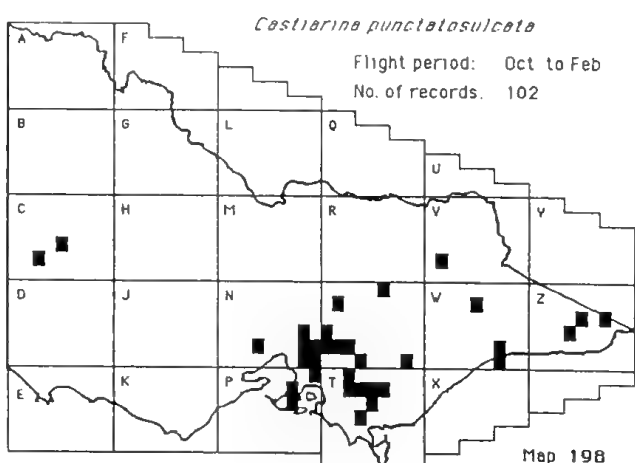
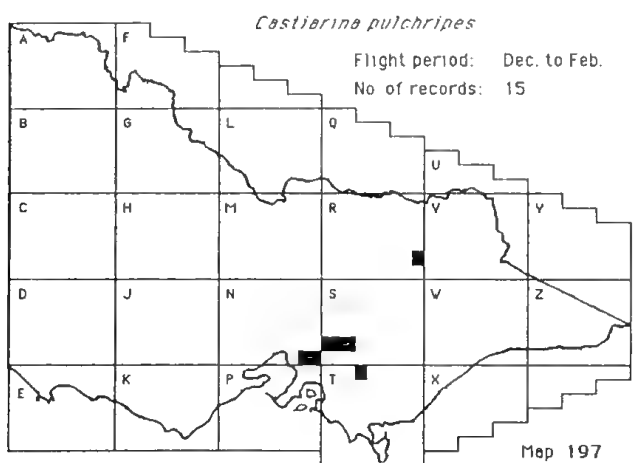
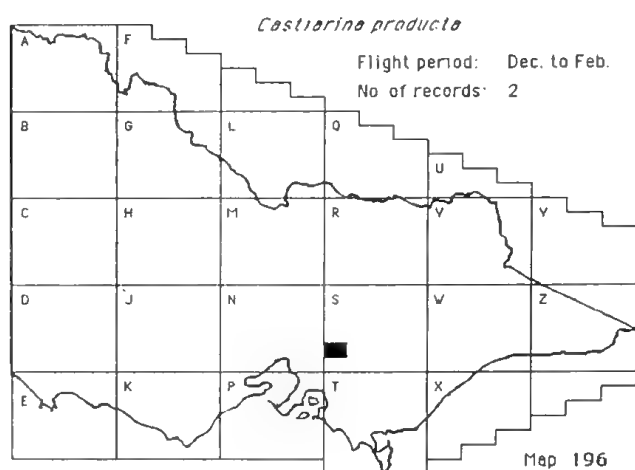
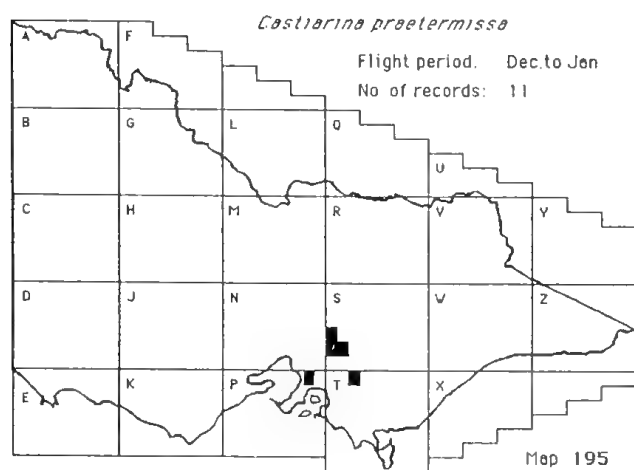
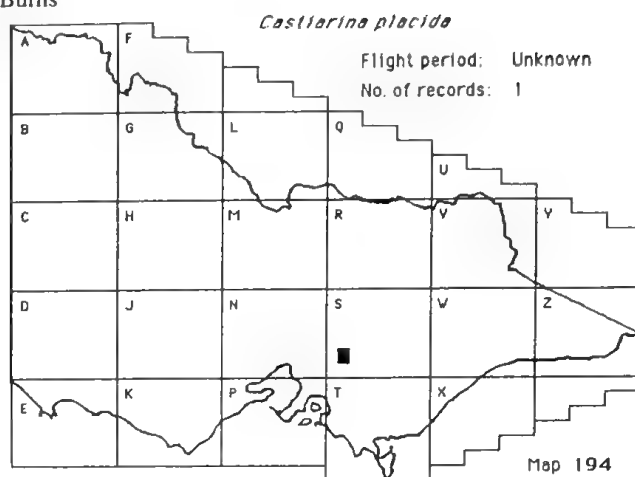
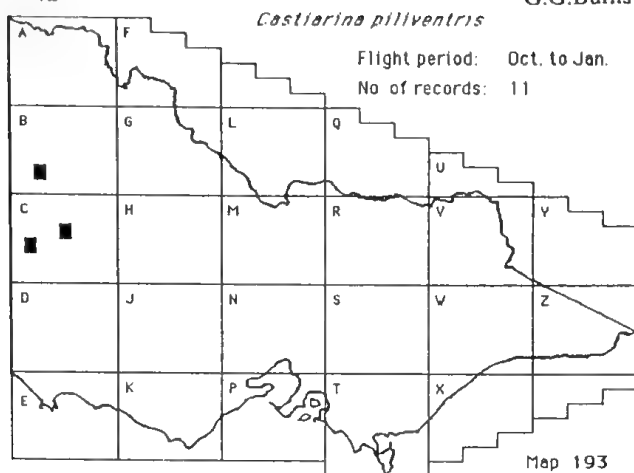
Map 191

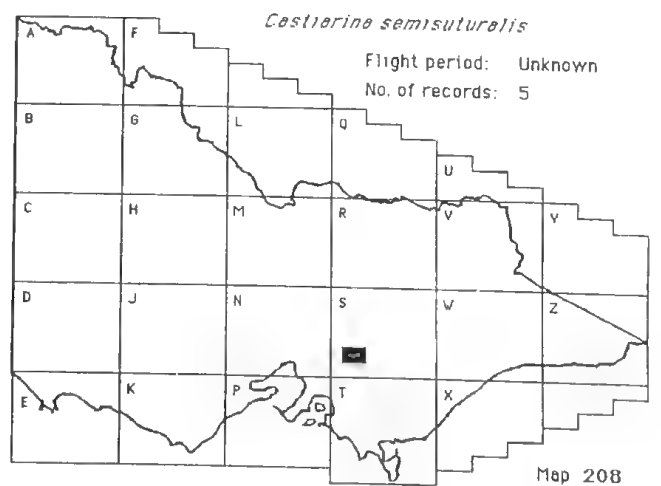
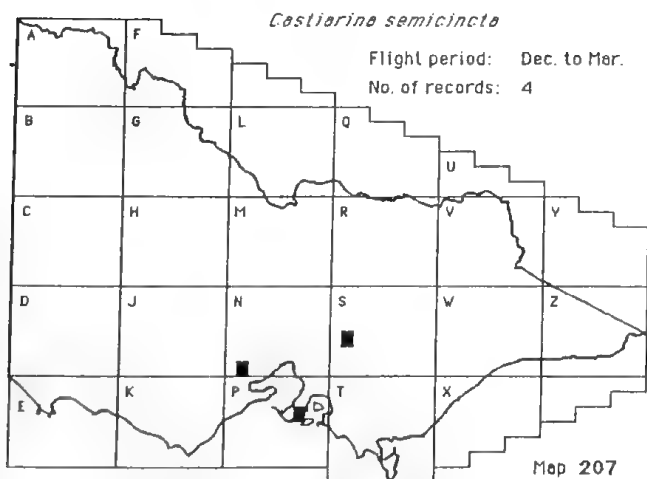
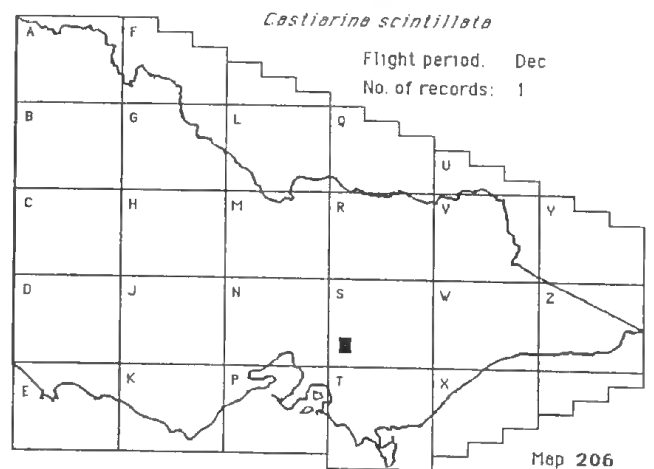
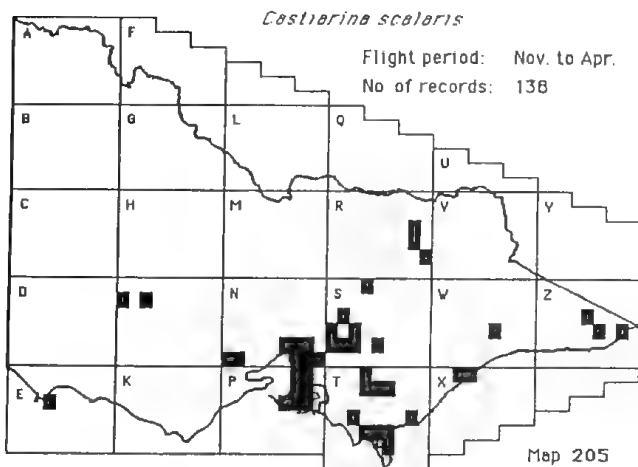
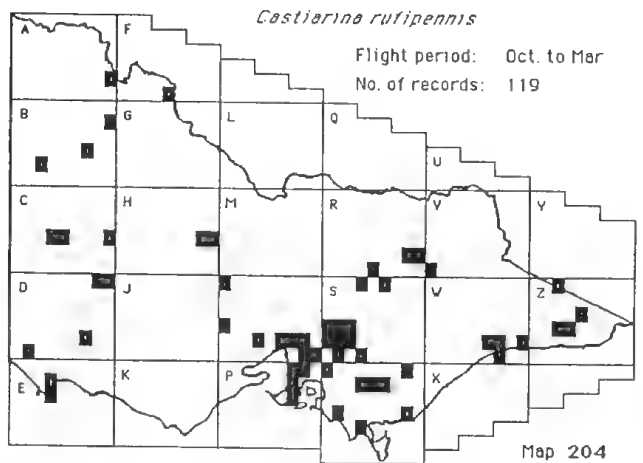
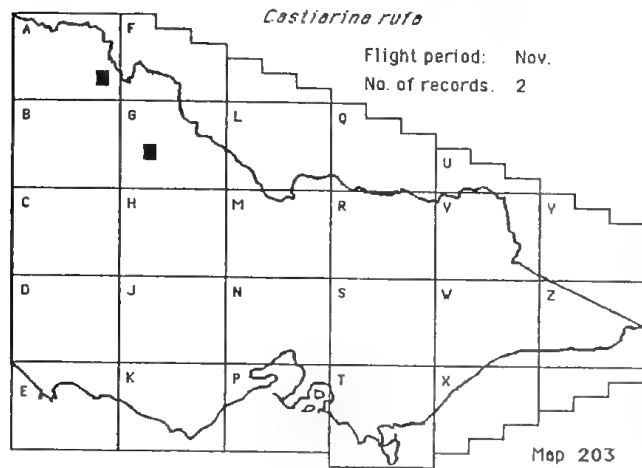
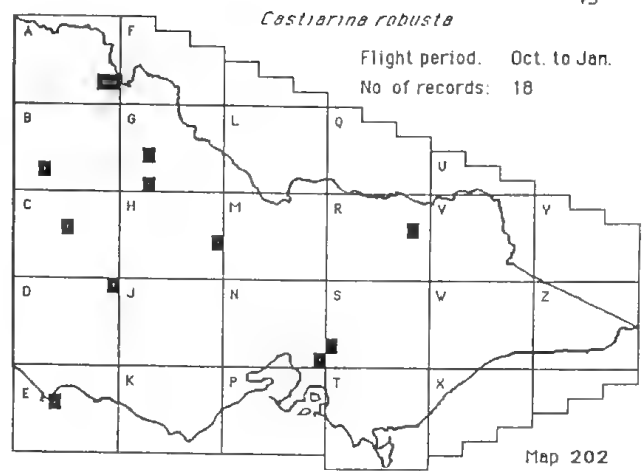
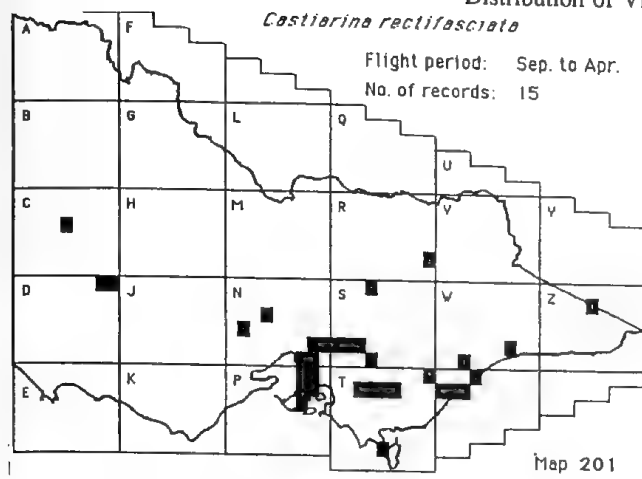
Castiarina perlonga

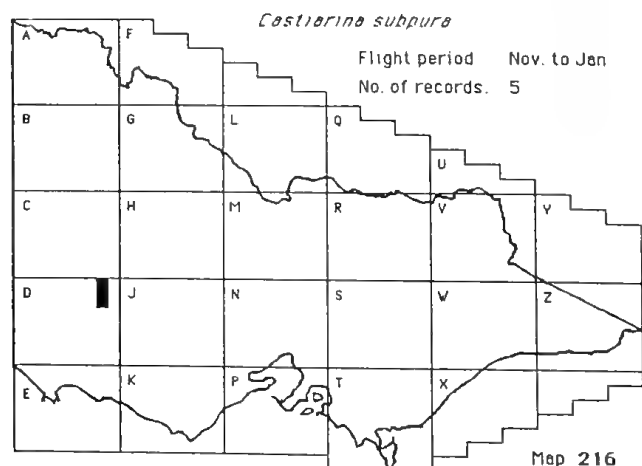
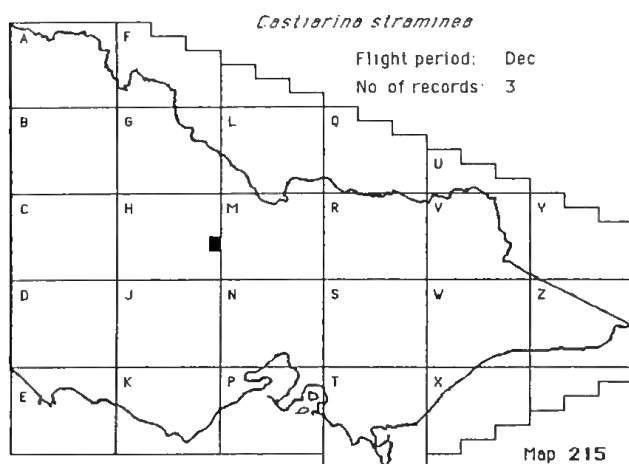
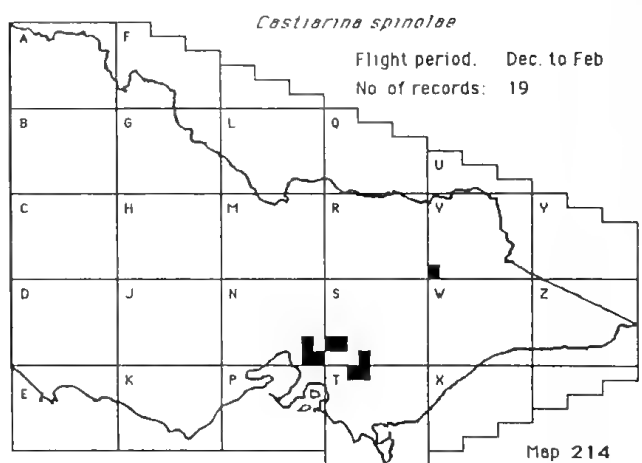
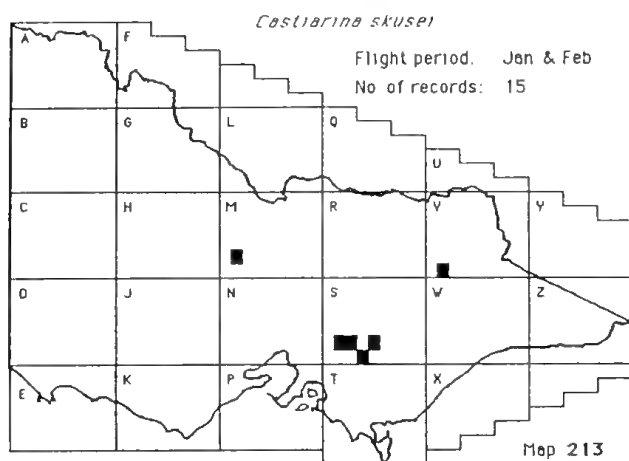
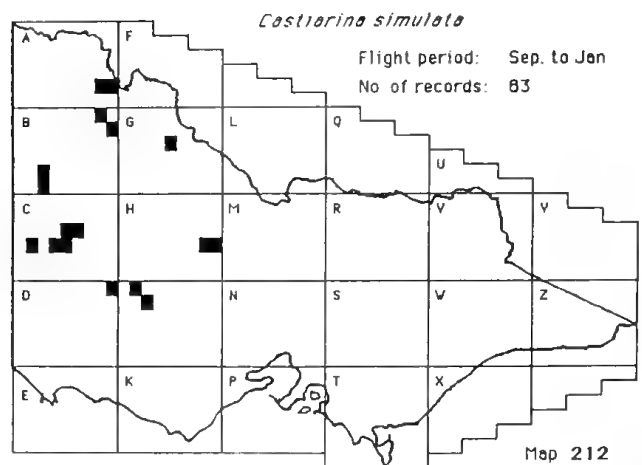
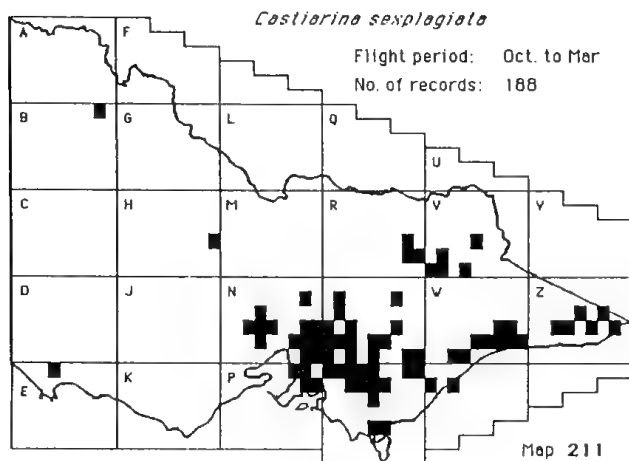
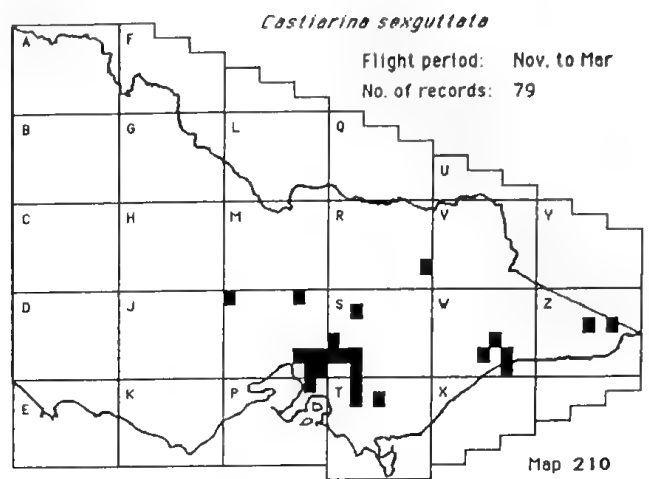
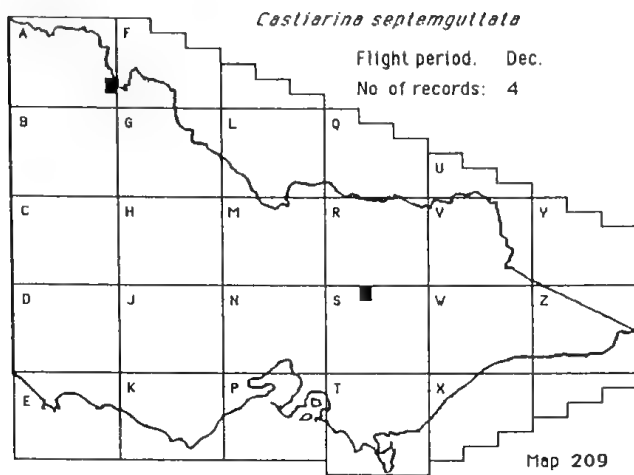
Flight period: Sep. to Nov.
No. of records: 9

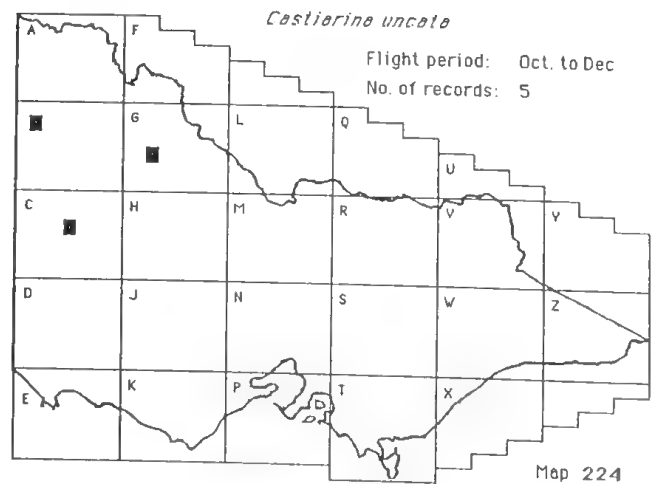
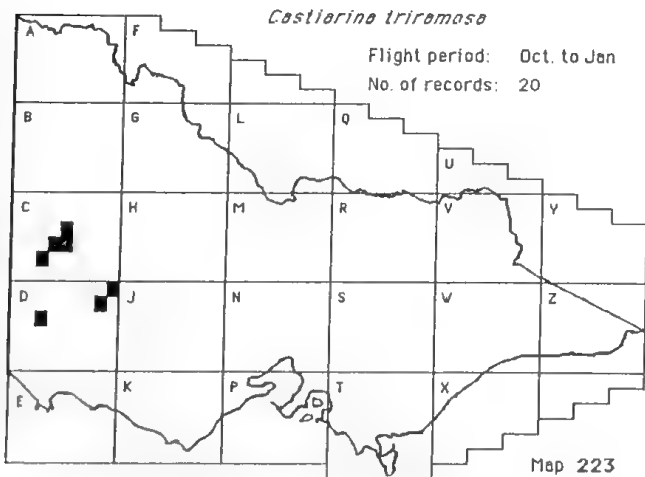
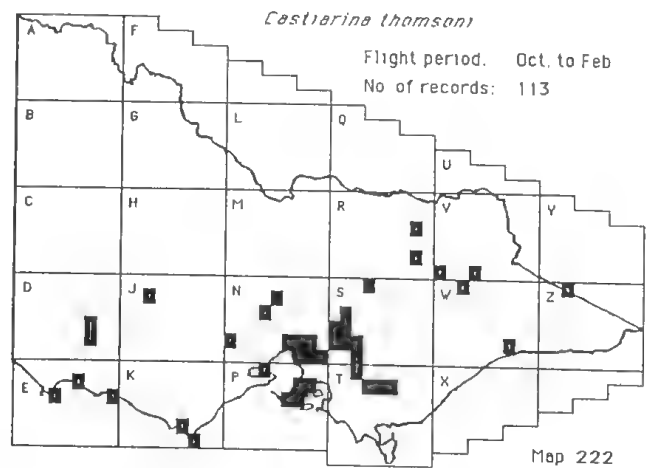
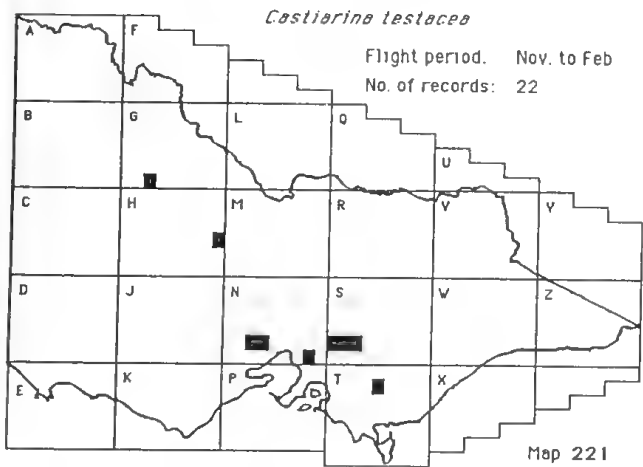
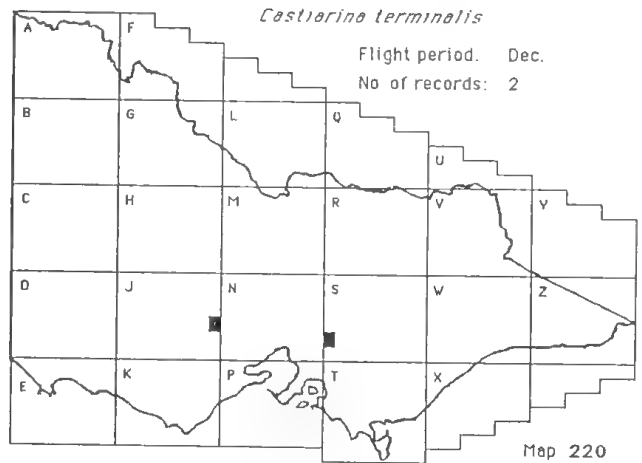
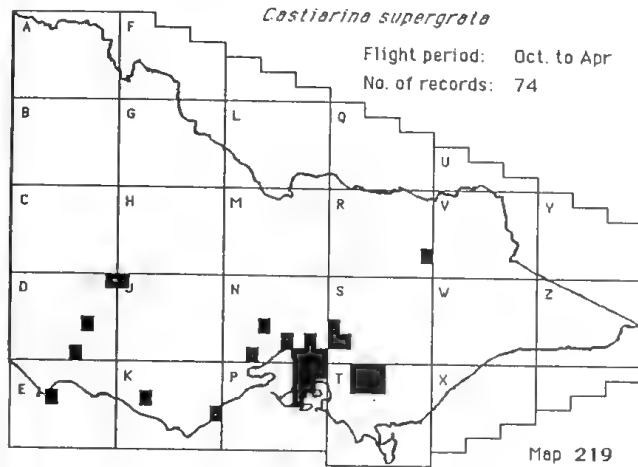
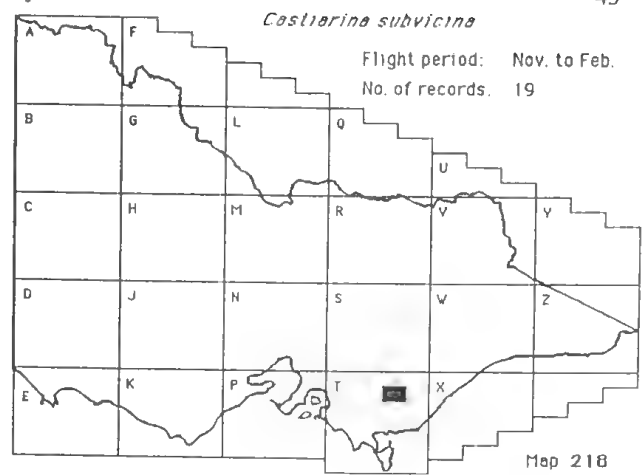
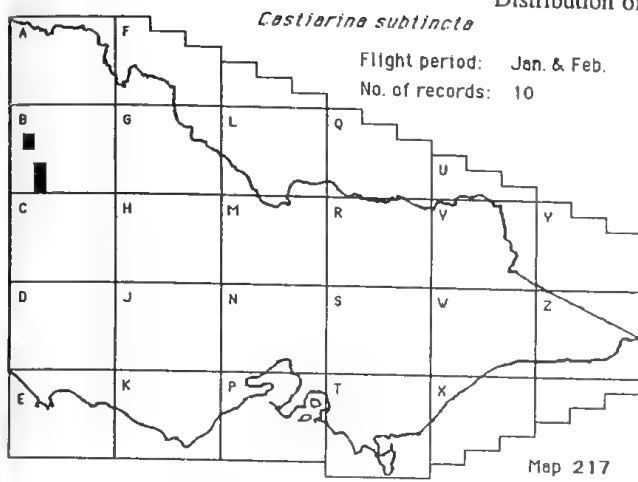


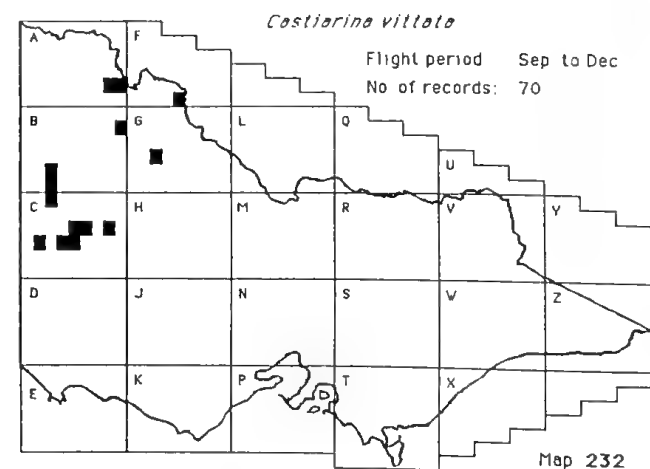
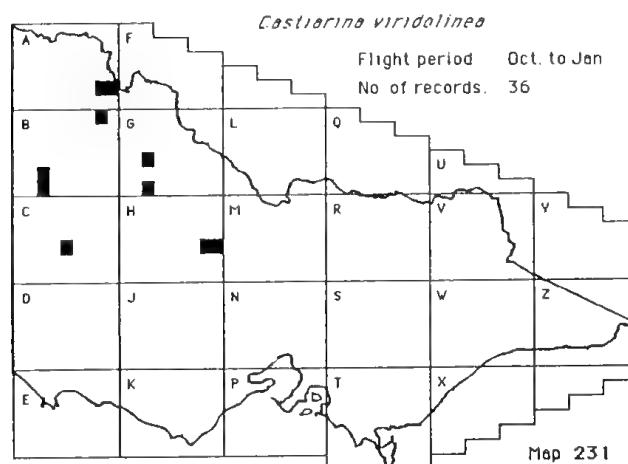
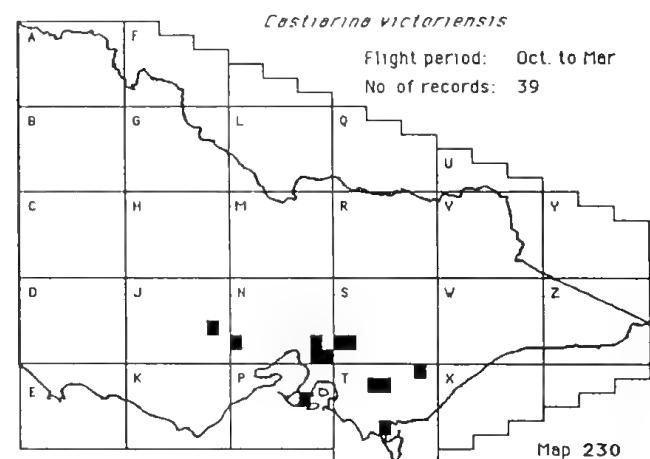
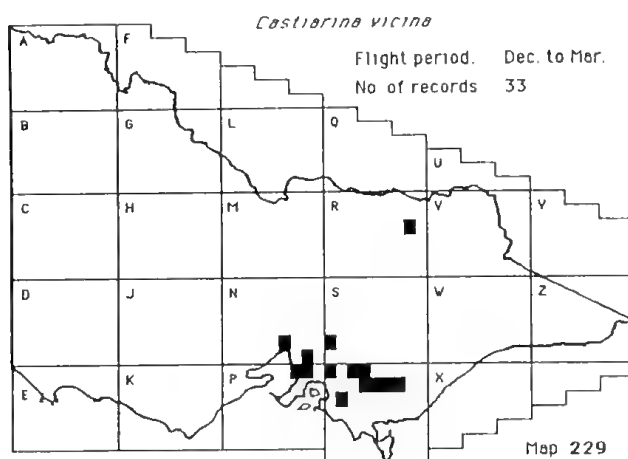
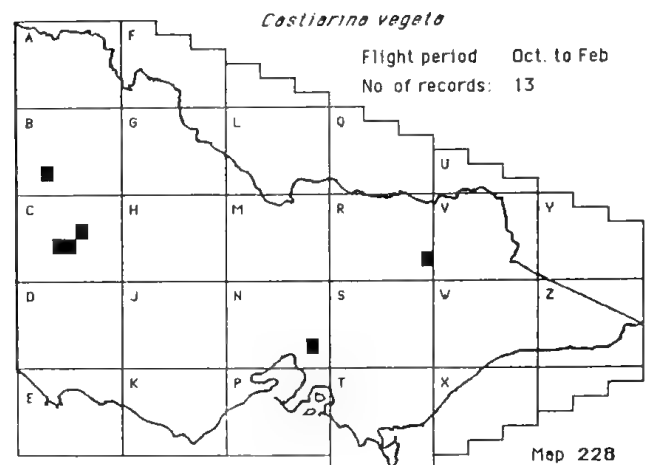
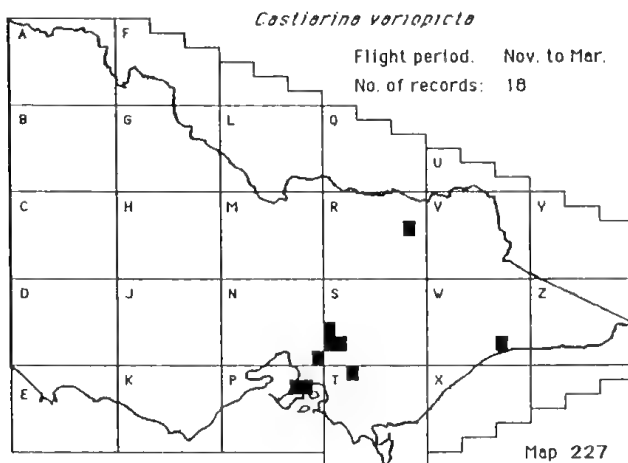
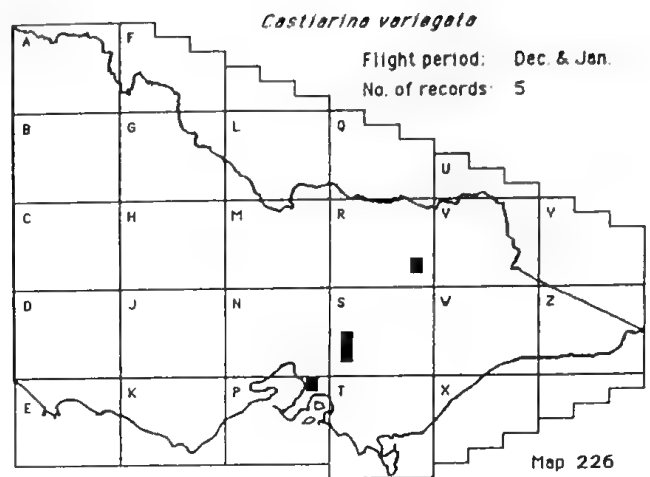
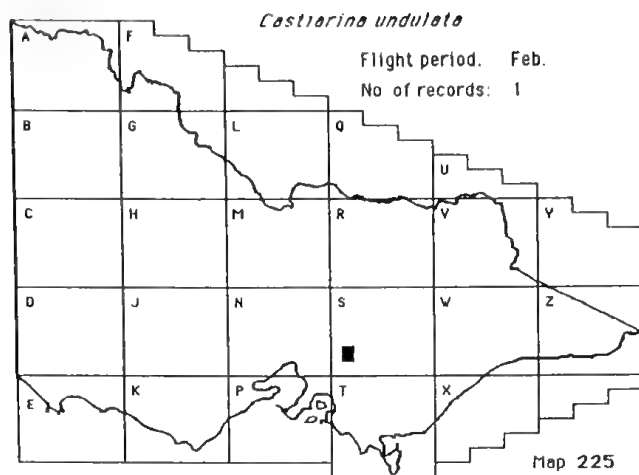
Map 192





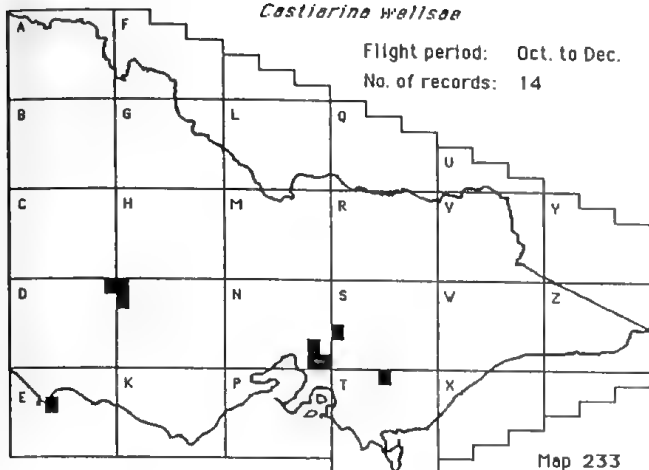






Castiarina wellsoni

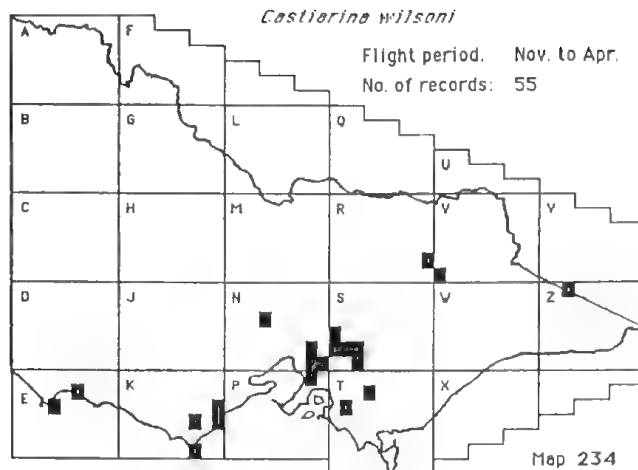
Flight period: Oct. to Dec.
No. of records: 14



Map 233

Castiarina wilsoni

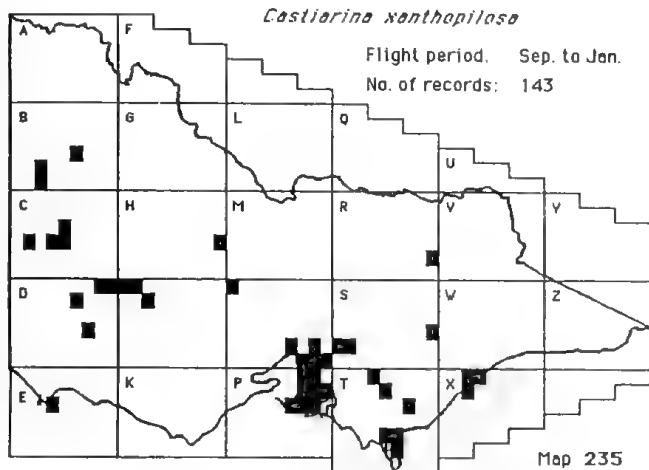
Flight period: Nov. to Apr.
No. of records: 55



Map 234

Castiarina xanthopilosa

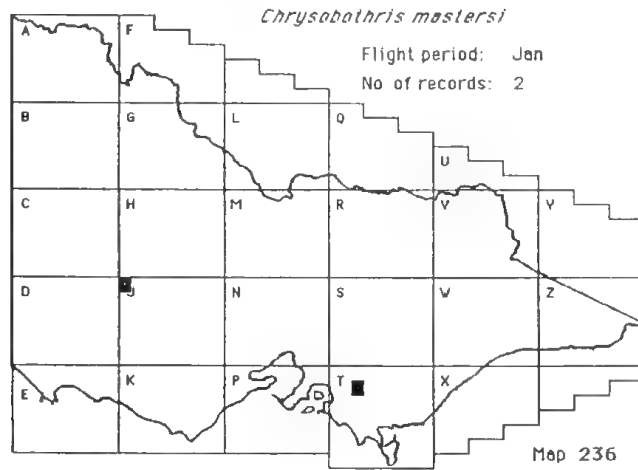
Flight period: Sep. to Jan.
No. of records: 143



Map 235

Chrysobothris mastersi

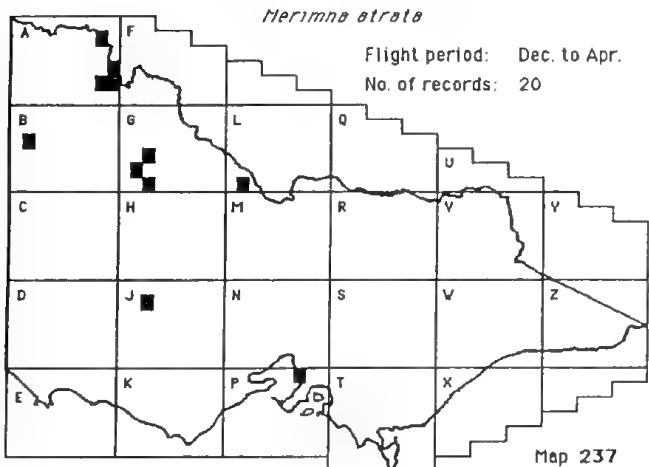
Flight period: Jan
No. of records: 2



Map 236

Merimna atrata

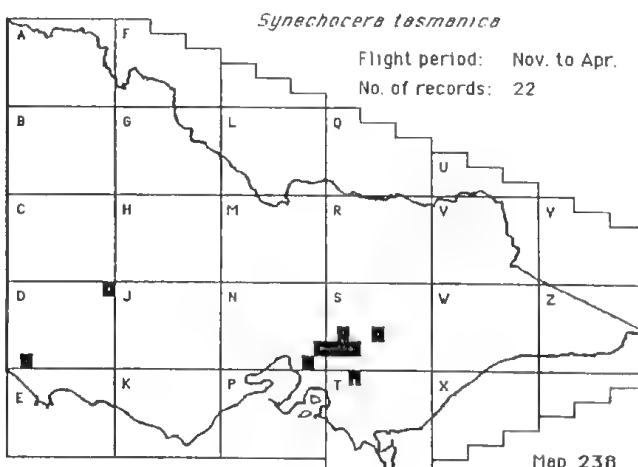
Flight period: Dec. to Apr.
No. of records: 20



Map 237

Synechocera tasmanica

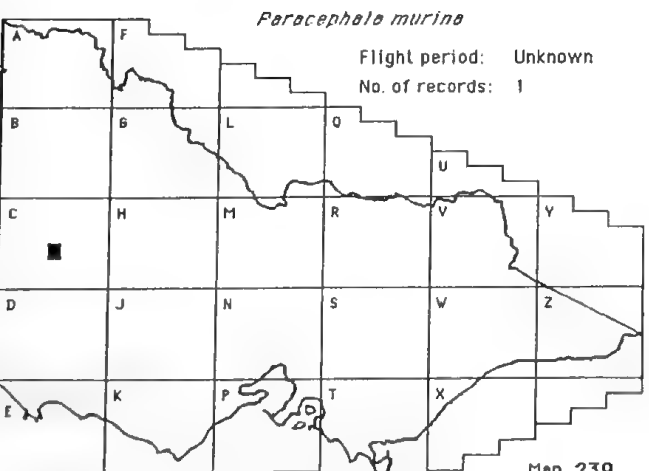
Flight period: Nov. to Apr.
No. of records: 22



Map 238

Paracephala murina

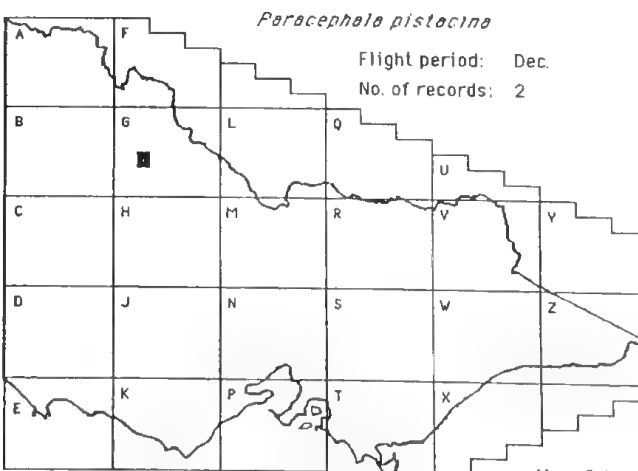
Flight period: Unknown
No. of records: 1



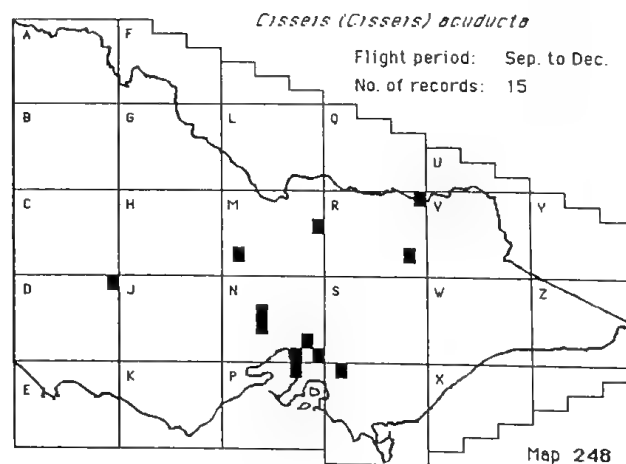
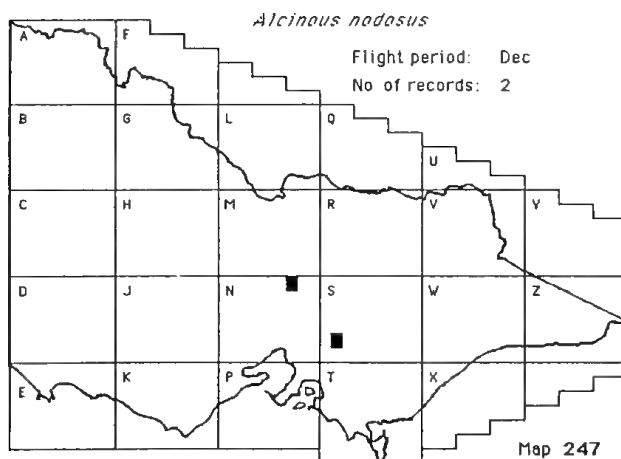
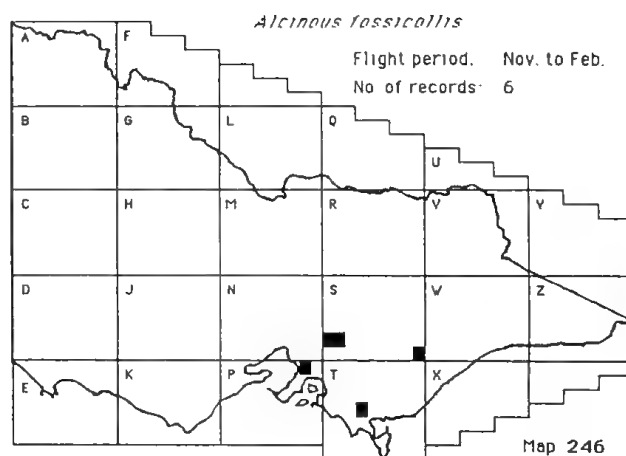
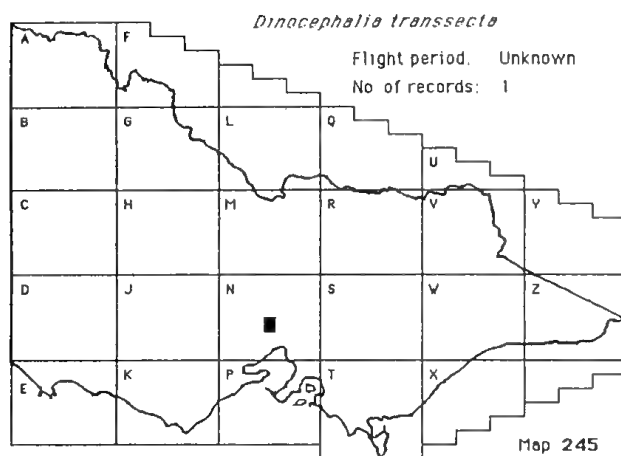
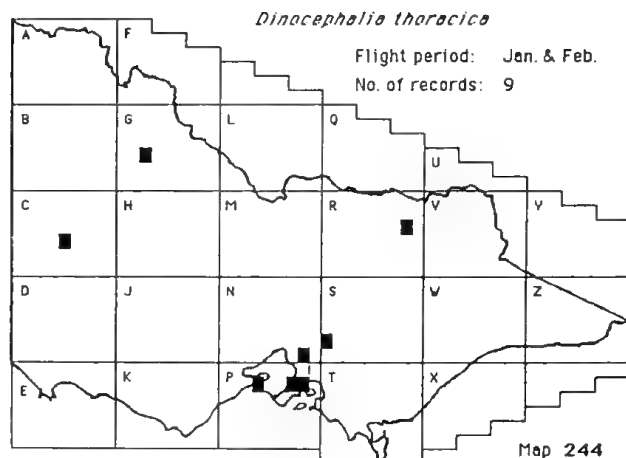
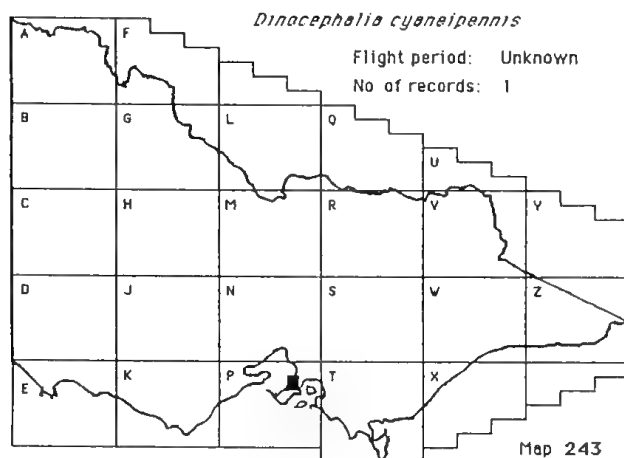
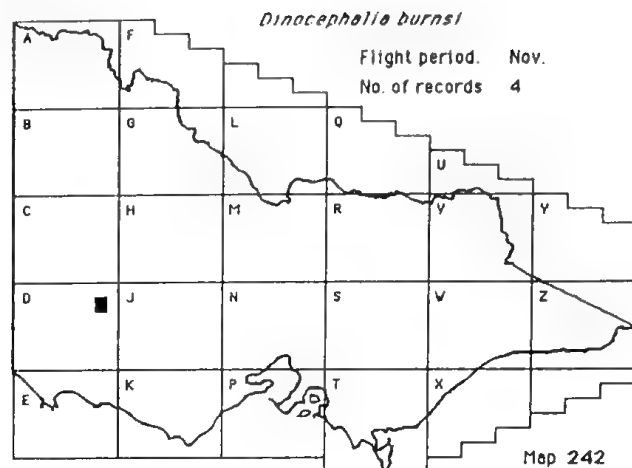
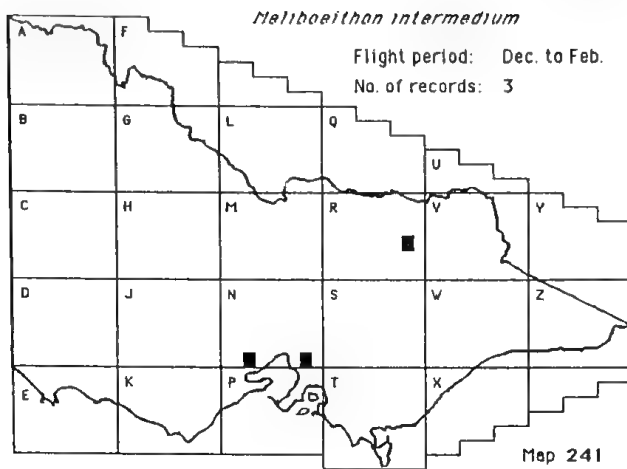
Map 239

Paracephala pistacina

Flight period: Dec.
No. of records: 2

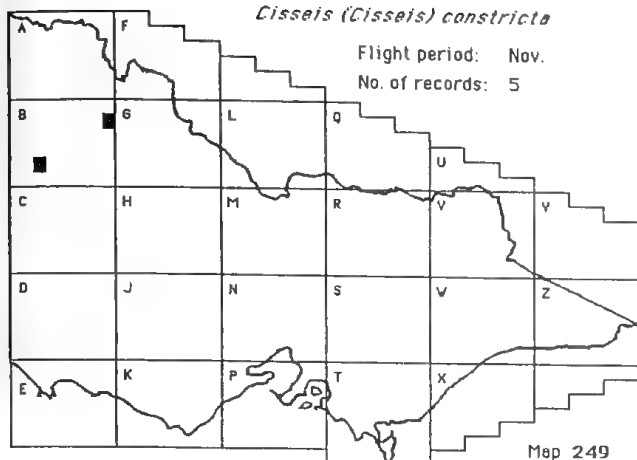


Map 240



Cisseis (Cisseis) constricta

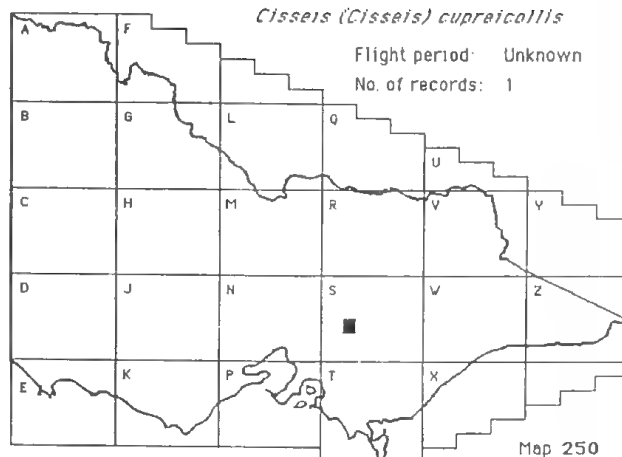
Flight period: Nov.
No. of records: 5



Map 249

Cisseis (Cisseis) cupreicollis

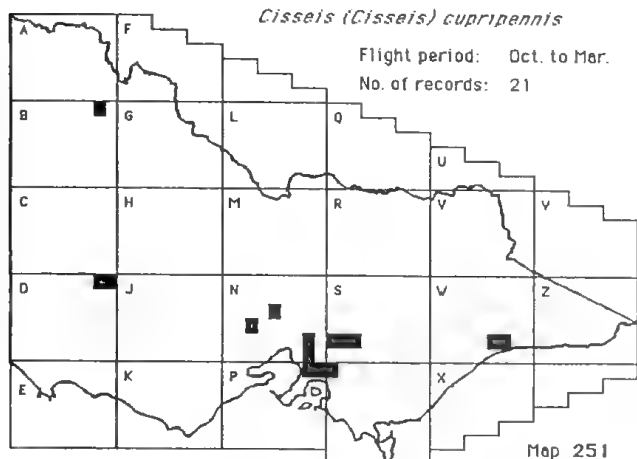
Flight period: Unknown
No. of records: 1



Map 250

Cisseis (Cisseis) cupripennis

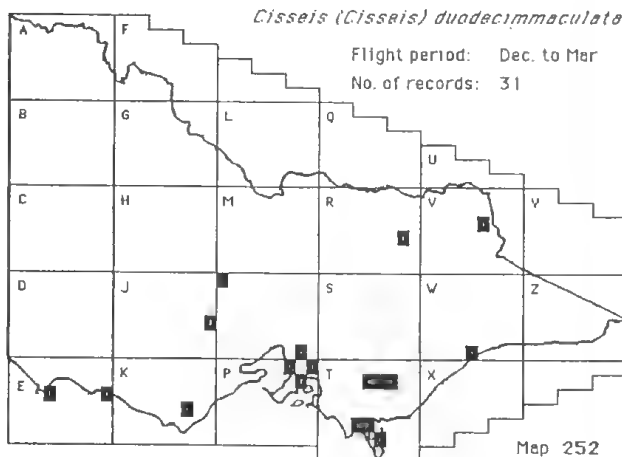
Flight period: Oct. to Mar.
No. of records: 21



Map 251

Cisseis (Cisseis) duodecimmaculata

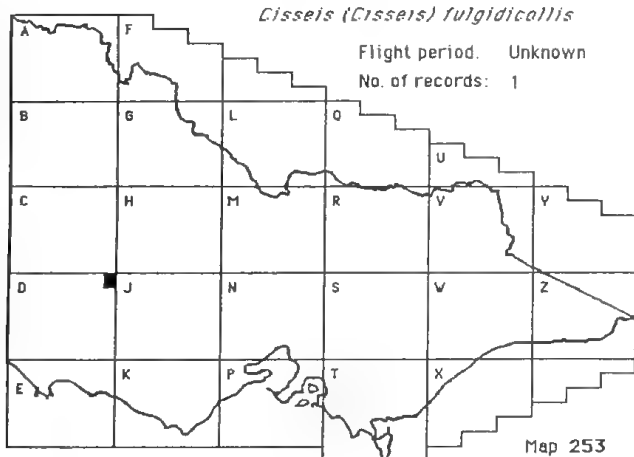
Flight period: Dec. to Mar.
No. of records: 31



Map 252

Cisseis (Cisseis) fulgidicollis

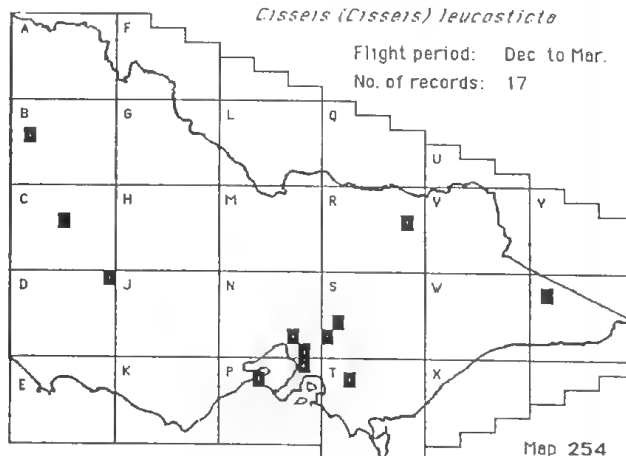
Flight period: Unknown
No. of records: 1



Map 253

Cisseis (Cisseis) leucosticta

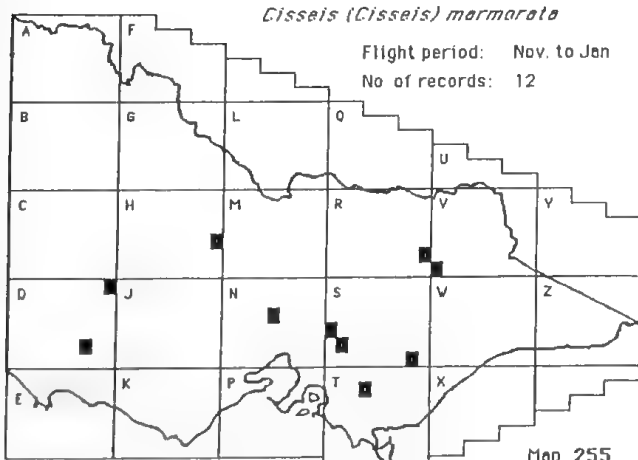
Flight period: Dec to Mar.
No. of records: 17



Map 254

Cisseis (Cisseis) marmorata

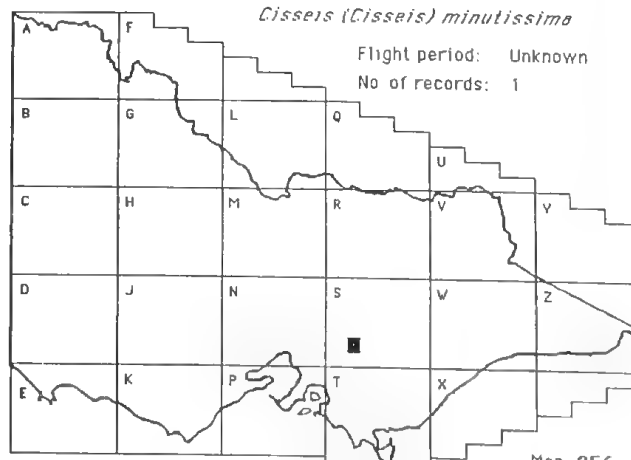
Flight period: Nov. to Jan.
No. of records: 12



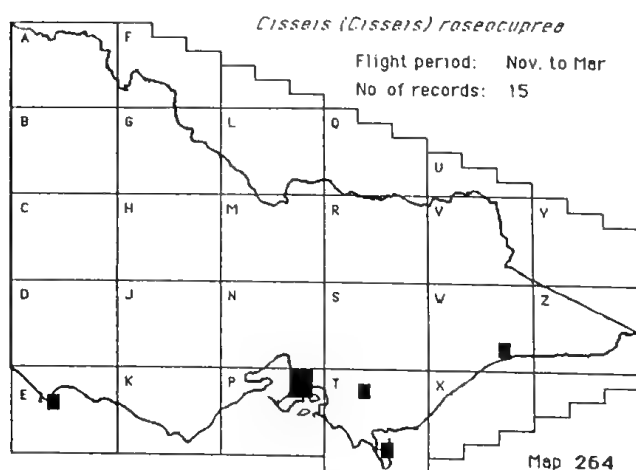
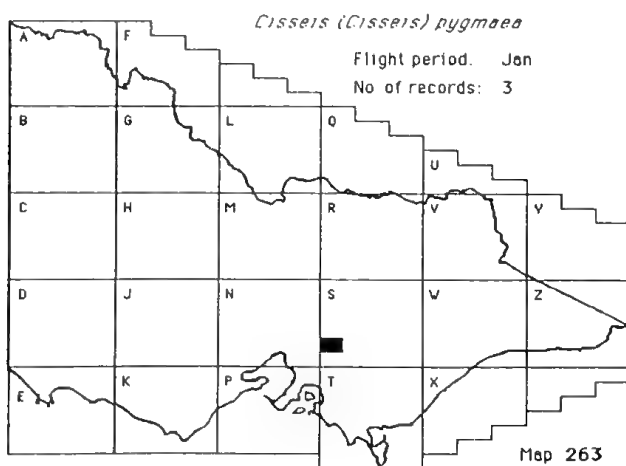
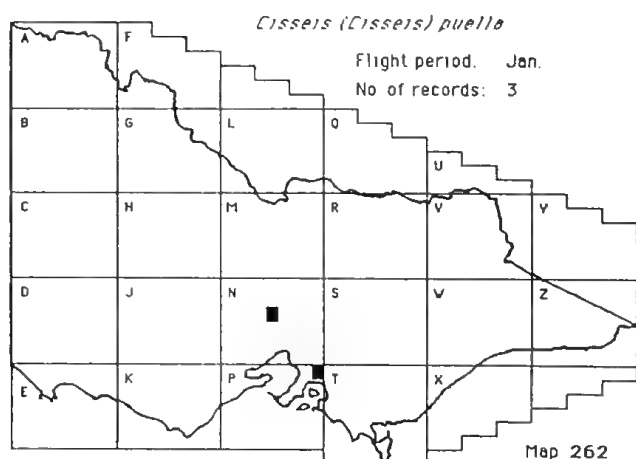
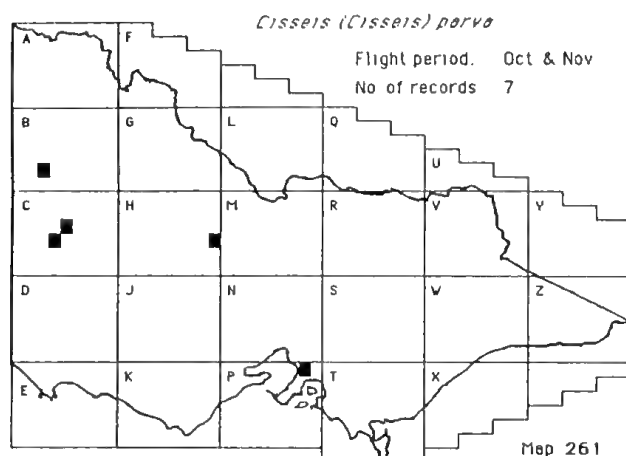
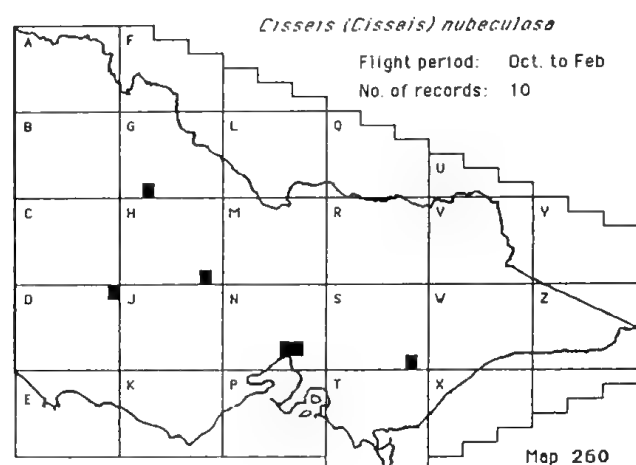
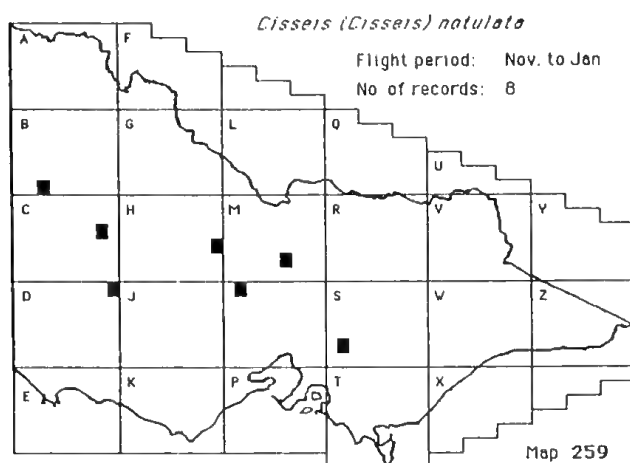
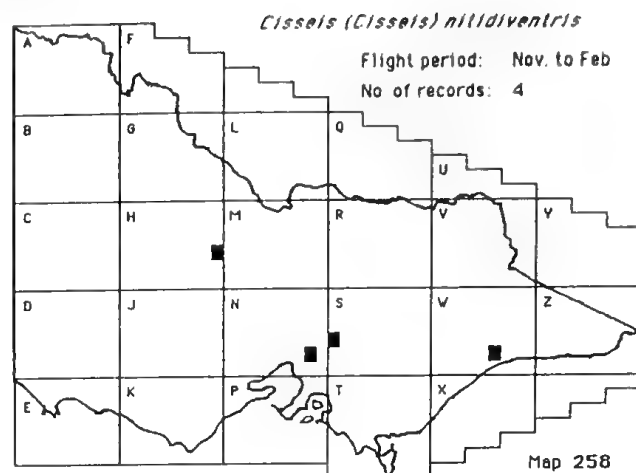
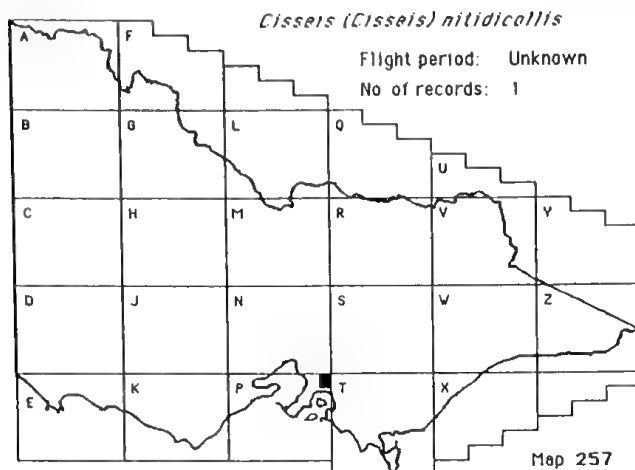
Map 255

Cisseis (Cisseis) minutissima

Flight period: Unknown
No. of records: 1

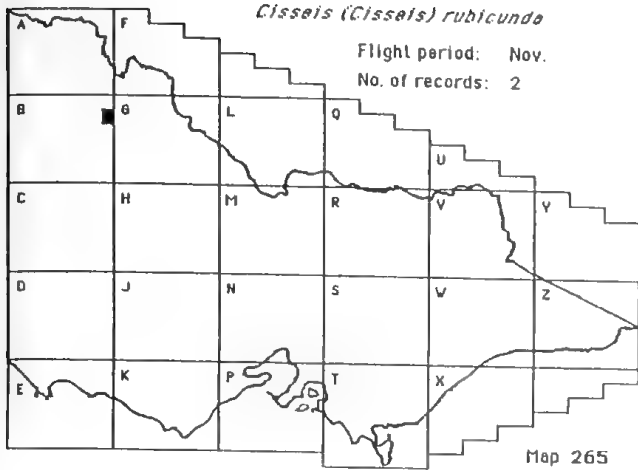


Map 256



Cisseis (Cisseis) rubicunda

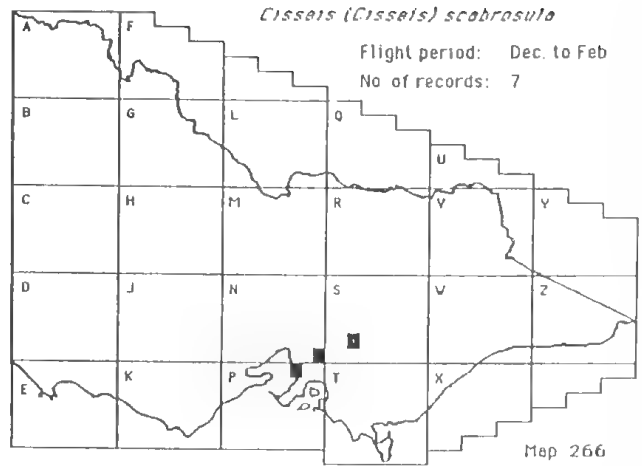
Flight period: Nov.
No. of records: 2



Map 265

Cisseis (Cisseis) scabrosula

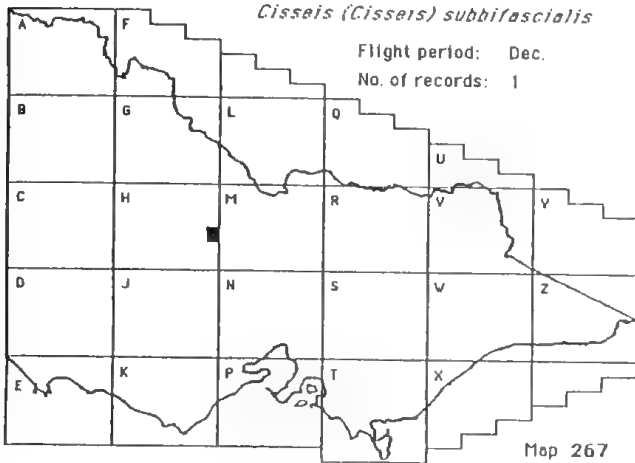
Flight period: Dec. to Feb
No. of records: 7



Map 266

Cisseis (Cisseis) subfascialis

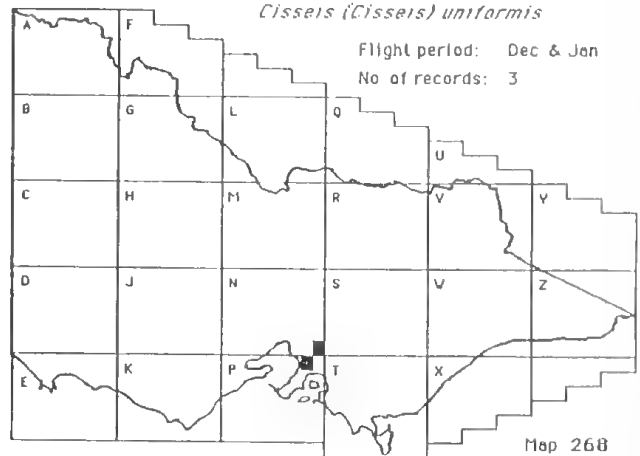
Flight period: Dec.
No. of records: 1



Map 267

Cisseis (Cisseis) uniformis

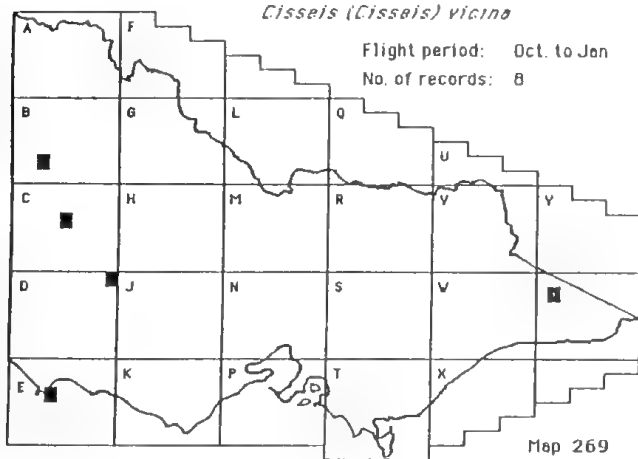
Flight period: Dec & Jan
No. of records: 3



Map 268

Cisseis (Cisseis) vicina

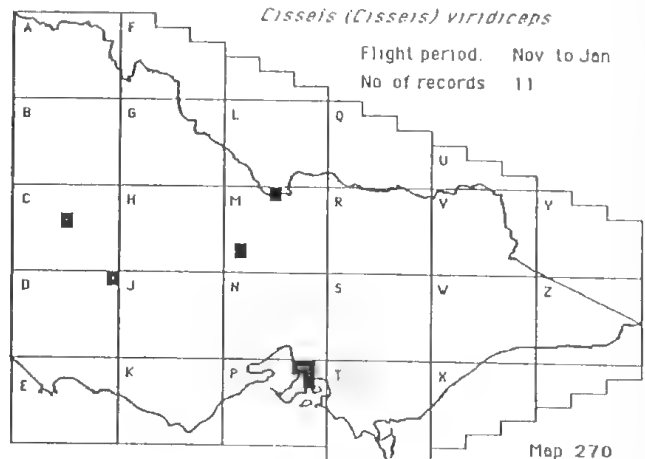
Flight period: Oct. to Jan
No. of records: 8



Map 269

Cisseis (Cisseis) viridiceps

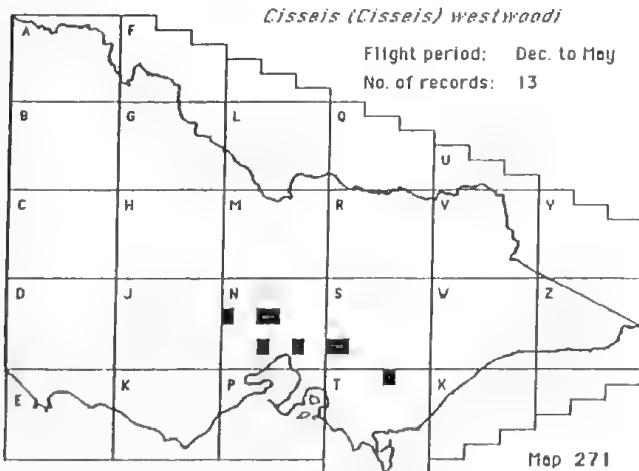
Flight period: Nov to Jan
No. of records: 11



Map 270

Cisseis (Cisseis) westwoodi

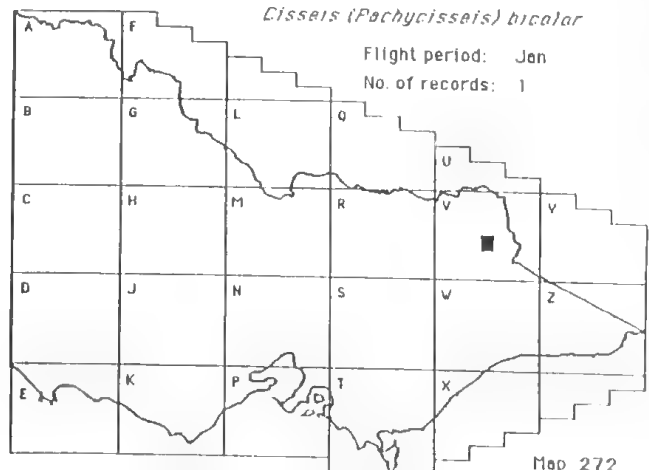
Flight period: Dec. to May
No. of records: 13



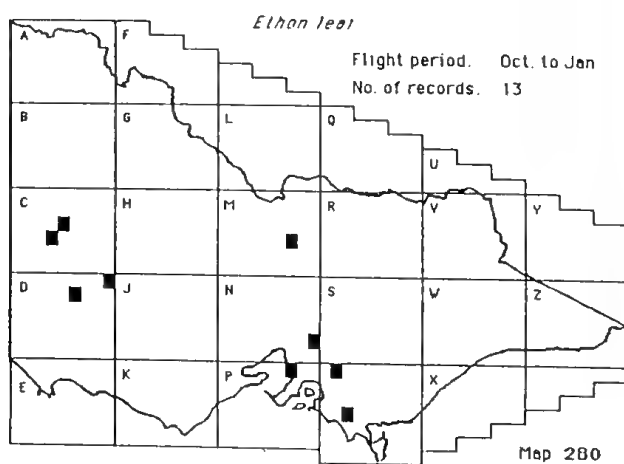
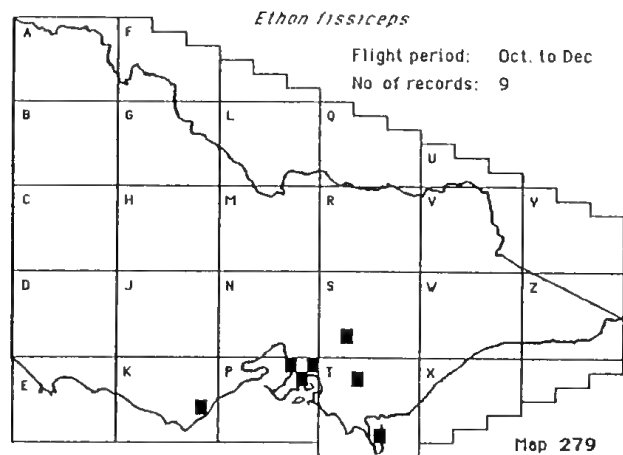
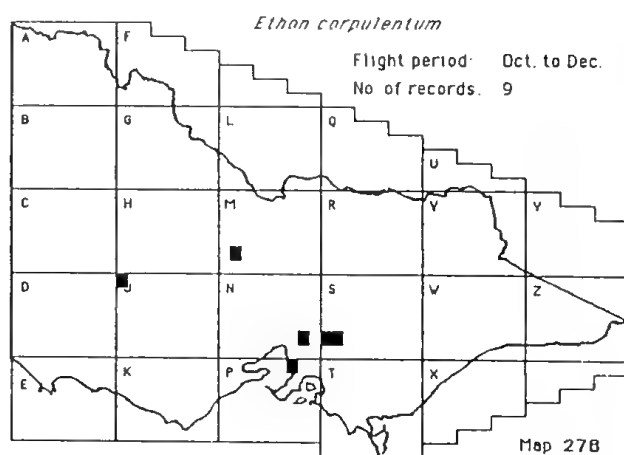
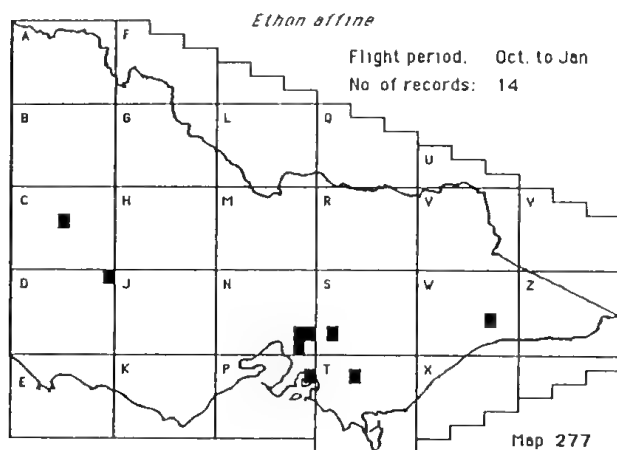
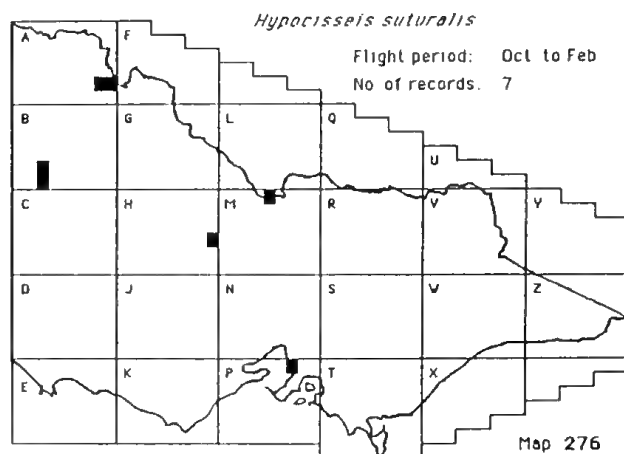
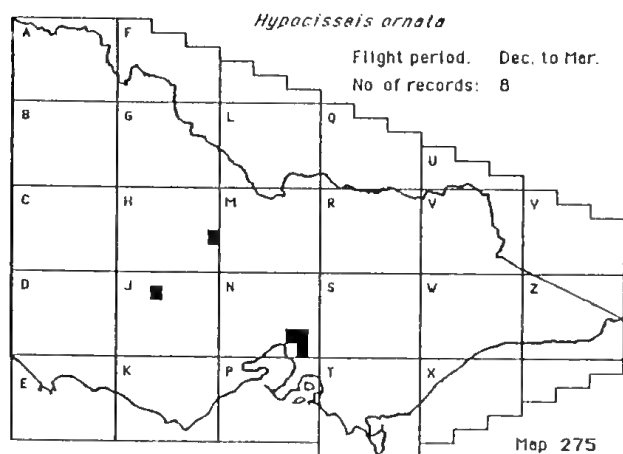
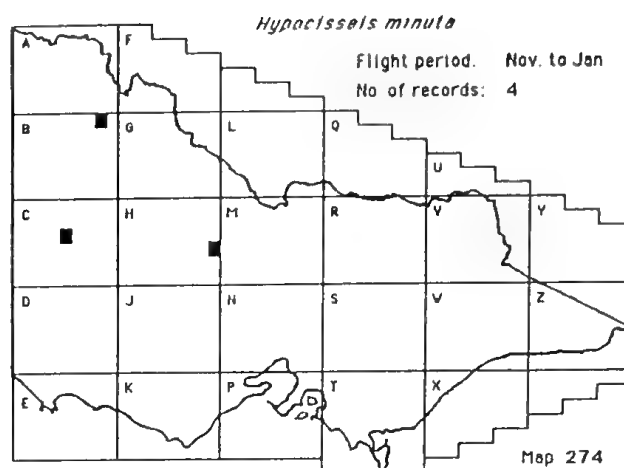
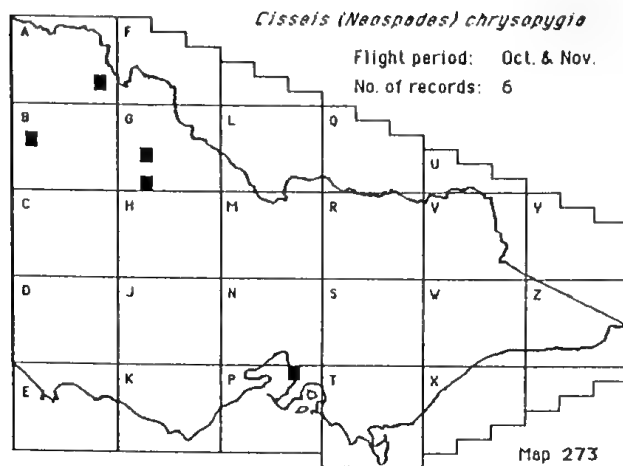
Map 271

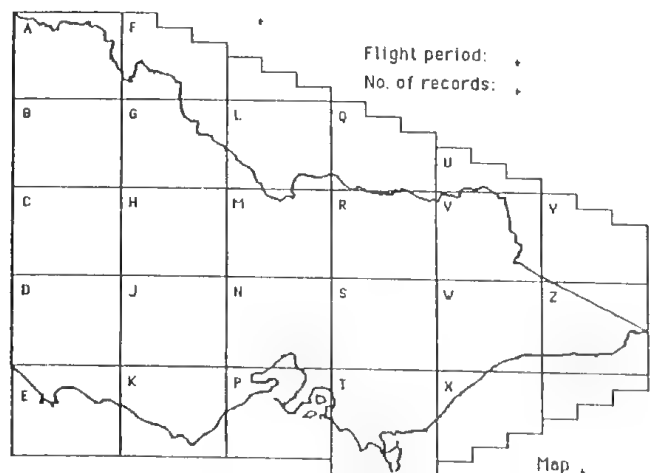
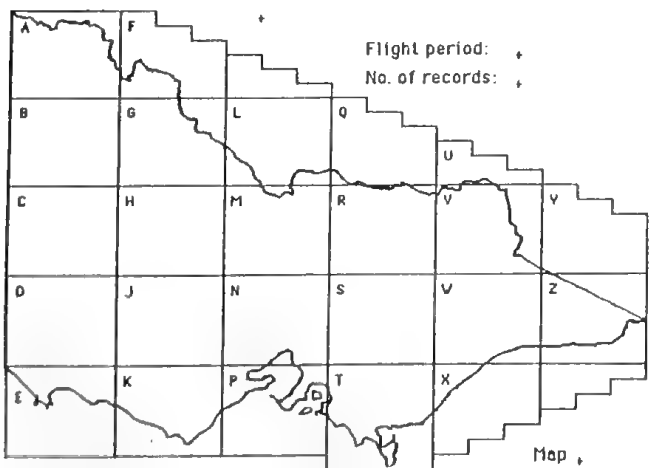
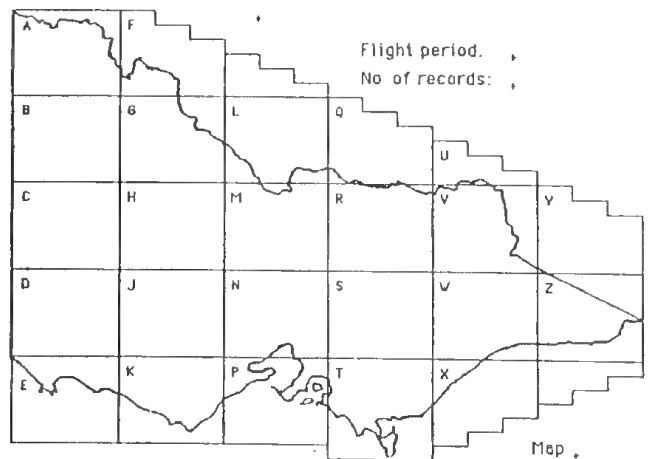
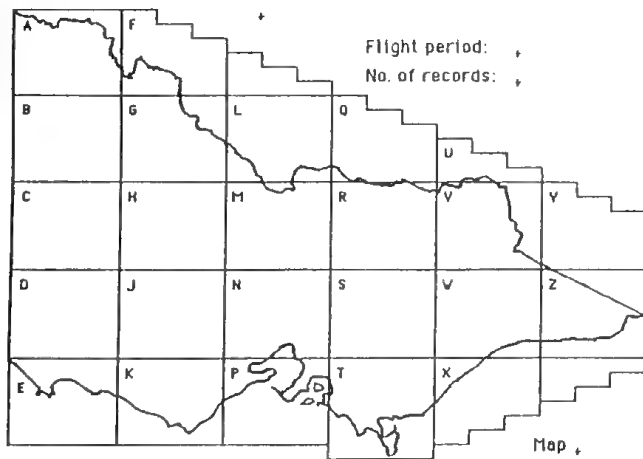
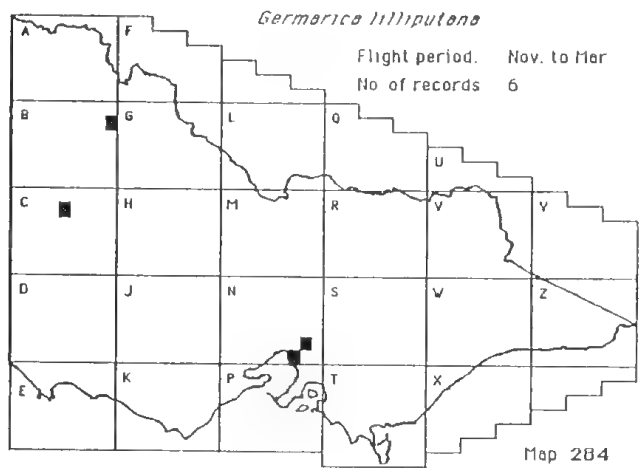
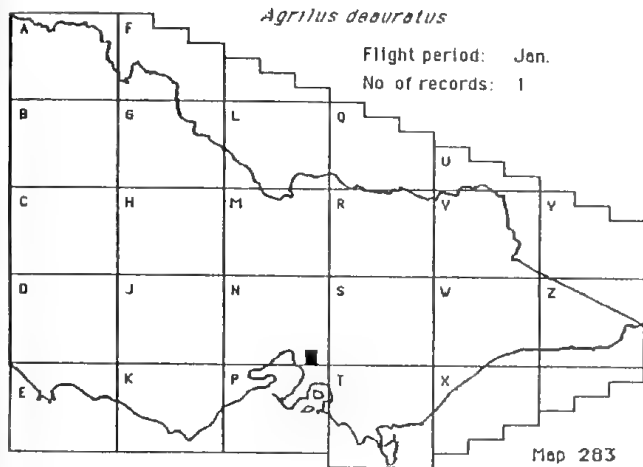
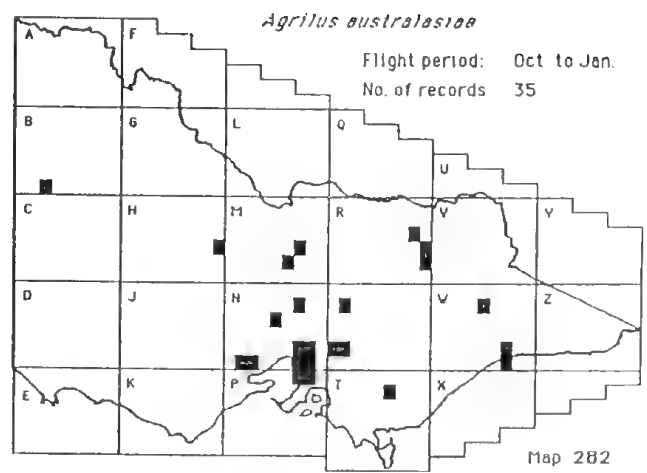
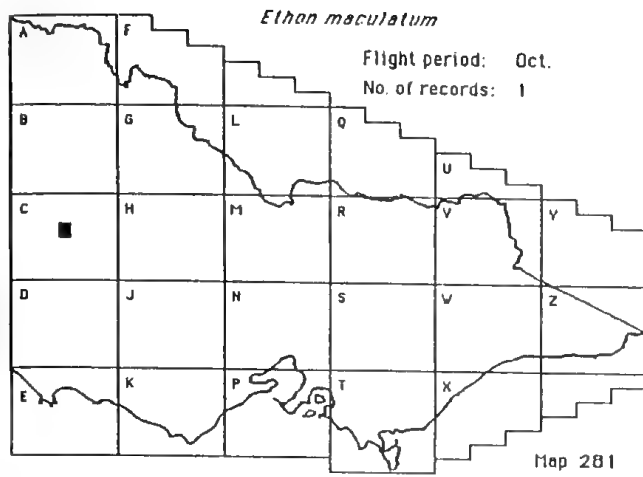
Cisseis (Pachycisseis) bicolor

Flight period: Jan
No. of records: 1



Map 272





The macroinvertebrate assemblages in pools and riffles in two intermittent streams (Werribee and Lerderderg Rivers, southern central Victoria)

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*Present address: Department of Zoology, University of Adelaide, GPO Box 498, Adelaide 5001, South Australia

Abstract. Boulton, A.J. and Lake, P.S. 1992. The macroinvertebrate assemblages in pools and riffles in two intermittent streams (Werribee and Lerderderg Rivers, southern central Victoria). *Occasional Papers from the Museum of Victoria* 5: 55-71.

The macroinvertebrate assemblages in pools and riffles at four study sites in the upper reaches of the Werribee and Lerderderg Rivers in central Victoria were sampled from 1982 to 1984 using box and suction samplers and sweep nets. A total of 258 invertebrate taxa was collected in 1175 samples: Mollusca (6 taxa), Crustacea (9), Ephemeroptera (5), Odonata (5), Plecoptera (15), Hemiptera (9), Coleoptera (64), Diptera (89), Trichoptera (29), and 27 other taxa. These taxa and their occurrences in each habitat at each site during each year are listed. Species richness was greater in pools than in riffles in both rivers, probably reflecting the importance of these habitats as refuges when the streams dry. Each taxon was assigned to a functional feeding group based on mouth-part morphology, gut contents, observations in the laboratory, and published dietary studies. Predators were always the most diverse feeding group in all habitats (39% of all aquatic taxa) whereas relatively few taxa were collector-filterers (6%) or collector-shredders (5%). Collector-scrappers and collector-gatherers comprised 19% and 17% respectively, representing most of the individuals, and illustrating the generalized feeding modes of many stream invertebrates.

Introduction

Despite the fact that intermittent and ephemeral streams drain over half of mainland Australia (Williams, 1983), we know very little about the ecology of these streams (Boulton and Suter, 1986). With the growing recognition of the importance of water in arid zones and the fragility of their aquatic systems (e.g., Ponder, 1986), it is crucial that fundamental survey data collected in intermittent streams in Australia are readily available. This need has partly been met for permanent streams in south-eastern Australia, by the publication of distribution lists of aquatic invertebrates (e.g., LaTrobe River, Victoria, Marchant et al., 1984).

It has also been suggested that intermittent streams in arid and semi-arid zones are likely to be highly sensitive harbingers of global climate change (Dahm and Molles, 1990). However, their variability in flow, especially in Australia (Finlayson and McMahon, 1988), means that relatively long-term and intensive studies are required in order to describe fully the resulting variation in community and ecosystem variables. This intensive faunal study of the Werribee and Lerderderg Rivers provides a baseline spanning two years of different flow regimes at four sites for comparison with data collected in the future during studies on long-term climatic change (cf. Gore et al., 1990). Furthermore, understanding the spatial and temporal heterogeneity of lotic systems and the "ecological strategies" used by the biota to persist

in these habitats are critical to assessing recovery after anthropogenic disturbances (Poff and Ward, 1990).

This paper presents a complete species list and summarizes distributional data for all aquatic taxa collected from pools and riffles at four sites along the upper reaches of the Werribee and Lerderderg Rivers from 1982 to 1984. Furthermore, each taxon was assigned to a functional feeding group (cf. Cummins and Klug, 1979; Hawkins and Sedell, 1981) because this classification can provide a useful basis for comparisons among studies involving different taxa or dealing with functional variation along river systems (Vannote et al., 1980; Marchant et al., 1985) or among sites (Bunn, 1986). We do not attempt to analyze statistically these data in this paper; such results were given in Boulton (1988).

Study Area

Full details of the study sites were given by Boulton and Smith (1985) and Boulton and Lake (1990). Briefly, two study sites (upstream of the Werribee Picnic Spot (WPS), and Spargo Creek (SC)) were located on the upper reaches of the Werribee River, and two (Fireplace Ford (FF), and Wheeler Road (WR)) on the Lerderderg River. These rivers rise on the southern edge of the Great Dividing Range some 100 km NW of Melbourne, join near Bacchus Marsh and flow south-east to empty into Port Phillip Bay (Fig. 1 in Boulton and Lake, 1990). An annual average rainfall of 950 mm (Smith, 1976) falls on the catchment which is vegetated

by dry sclerophyll forest dominated by Manna Gum (*Eucalyptus viminalis* Labillard). Annual discharge reflects rainfall, peaking in spring, and ceasing in summer almost annually at the Werribee River sites and one year in three at the Lerderderg sites. Mean daily discharge is 24.2 Ml in the Werribee and 27.3 Ml in the Lerderderg River (Australian Water Resources Council, 1979) and highly variable (coefficient of variation of annual flow is 77% and 62% respectively (McMahon, 1979)).

The study commenced in a drought year, 1982. The Werribee River did not flow at all at the upstream site (SC) and only flowed for 5 months (late June to late November) at WPS. The study pool at WPS dried during the ensuing summer. In 1983, flow began in late May and continued for 7.5 months at both sites. At FF and WR, the Lerderderg began to flow in late May, 1982 and ceased in December. At WR, the study pool dried for six weeks that summer whereas at FF it shrank to a moist patch of leaves before refilling in mid-March. Flow resumed in May, 1983 and continued beyond the end of the study in March, 1984.

Methods

Sampling

Physicochemical data were collected each trip, and were discussed elsewhere (Boulton, 1988; Boulton and Lake, 1990). In 1982-1983, five 30-second sweep-net samples were taken in the pools using an FBA pond net (300 μ m mesh). While shuffling vigorously along the bottom of the pool, the pond net was swept from side to side across the disturbed path. When necessary, large wood fragments and cobbles retained in the net were thoroughly rinsed to remove clinging invertebrates before being discarded. Net contents were fixed in 5% formalin.

In adjacent riffles, five samples were collected using a Portable Invertebrate Sampler resembling that illustrated by Merritt and Cummins (1984). This device is a galvanized-iron box whose lower edges are lined with sponge rubber to ensure a seal with the uneven substratum. The upstream face of the box is removable, allowing water currents to flush the box contents into a collecting net (50 μ m mesh) attached to the downstream face. The sampler encloses 0.05 m², exceeding the diameter of most rocks at the sites. The sampling plot was located using random numbers as co-ordinates, the sampler was placed carefully, and the box's contents agitated and scrubbed to a depth of 10 cm while the current washed suspended material and invertebrates into the collecting net. Samples were fixed in 5% formalin.

Often the current was insufficient to flush the box's contents into the collecting net or the water level was too low (< 5 cm) to flow into the box. This meant that the contents had to be manually scooped into the net - an inconsistent and unsatisfactory sampling method remedied by the development of a flow-independent pump sampler described by Boulton (1985). This device was used in 1983-1984 to collect four samples each from pool and riffle habitats at each site except SC where the pool was too deep (> 100 cm) for the sampler. A pilot study based on 40 samples (Boulton, 1988) confirmed that four samples sufficed to provide estimates of the abundances of the 15 most common taxa to within 40% of their means (cf. Elliott, 1977). For comparisons with pool samples collected during 1982, three sweep net samples were taken in each pool during 1983-1984. Fewer samples were collected when pools were smaller than normal (filling or drying) to avoid biases due to habitat disruption or artificial predation. During floods in September, 1983, pump samples could not be taken, necessitating four "kick samples" (Frost et al., 1971) instead.

In 1982, samples were taken approximately monthly (Table 1). As some taxa (e.g., Simuliidae, Chironomidae) can complete their aquatic stages in less than a month at high water temperatures (Chutter, 1968; Oliver, 1971), sampling frequency was increased to fortnightly in 1983-1984 (Table 1) to avoid missing these taxa. Taxonomy of larval Trichoptera and nymphal Ephemeroptera and

Plecoptera was aided by the collection of adult material from light traps and by regular sweep-netting over riparian vegetation.

Laboratory processing

Preserved samples were washed through a nest of sieves with mesh sizes of 2 mm, 1 mm, and 300 μ m. Taxa retained by the two larger meshes were hand-sorted under 12 \times magnification. Organisms retained by the 300 μ m mesh were separated from accompanying organic detritus using the kerosene-ethanol phase technique of Barmuta (1984). Macroinvertebrates were counted and identified as far as practicable using available keys and the voucher collection of the Museum of Victoria; most of the identifications were confirmed or carried out by a variety of experts (see Acknowledgements). Adults and larvae of some taxa (e.g., Elmidae, Hydrophilidae) were treated as separate entities because we were uncertain whether they were conspecific and in many cases they occupied different functional feeding groups.

Each taxon was assigned to a functional feeding group (*sensu* Cummins, 1975; Cummins and Klug, 1979) according to mouth-part morphology and feeding behaviour observed in the laboratory or reported in the literature. Additional information on dietary habits was provided by teasing out the foregut contents of at least 20 specimens of the more common species into polyvinyl lactophenol and comparing them with reference mounts of macerated wood, coarse particulate organic matter (CPOM), fine POM, periphyton, and fine inorganic material. No attempt was made to quantify dietary items (cf. Chessman, 1986). Animals using more than one mode of food collection (e.g., collector-shredder) were treated as separate functional groups because we deemed it unrealistic simply to divide the combination equally between constituent feeding modes (cf. Hawkins and Sedell, 1981). Several taxa were too rare to be unequivocally classified.

Results

A total of 258 taxa was collected from pools and riffles at the four study sites in 1982-1984 (Table 2). Almost a quarter of these were Coleoptera while Diptera comprised just over a third of the total number of taxa (Table 3). Other species-rich groups included aquatic arachnids, Plecoptera, and Trichoptera. Four species of Anura (*Litoria ewingi* (Dumeril and Bibron); Hylidae; *Geocrinia victoriana* (Boulenger), *Neobatrachus sudelli* (Lamb), *Ranidella signifera* (Girard); Myobatrachidae) and two teleost fish (*Galaxias olidus* Günther; Galaxiidae and *Edelia ?obscura* (Klunzinger); Kuhliidae) were also recorded.

Most of the species were predators (Table 4). Collector-scrappers and collector-gatherers were also relatively diverse whereas few taxa were collector-filterers and collector-shredders (Table 4). Most of the individuals (chironomids, oligochaetes, mayfly and stonefly nymphs) collected during this study were collector-gatherers and collector-scrappers.

Discussion

The large number of species of aquatic macroinvertebrates in these intermittent streams has been discussed elsewhere (Boulton and Suter, 1986; Boulton and Lake, 1988) and more recent studies in arid-zone streams in central Australia (Davis et al., in press) suggest that this phenomenon is widespread. The considerable habitat diversity offered by intermittent streams, coupled with the adaptations of many species (e.g., Towns, 1985; Boulton and Lake, 1988) to cope with a long history of aridity is probably responsible for such high species richness in these systems (Lake et al., 1986).

The higher species diversity found in pools (when equivalent numbers of samples were collected using a flow-independent sampling device) differs from the patterns observed in a review of 17 upland permanent streams in North America and the United Kingdom where numbers of taxa were similar between pools and riffles (Logan and Brooker, 1983). The greater species richness probably reflects the importance of pools in the Werribee and Lerderderg Rivers as over-summering refuges (Boulton, 1989).

Many of the species found in late summer are able to tolerate low dissolved oxygen levels or are air-breathing, and most are mobile (e.g., adult dytiscids) and able to escape when the pools dry. Relatively few taxa appear to be obligate inhabitants of intermittent streams; the majority are opportunistic generalists that also occur in nearby permanent streams (Boulton, 1988) or lentic waterbodies (e.g., Hemiptera, Coleoptera).

There are practical difficulties associated with assigning stream invertebrates to functional feeding groups because many of the animals appear to be opportunistic generalists that feed on a wide range of food resources. Diets also tend to change with age. For example, the stonefly *Iliesoperla australis* is a collector-gatherer in its early stages, switching to predation just before its final instars (Yule, 1990). Many taxa possess relatively undifferentiated mouth-parts, richly endowed with brushes and setae that confound attempts to classify them readily into feeding groups. Therefore, generalizations must be cautiously considered.

The high diversity of predators reflects the rich fauna of water mites, dytiscids, hemipterans, odonates, tanypods and other dipterans recorded in this study. The high species richness of predators in receding temporary waters has been reported elsewhere in Australia (Lake et al., 1989) and world-wide (Williams, 1987); densities tend to rise as pools dry up and prey become concentrated (Boulton, 1988; Lake et al., 1989). Most of the individuals collected from the Werribee and Lerderderg Rivers were collector-gatherers and collector-scrappers although these groups were only moderately diverse. This undoubtedly reflects the generalist feeding strategies discussed above, and reiterates the opportunistic nature of many of the lotic macroinvertebrates in these two intermittent streams.

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Table 1. Dates of sampling trips to the study sites during 1982-1984.

Month	1982	1983	1984
January		20	4, 18, 31
February		8	14, 25
March		8, 21, 29	23
April		5, 19, 30	
May	25	10, 24	
June	2-3, 27	7, 22	
July	8, 23	5, 19	
August	22	2, 16, 30	
September	13, 23-24	13, 27	
October	26	11, 25	
November	25	8, 24	
December	19, 23	7, 20	

Table 2. Aquatic macroinvertebrate taxa collected in pools (P) and riffles (R) at the four study sites on the Werribee and Lerderderg Rivers in 1982 ('82) and 1983-84 ('83). The collecting technique (SW = FBA kick-sweep sample, PS = pump sample, BS = box sample) and number of samples are listed below site (SC = Spargo Creek, WPS = Werribee River Picnic Spot, FF = Fireplace Ford, WR = Wheeler Road). Functional feeding groups abbreviated as follows: Cf = Collector-filterers, Cscr = Collector-scrapers, Cg = Collector-gatherers, Cshr = Collector-shredders, Pred = Predators (includes 'piercing' and 'engulfing' predators), Scr = Scrapers, Shr = Shredders, Gen = Generalist feeders, Nf = Nonfeeding stages, Un = Uncertain diet. NMV refers to the National Museum of Victoria voucher number.

[illegible]

[illegible]

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River							
		SC				WPS				FF				WR			
		'83	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'82	'83	'82	'83
		SW	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS
		54	75	58	74	89	28	88	58	75	92	36	92	58	77	93	36
		P	R	P	P	P	R	R	P	P	P	R	R	P	P	P	R
Hygrobatidae																	
<i>Hygrobat</i> sp.	Pred	x	x	x	x	x	x	x		x	x	x	x		x	x	x
<i>Australiobates</i> spp.	Pred	x	x	x	x	x	x	x	x	x	x		x	x	x	x	x
<i>Corticacarus</i> spp.	Pred	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Aspidiobates</i> sp.	Pred	x		x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Gondwanabates</i> sp.	Pred																x
Pionidae																	
<i>Piona ?uncatiformis</i> Lundblad	Pred			x	x		x		x								
Aturidae																	
<i>Frontipodopsis</i> sp.	Pred											x	x				
<i>'Notoaturus'</i> sp.	Pred	x	x		x	x		x	x	x	x	x	x	x		x	x
Arrenuridae																	
<i>Arrenurus</i> sp.	Pred	x			x	x	x										
Mesostigmata																	
Aquatic species 1	Pred				x	x	x		x							x	
Aquatic species 2	Pred	x			x	x	x	x	x	x		x				x	x
Aquatic species 3	Pred				x	x									x	x	
INSECTA																	
Ephemeroptera																	
Baetidae																	
<i>Pseudocloeon</i> sp.	Cscr	x	x		x	x		x					x		x	x	x
Leptophlebiidae																	
<i>Atalophlebia</i> sp. (nr <i>albiterminata/tuhla/hudsoni</i> ¹)	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Nousia</i> spp. (N. nr <i>inconspicua</i> - Werribee R., N. nr <i>fuscata</i> - Lerderderg R. ¹)	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Jappa</i> sp.	Cscr									x	x				x	x	
Odonata																	
Zygoptera																	
Lestidae																	
<i>Austrolestes</i> ?io (Selys)	Pred	x		x	x	x	x	x								x	
Amphipterygidae																	
<i>Diphlebia</i> sp.	Pred			x							x				x		
Anisoptera																	
Aeschnidae																	
<i>Austroaeschna</i> ?parvistigmata																	
Martin	Pred									x		x					

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River									
		SC				WPS				FF					WR				
		'83	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'83
		SW	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	PS
		54	75	58	74	89	28	88	58	75	92	36	92	58	77	93	36	92	
		P	R	P	P	P	R	R	P	P	P	R	R	P	P	P	R	R	
<hr/>																			
Synthemidae																			
<i>Synthemis</i> sp.	Pred	x			x	x		x		x	x						x		
Corduliidae																			
<i>Hemicordulia</i> ? <i>tau</i> Selys	Pred			x	x	x													
Plecoptera																			
Austroperlidae																			
<i>Austropentura victoria</i> Illies	Cshr								x	x	x	x	x	x	x	x	x	x	x
<i>Acruroperla atra</i> (Samal)	Shr								x	x	x		x		x	x	x	x	x
Notonemouridae																			
<i>Austrocercia tasmanica</i> (Tillyard)	Cshr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
Gripopterygidae																			
<i>Illiesoperla australis</i> (Tillyard) ²	Cg	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Newmanoperla thoreyi</i> McLellan	Cscr	x	x																
<i>Leptoperla bifida</i> McLellan	Cscr	x		x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Leptoperla</i> sp.	Cscr	x	x		x	x		x		x	x				x	x			
<i>Riekoperla rugosa</i> (Kimmins)	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>R. tuberculata</i> McLellan	Cscr					x		x			x	x	x		x		x	x	
<i>R. karki-reticulata</i> group ³	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>Trinotoperla montana</i> (Riek) ⁴	Scr				x	x		x	x	x	x	x	x		x	x	x	x	
<i>Dinotoperla fontana</i> Kimmins	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>D. thwaitesi</i> Kimmins	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>D. eucumbene</i> McLellan	Cshr										x	x	x						
<i>D. brevipennis</i> Kimmins	Cscr	x	x		x		x	x		x	x	x	x	x		x	x	x	
Heteroptera																			
Gerridae																			
<i>Rheumatometra philarete</i> Kirkaldy	Pred													x					
Veliidae																			
<i>Microvelia dubia</i> Hale	Pred	x		x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
<i>M. distincta</i> Malipatil	Pred	x		x	x	x	x		x	x				x	x				
Notonectidae																			
<i>Enithares woodwardi</i> Lansbury	Pred	x		x					x	x									
<i>Anisops deanei</i> Brooks	Pred	x		x	x				x	x				x	x				
<i>A. hackeri</i> Brooks	Pred			x	x									x	x				
Corixidae																			
<i>Micronecta annae illiesi</i> Wróblewski	Cg	x	x	x	x	x			x	x	x			x	x	x			x
<i>M. a. tasmanica</i> Wróblewski	Cg				x	x			x						x	x			

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River								
		SC		WPS						FF				WR				
		'83	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83
		SW	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS
		54	75	58	74	89	28	88	58	75	92	36	92	58	77	93	36	92
		P	R	P	P	P	R	R	P	P	P	R	R	P	P	P	R	R
<i>Sigara (Tropocorixa) truncatipala</i> (Hale)	Cg	x		x	x	x			x	x				x	x			
Megaloptera																		
Corydalidae																		
<i>Protochauliodes</i> sp.	Pred													x				
Coleoptera ⁵																		
Noteridae																		
Unidentified larvae (Watts, pers. comm., 4.2.85)	Pred								x									
Dytiscidae																		
<i>Antiporus blakei</i> (Clark)	Pred	x		x	x	x	x		x	x	x		x	x		x		
<i>A. femoralis</i> (Boheman)	Pred	x		x	x	x			x	x	x			x	x	x		
<i>Antiporus</i> spp. larvae	Pred	x		x	x	x	x		x	x	x				x	x		
<i>Australphilus saltus</i> Watts	Pred				x				x	x				x	x			
<i>Australphilus</i> sp. larvae	Pred				x	x		x		x				x	x	x		x
<i>Chostonectes gigas</i> (Boheman)	Pred								x					x	x			
<i>C. johnsoni</i> (Clark)	Pred	x		x	x	x	x		x	x	x	x		x	x	x		
<i>Chostonectes</i> spp. larvae	Pred	x	x	x	x	x	x	x	x	x	x		x		x	x	x	x
<i>Copelatus australiae</i> Clark	Pred	x	x		x		x							x				
<i>Copelatus</i> sp. larvae	Pred	x			x										x			
<i>?Hydaticus</i> sp. larvae	Pred														x	x		
<i>Hyderodes schuckardi</i> Hope	Pred	x				x												
<i>Lancetes lanceolatus</i> (Clark)	Pred	x	x	x	x			x	x		x			x	x	x		
<i>Lancetes</i> sp. larvae	Pred	x	x		x	x	x	x		x	x				x	x	x	x
<i>Liodesuss schuckardi</i> (Clark)	Pred	x		x	x	x	x	x	x	x	x		x	x	x	x	x	x
<i>Megaporus hamatus</i> (Clark)	Pred	x			x	x												
<i>Necterosoma penicillatum</i> (Clark)	Pred	x		x	x	x	x	x	x	x	x			x	x	x	x	x
<i>Necterosoma</i> sp. larvae	Pred	x	x	x	x	x	x	x	x	x	x		x	x	x	x		
<i>Platynectes decempunctatus</i> (Fabricius)	Pred	x	x	x	x	x									x			x
<i>Platynectes</i> sp. larvae	Pred	x	x		x											x		
<i>Rhantus suturalis</i> (Macleay)	Pred	x		x		x			x	x	x			x	x	x		
<i>Sternopriscus hansardi</i> (Clark)	Pred	x		x														
<i>S. multimaculatus</i> (Clark)	Pred									x								
<i>S. mundanus</i> Watts	Pred	x		x	x	x	x		x	x	x	x		x	x	x	x	x
<i>Sternopriscus</i> sp.	Pred															x		
<i>?Eretes</i> sp. larvae	Pred				x	x												
<i>?Rhantus</i> sp. larvae	Pred	x		x	x	x									x	x		
Gyrinidae																x		
<i>Aulonogyrus strigosus</i> Guérin	Pred								x									
<i>Macrogyrus</i> sp.	Pred													x	x			

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River									
		SC				WPS				FF					WR				
		'83	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	
		SW	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	
		54	75	58	74	89	28	88	58	75	92	36	92	58	77	93	36	92	
		P	R	P	P	P	R	R	P	P	P	R	R	P	P	P	R	R	
Gyrinidae sp. larvae	Pred														x	x			
Hydraenidae																			
<i>Hydraena ?ambiflagellata</i>																			
Zwick	Cscr		x			x	x								x	x			
<i>H. luridipennis</i> Macleay	Cscr	x			x	x	x		x	x	x			x	x	x		x	
<i>H. ?tricantha</i> Zwick	Cscr	x		x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	
<i>Hydraena</i> sp.	Cscr		x		x	x							x			x		x	
<i>Ochthebius</i> sp. 1	Cscr			x															
<i>Ochthebius</i> sp. 2	Cscr	x			x				x						x		x		
<i>Ochthebius</i> sp. 3	Cscr	x			x	x		x			x		x				x		
Hydrochidae																			
<i>Hydrochus</i> sp. 1	Cscr				x					x									
<i>Hydrochus</i> sp. 2	Cscr		x											x					
Hydrophilidae																			
<i>Berosus</i> sp.	Cscr	x			x						x		x						
<i>Berosus</i> sp. larvae	Pred				x						x								
<i>Enochrus</i> sp.	Cscr				x									x					
<i>Helochaeres australis</i> Blackburn	Cscr	x			x					x					x				
<i>Limnoxenus zeelandicus</i> (Broun)	Cscr		x		x				x				x						
<i>Paracymus pygmaeus</i> (Macleay)	Cg	x	x	x	x	x		x	x	x	x	x		x	x	x	x		
<i>Paracymus</i> sp. larvae	Pred	x		x	x	x			x	x	x	x	x		x		x		
<i>Paranacaena lindi</i> Blackburn	Cg	x	x	x	x								x	x	x				
Helodidae																			
Helodid sp. 1 larvae	Cg													x		x			
Helodid sp. 2 larvae	Cg	x	x	x	x	x	x		x	x	x	x	x	x	x	x	x	x	
Helodid sp. 3 larvae	Cg									x	x			x	x	x		x	
Helodid sp. 4 larvae	Cg											x			x				
Helodid sp. 5 larvae	Cg				x		x												
?Limnichidae sp.	Cg		x											x					
Psephenidae																			
<i>Sclerocyphon striatus</i>																			
Lea larvae	Scr				x	x	x	x	x	x	x	x	x	x		x	x	x	
Ptilodactylidae sp. larvae	Shr								x	x	x	x	x		x	x	x	x	
Elmidae																			
<i>Austrolimnius (Tiphonaetes)</i>																			
<i>hebrus</i> (Hinton)	Scr		x					x	x	x	x	x	x			x	x	x	
<i>A. (Limnelmis) maro</i> Hinton	Scr							x		x	x	x	x	x	x	x	x	x	
<i>A. (L.) miletus</i> Hinton	Scr										x	x	x			x	x	x	
<i>A. "mormo"</i> larvae	Scr	x			x			x	x	x	x	x	x				x	x	
<i>Austrolimnius</i> sp. larvae	Scr								x	x	x	x	x			x	x	x	

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River									
		SC		WPS						FF					WR				
		'83	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'83
		SW	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	PS
		54	75	58	74	89	28	88	58	75	92	36	92	58	77	93	36	92	
		P	R	P	P	P	R	R	P	P	P	R	R	P	P	P	R	R	
<hr/>																			
<i>Notriolus quadriplagiatus</i> Carter	Cscr		x	x	x	x	x	x			x	x	x						
<i>N. quadriplagiatus</i> larvae	Cscr			x	x	x		x											
<i>Simsonia tasmanica</i> (Blackburn)	Cscr								x	x	x	x	x						
<i>S. tasmanica</i> larvae	Cscr								x	x	x	x	x					x	
<i>Kingolus</i> sp. larvae (NMV sp. I)	Cscr								x							x			
<hr/>																			
Mecoptera																			
Nannochoristidae																			
<i>Nannochorista</i> sp.	Pred												x						
<hr/>																			
Diptera																			
Tipulidae																			
<i>Cryptolabis</i> sp. (nr NMV sp. 4)	Cg													x	x	x			x
<i>Ctenophora</i> sp.	Shr	x	x								x							x	
<i>Dactylolabis</i> sp.	Pred										x		x						
<i>Hexatoma</i> nr <i>megacera</i>																			
(nr NMV sp. 39)	Pred		x	x	x	x		x											
<i>Hexatoma</i> sp. 1 (nr NMV sp. 24)	Pred			x	x									x	x	x			
<i>Hexatoma</i> sp. 2 (nr NMV sp. 45)	Pred		x										x						
<i>Limnophila</i> sp. 1 (nr NMV sp. 1)	Pred									x						x			
<i>Limnophila</i> sp. 2	Pred		x	x	x	x	x		x	x	x	x		x		x	x	x	
<i>Limonia</i> sp. 1 (NMV sp. 3)	Cg		x		x			x		x			x		x	x			x
<i>Limonia</i> sp. 2	Cg			x			x				x	x	x			x	x	x	
<i>Ormosia</i> sp. (NMV sp. 5)	Cshr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	
<i>Oropeza</i> sp. 1	Shr												x						
<i>Oropeza</i> sp. 2	Shr		x		x	x	x	x							x	x			
<i>Tipula</i> sp. 1	Cg					x		x											x
<i>Tipula</i> sp. 2	Pred	x	x		x	x	x	x			x	x	x	x	x	x	x	x	
Tipulidae sp. 1	Cg									x			x						
Tipulidae sp. 2	Pred						x	x											
Tipulidae sp. 3	Cg																x	x	x
<hr/>																			
Psychodidae																			
<i>Maruina</i> sp.	Cscr			x		x						x	x			x			x
<i>Psychoda</i> sp. (NMV sp. 3)	Cg	x	x		x	x	x	x	x	x	x	x	x		x	x	x		
<hr/>																			
Dixidae																			
<i>Dixa</i> sp. (NMV sp. 1)	Cg				x	x	x			x	x	x		x	x	x			x
<hr/>																			
Culicidae																			
<i>Aedes</i> sp. 1	Cf	x		x	x	x			x	x	x			x	x	x			x
<i>Aedes</i> sp. 2	Cf	x			x	x			x	x	x			x	x	x			
<i>Anopheles annulipes</i> Walker	Cf	x		x	x					x		x		x	x	x			
<i>Culex australicus</i> Skuse	Cf	x		x					x	x					x	x			
<i>C. annulirostris</i> Skuse	Cf	x		x			x			x				x	x	x			
<i>C. fatigans</i> Weidmann	Cf				x					x	x			x	x	x			

[illegible]

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River								
		SC				WPS				FF				WR				
		'83	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83	'82	'83	'83	'82	'83
		SW	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS	SW	SW	PS	BS	PS
		54	75	58	74	89	28	88	58	75	92	36	92	58	77	93	36	92
		P	R	P	P	P	R	R	P	P	P	R	R	P	P	P	R	R
<i>Cricotopus</i> complex																		
(includes NMV spp. 160E, 12E) ⁶																		
	Cscr	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
nr <i>Brillia</i> sp. 1	Cg	x			x	x	x	x	x	x	x	x	x	x	x	x	x	x
nr <i>Brillia</i> sp. 2	Cg	x	x	x	x	x	x	x	x	x	x	x			x	x	x	x
<i>Heterotrissocladius</i> sp.	Cg	x	x	x	x	x	x	x	x	x	x		x		x	x	x	x
<i>Corynoneura</i> sp.	Cscr								x	x						x		
<i>Stictocladius</i> sp. 1 (NMV sp. 9E)	Cscr		x			x	x	x		x	x	x	x			x	x	x
nr <i>Cordites</i> sp.	Cscr	x	x		x		x	x		x	x	x	x		x	x		x
<i>Rheocricotopus</i> sp.																		
(NMV sp. 91E)	Cg								x	x					x			
<i>Thienemanniella</i> sp.																		
(NMV sp. 10E)	Cg						x											
<i>Orthoclaudiinae</i> sp. 1	Cscr	x	x		x	x	x	x		x	x			x	x	x		
<i>Orthoclaudiinae</i> sp. 2	Cscr	x															x	x
<i>Orthoclaudiinae</i> sp. 3	Cscr		x															
<i>Orthoclaudiinae</i> sp. 4	Cscr																x	
<i>Orthoclaudiinae</i> sp. 5	Cscr		x						x									
<i>Orthoclaudiinae</i> sp. 6																		
(NMV sp. 34E)	Cg			x	x	x	x		x	x	x	x		x	x	x		
Aphroteniinae																		
<i>Aphroteniella</i> sp.(nr NMV sp. 18E)	Pred									x	x	x	x	x	x	x	x	x
Ceratopogonidae																		
<i>Atrichopogon</i> sp.	Cscr	x	x		x	x		x							x			x
<i>Bezzia</i> sp.	Pred	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x		x
<i>Culicoides</i> sp. 1	Pred	x	x			x		x										
<i>Culicoides</i> sp. 2	Pred							x										
<i>Dasyhelea</i> sp.	Cg		x	x	x	x	x	x		x			x			x		x
<i>Forcipomyia</i> sp.	Cscr	x	x	x	x			x	x	x	x		x					
<i>Monohela</i> sp.	Pred		x	x	x	x	x	x						x	x		x	
<i>Nilobezzia</i> sp.	Pred		x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x
Simuliidae																		
All taxa listed below were "lumped" in this study																		
<i>Austrosimulium furiosum</i> (Skuse)																		
<i>A. victoriae</i> (Roubaud)																		
<i>Austrosimulium</i> nr sp. Y ⁷																		
<i>Austrosimulium</i> sp. (nr <i>victoriae</i>)																		
? " <i>Cnephia</i> " <i>umbratorum</i> ⁸	Cf	x	x	x	x	x	x	x	x	x	x	x	x		x	x	x	x
Thaumaleidae																		
<i>Thaumaleidae</i> sp.	Scr			x	x	x			x	x								x

Taxa	Functional Feeding Group	Werribee River								Lerderderg River									
		SC				WPS				FF					WR				
		'83 SW 54 P	'83 PS 75 R	'82 SW 58 P	'83 SW 74 P	'83 PS 89 P	'82 BS 28 R	'83 PS 88 R	'82 SW 58 P	'83 SW 75 P	'83 PS 92 P	'82 BS 36 R	'83 PS 92 R	'82 SW 58 P	'83 SW 77 P	'83 PS 93 P	'82 BS 36 R	'83 PS 92 R	
Tabanidae																			
Tabanidae sp. 1	Pred											x	x					x	
Tabanidae sp. 2	Pred											x					x		
Stratiomyidae																			
Stratiomyidae spp.	Cg	x	x		x	x	x	x		x		x	x		x	x		x	
Empididae																			
Empididae sp. 1	Pred	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	
Empididae sp. 2	Pred	x	x		x		x	x	x	x	x	x	x	x	x	x	x	x	
Dolichopodidae																			
Dolichopodidae sp.	Pred	x	x		x	x		x		x			x	x	x	x		x	
Sciomyzidae																			
Sciomyzidae sp.	Pred			x		x									x				
Ephydriidae																			
Ephydriidae sp.	Cg						x												
Muscidae																			
Muscidae sp. 1	Pred	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	x	
Muscidae sp. 2	Pred					x	x	x	x	x	x				x	x		x	
Muscidae sp. 3	Pred	x	x		x	x	x	x	x	x	x	x	x	x	x	x		x	
Trichoptera																			
Hydrobiosidae																			
<i>Apsilochorema gisburni</i> (Mosely)	Pred		x		x			x			x		x			x		x	
<i>A. obliquum</i> (Mosely)	Pred								x		x	x	x			x	x	x	
<i>Ptychobiosis nigrata</i> (Banks)	Pred									x	x	x	x		x	x	x	x	
<i>Taschorema</i> sp. (?evansi)	Pred		x		x	x	x	x		x	x	x	x			x	x	x	
Glossosomatidae																			
<i>Agapetus</i> spp. (includes sp. 7 ⁹)	Scr		x				x	x		x	x	x	x		x	x	x	x	
Hydroptilidae																			
<i>Hellyethira ?simplex</i> (Mosely)	Scr	x	x	x	x	x	x	x		x	x	x	x	x	x	x		x	
<i>Maydenoptila</i> sp.	Scr	x			x	x		x											
<i>Oxyethira columba</i> (Neboiss)	Scr					x		x											
Philopotamidae																			
<i>Hydrobiosella</i> sp.	Cf												x						
Hydropsychidae																			
<i>Cheumatopsyche</i> sp. 3 ⁹																			

Table 2 (continued)

Taxa	Functional Feeding Group	Werribee River								Lerderderg River								
		SC				WPS				FF					WR			
		'83 SW 54 P	'83 PS 75 R	'82 SW 58 P	'83 SW 74 P	'83 PS 89 P	'82 BS 28 R	'83 PS 88 R	'82 SW 58 P	'83 SW 75 P	'83 PS 92 P	'82 BS 36 R	'83 PS 92 R	'82 SW 58 P	'83 SW 77 P	'83 PS 93 P	'82 BS 36 R	'83 PS 92 R
Calocidae																		
<i>Coenota plicata</i> Mosely	Shr								x	x		x			x		x	
Calocidae sp. 1	Shr					x		x	x	x	x	x	x	x	x	x	x	
Atriplectididae																		
<i>Atriplectides dubius</i> Mosely	Cg									x	x		x				x	
Philorheithridae																		
<i>Austrheithrus</i> sp.	Pred										x					x		
Helicopsychidae																		
? <i>Helicopsyche</i> sp.	Scr			x			x		x	x	x	x	x	x	x	x	x	
Calamoceratidae																		
<i>Anisocentropus</i> sp.	Shr													x				
Leptoceridae																		
<i>Leptorussa darlingtoni</i> (Banks)	Cscr	x		x					x	x	x		x	x	x	x	x	
<i>Oecetis</i> sp.	Pred	x		x	x	x	x		x	x	x	x	x	x	x	x	x	
<i>Triaenodes</i> sp. (NMV sp. PT773)	Shr			x	x	x	x											
<i>Lectrides varians</i> Mosely	Shr			x					x	x	x		x	x	x			
<i>Notalina bifaria</i> Neboiss	Cshr				x											x		
<i>N. spira</i> St Clair	Cshr					x											x	
<i>Triplectides ciuskus ciuskus</i> Mosely	Shr			x					x	x	x		x	x	x		x	
<i>T. similis</i> Mosely	Shr			x	x		x		x	x	x		x	x	x	x	x	
<i>T. truncatus</i> Neboiss	Shr								x	x	x	x		x	x	x	x	
Lepidoptera																		
Pyralidae sp.	Shr	x	x	x	x	x				x							x	

¹ Dr P.J. Suter (pers. comm. 17.7.85) believes that the species of *Atalophlebia* and the species of *Nousia* is probably new because the adult genitalia are quite distinctive. They most resemble those named species in parentheses. *N. inconspicua* is now named *Koorrnonga inconspicua* (Dr R. Marchant, pers. comm. 18.4.91).

² Mr G. Theischinger (pers. comm. 14.5.85) identified these specimens as *Illiesoperla* (*Illiesoperla*) *mayi* (Perkins), a species earlier synonymized with *I. australis* by Hynes (1974). Theischinger (1982) reinstated *I. mayi* as a species clearly distinct from *I. australis* and illustrated some of the larval features (Theischinger 1984) associated with the adult after breeding out. The validity of the specific status of *I. mayi* is in doubt (C. M. Yule, pers. comm.) and a conservative approach, retaining the original name *Illiesoperla australis*, has been adopted here.

- ³ Of the four species that comprise this complex (Hynes 1978), only *Riekoperla williamsi* McLellan occurred in collections of adults examined by Mr G. Theischinger (pers. comm., 14.5.85). Other adults, that could not be matched with nymphs, were *R. naso* Theischinger and *R. triloba triloba* McLellan and it is likely that the nymphs of these species were included in counts of nymphs assigned to the *R. karki-reticulata* complex. Difficulty was encountered separating some *R. rugosa* nymphs from those of the *R. karki-reticulata* complex. These may have been the nymphs of *R. naso*, a species found co-occurring with the closely related *R. rugosa* for the first time in Australia in this study (G. Theischinger pers. comm. 14.5.85).
- ⁴ Ms C.M. Yule (pers. comm. 18.2.84) states that this is the correct name for adults and nymphs previously assigned to *Trinotoperla yeoi* Perkins.
- ⁵ Unless otherwise indicated, all these beetles were adults.
- ⁶ The *Cricotopus* complex is made up of at least seven different chironomid species whose characteristics are difficult to distinguish, especially when the mouth-parts are worn. As they all appear to share a similar diet, they have been lumped in this study.
- ⁷ *Austrosimulium* nr sp. Y follows the name used in Ms J. Prince's collection.
- ⁸ "*Cnephia*" definitely is not in the genus *Cnephia* although that is the name currently used (Ms J. Prince pers. comm. 7.8.84).
- ⁹ These numbers refer to the voucher specimens held in the MMBW collection of Mr J. Dean and Mr D. Cartwright.

Table 3. Numbers and percentage composition of aquatic macroinvertebrate taxa in major groups collected from pools and riffles at four study sites on the Werribee and Lerderg Rivers in 1982–1984.

Taxon	Total	Percentage
Mollusca	6	2.32
Other non-arthropods	7	2.71
Crustacea	9	3.49
Arachnida	17	6.59
Ephemeroptera	5	1.94
Odonata	5	1.94
Plecoptera	15	5.81
Hemiptera	9	3.49
Coleoptera	64	24.81
Non-chironomid Diptera	52	20.16
Chironomidae	37	14.34
Trichoptera	29	11.24
Other insects	3	1.16
Total	258	

Table 4. Numbers and percentage composition of taxa in several functional feeding groups.

Taxon	Total	Percentage
Predators	101	39.15
Collector-scrappers	49	18.99
Collector-gatherers	43	16.67
Scrappers	17	6.59
Shredders	17	6.59
Collectors-filterers	15	5.81
Collector-shredders	12	4.65
Others	4	1.55

Plecoptera, Ephemeroptera and Trichoptera of the Pelion Valley, Tasmanian World Heritage Area

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Abstract. Dean, J.C. and Cartwright, D.I. 1992. Plecoptera, Ephemeroptera and Trichoptera of the Pelion Valley, Tasmanian World Heritage Area. *Occasional Papers from the Museum of Victoria* 5: 73-79.

A survey of the Pelion Valley yielded eighty species of aquatic insects, consisting of seventeen species of stoneflies, thirteen species of mayflies and fifty species of caddisflies. Most species featured orderly distribution patterns along the river continuum, and several examples of spatial separation of congeners are reported. There was an obvious trend of increasing species richness as one moved from small headwater streams to larger downstream sites. The Lake Ayr outlet stream was unique in yielding a number of species which are widely distributed at lower altitudes, and their presence in the study area can probably be attributed to benign physical conditions at that site. There was no evidence of endemism among the aquatic insects we collected. With a single exception, there are extensive records from elsewhere in Tasmania of all the species we have identified. The exception is the stonefly *Eusthenia reticulata*, which has seldom been collected and is apparently a rare species (Hynes, 1989). Its collection from Douglas Creek is significant. The status of species we are unable to identify is unknown. These include most of the mayflies and some caddisflies, and highlights the need for basic taxonomic investigations of these groups.

Introduction

Running water communities worldwide are usually dominated by aquatic insects. In terms of species richness and abundance, the Orders Plecoptera, Diptera, Ephemeroptera and Trichoptera are prominent. This report presents the results of a survey of three of these: Orders Plecoptera (stoneflies), Ephemeroptera (mayflies) and Trichoptera (caddisflies), in the Mount Ossa - Pelion Valley area of the Cradle Mountain - Lake St Clair National Park.

Cradle Mountain - Lake St Clair National Park is an extensive reserve which forms the northern boundary of the Tasmanian World Heritage Area. Access to most of the country within the Park is relatively difficult, and the area has not previously been surveyed for aquatic insects. Neboiss et al. (1988) have reported on the caddisflies of the extreme northern and southern ends of the Park, but pointed out that "the greatest and most exciting part of the Park from Cradle Mountain to Lake St Clair still remains unexplored as far as the aquatic insects (Trichoptera) are concerned". Limited collections of Plecoptera and Ephemeroptera have also been made in accessible parts of the Park, especially near Cradle Mountain. These collections have been far from comprehensive, and have only been reported as locality records for a few species in the taxonomic literature.

The results presented below are based on a series of

samples collected in mid-January 1990. Limited time was available for sampling at each location and, although taxa lists are extensive, it is likely that they include only the more common species from the survey area. In addition, species with life histories characterised by an absence of nymphs or larvae from streams in summer are unlikely to have been collected.

Methods

Sampling

Samples were collected from seventeen locations (Figure 1). With the exceptions of sites 1 and 6, these were all running water sites, ranging from first order headwater streams to larger third order streams (see Appendix).

Aquatic stages were primarily collected using a kick sampling technique. Bottom sediments were vigorously disturbed, either by foot or by overturning larger rocks and woody substrates. The dislodged fine material was collected in a downstream pond net, transferred to a large white tray, and individual insects picked from the debris using forceps. Larvae and pupae were also handpicked directly from the surfaces of larger rocks and solid substrates.

Adults were collected by sweeping riparian vegetation with hand nets. Battery operated light traps were employed after dark, although with limited success.

Taxonomy

The Tasmanian stoneflies are taxonomically well known. Material collected during the present study has been identified using keys to adults and nymphs (Hynes, 1989).

By contrast, the taxonomy of Tasmanian mayflies is very incomplete. Adults of fifteen species have been described from the state (Tillyard, 1936; Harker, 1954, 1957), but descriptions of many of these are inadequate for identification of collected material. This problem is compounded by the presence of an unknown number of undescribed species. Although taxonomy of the nymphs is an even more parlous state, we have concentrated our efforts on this stage. All specimens have been identified to genus using the keys of Suter (1979) and Dean (1989), and we have allocated voucher numbers to distinguish presumptive species. A voucher collection has been maintained for several years in Victoria, and the Tasmanian material has now been incorporated into this. Formal identification must, however, await a revision of the adults and the rearing of nymphs.

While adult taxonomy of the Tasmanian Trichoptera is well advanced, primarily due to the efforts of Neboiss (1977, 1986), little published information is available on immature stages. Despite this, we have been able to identify most of the larvae collected. Over the past ten years, we have accumulated a large amount of larval and reared material from both Tasmania and the mainland, and this unpublished information has been augmented with reared material collected during the present study. Larval taxonomy of the families Conoesucidae, Calocidae and Helicophidae is currently being investigated by Jean Jackson at the University of Tasmania, and she has kindly identified some of our material in these families.

Distribution and abundance

Included in the species lists are assessments of the distribution and abundance of each taxa. These assessments are based solely on the set of samples which we have collected, and give an indication of relative rather than absolute abundance. The status of individual taxa would be likely to change if sampling intensity was increased, or if samples were collected at other times of the year. Categories we have adopted are:

Restricted distribution: recorded from 1 or 2 sites only.

Limited distribution: recorded from 3 or 4 sites.

Widespread distribution: recorded from 5 or more sites.

Rare species: a total of 1-3 specimens collected from all sites.

Common species: a total of 4-30 specimens collected from all sites.

Abundant species: more than 30 specimens collected from all sites.

Results and Discussion

Plecoptera (stoneflies)

The species. — At least seventeen stonefly species were recorded from the study area (Table 1). While most of the material consisted of nymphs, adults of eleven species were also collected, allowing confirmation of nymphal identifications.

Species richness ranged from 1 to 7 at the running water sites, but there was no obvious relationship between numbers of species and stream size. Similarly, there were few well defined patterns of distribution. Several species were widely distributed (e.g., *Eusthenia costalis*, *Tasmanoperla thalia* and *Cryptoperla paradoxa*), but the majority were sporadic in occurrence, with at least eleven species recorded from only 1 or 2 sites. This makes it difficult to draw conclusions about general distribution.

Notes on selected taxa. — *Eusthenia reticulata*. This species is apparently rare. Hynes (1989) comments that it has only been collected once in recent years, from a high stream on Mount McCall. The nymph is unknown, although possible nymphs from Mount McCall are grouped with two additional species as the *Eusthenia spectabilis* group in the key of Hynes (1989). In the present study a single female was collected from site 4, while four nymphs of the *E. spectabilis* group listed in Table 1 may be nymphs of *E. reticulata*.

Leptoperla beroe. A single male of this species was collected from the shore of Lake Ayr: the nymphs probably occur in the lake, but this has not been confirmed.

Cardioperla nigrifrons. Large numbers of nymphs of this species were collected from moss in swift water on the 45° face of a waterfall at site 12.

Cardioperla sp. A. A single, very early nymph of a third species of *Cardioperla* was collected from site 5. This nymph had a very strong dorsal ridge on the abdominal terga, but was too small for positive identification. Sampling at other times of the year may yield additional specimens.

Genus *Austrocercoides*. Nymphs of the two known Tasmanian species cannot be distinguished. Adults of *A. zwicki* emerge in spring-early summer, while *A. bullata* adults have been collected from January to April (Hynes, 1989). In the present study adults of *A. bullata* were collected from site 4, while fully developed nymphs from sites 4 and 7 were probably the same species. Nymphs less than half grown were collected from sites 8, 14, 15 and 16, and these could conceivably be *A. zwicki*.

Ephemeroptera (mayflies)

The species. — Thirteen mayfly species were collected, all but three belonging to the family Leptophlebiidae (Table 2). At least four, and perhaps as many as ten of these species are undescribed, reflecting the great need for taxonomic study of Tasmanian Ephemeroptera.

Distribution patterns. — Arrangement of the running water sites in order of increasing size highlights the fact that most species featured orderly, well defined patterns of distribution (Table 3). *Ameletoides* sp. was widely distributed in fast flowing habitats of moderate size streams, while *Tasmanophlebia lucustris* avoids currents and was only collected from Lake Ayr and those few running water sites with pools and areas of slow current (sites 2, 4 and 14). Within the genus *Nousia*, *Nousia* sp. 7 was extremely widespread and recorded from most stream sites, *Nousia* sp. 6 was restricted to intermediate size tributary streams in heavy forest, while *Nousia* sp. 5 and *Nousia* sp. 9 were only recorded from larger downstream sites. *Nousia* sp. 8 was restricted to flowing water sites above and below Lake Ayr. The two species of *Austrophlebioides* were also spatially separated. *Austrophlebioides* sp. 4 was restricted to open sites on the Pelion Plain, while *Austrophlebioides* sp. 5 was only recorded from heavily forested sites further upstream.

There was an obvious tendency for species richness to increase as stream size increased (Table 3). The number of species at individual sites ranged from one at site 8 (a small headwater stream) to seven species at the furthest downstream site (site 17).

Notes on selected taxa. — *Ameletoides* sp. This genus was first recorded from Tasmania by Campbell (1981), who collected nymphs from Pencil Pine River near Cradle Mountain. He suggested that they may have been an undescribed species, and comparison of nymphs collected in the present study with specimens from the Victorian Alps supports this suggestion.

Atalophlebia albiterminata. This species is common throughout much of Tasmania (Tillyard, 1936). In the present study it was only collected from the Lake Ayr outlet stream, 100 metres downstream from the lake. Its presence at this site may be a consequence of a reduced flow and stable substrate, or perhaps is due to slightly elevated temperatures.

Genus *Nousia*. With six species, the genus *Nousia* is extremely diverse in the study area. This diversity is apparently facilitated by spatial separation of nymphs. Four species in the genus have previously been described from Tasmania, three of them only from lowland localities. Identification of the Pelion material is not possible at present, despite the collection of adults of several of the species.

Genus *D* sp. *tristis*. This species was described by Harker (1954) (as *Jappa tristis*), the type material having been collected by Tillyard in 1917 from Cradle Mountain. In a guide to Australian Leptophlebiidae, Dean (1989) established "Genus D" to accommodate several undescribed species from south-eastern Australia. Examination of nymphs and an adult male from Pelion Valley confirms that the species *tristis* belongs to this undescribed genus. The genus *Jappa* is therefore removed from the Tasmanian fauna.

Genus *Austrophlebioides*. This is the first record of this genus from Tasmania, and both species are undescribed. *Austrophlebioides* sp. 5 is characterised by a striking modification of the labrum. The fringes of setae on the anterior margin have been formed into a "suction disc" which perhaps assists the nymph when clinging to rock surfaces in fast currents. In all other aspects the nymph agrees with typical *Austrophlebioides* nymphs.

Trichoptera (caddisflies)

The species. — At least fifty species of caddisflies were recorded from the study area (Table 4). The dominant families were Hydrobiosidae (14 species), Leptoceridae (8 species), Conoesucidae (5 species) and Philorheithridae (4 species). Of the fifty species, 38 were collected as either adults or larvae which have been reared to adults, and could be identified to the species level.

Distribution patterns. — Distributions of those species which were recorded from more than one site are presented in Table 5. As with the mayflies, there was a general tendency for species richness to increase with increasing stream size.

Most of the running water species featured distribution patterns which were orderly with respect to position along the river continuum. For example, *Dipletrona* sp. was limited to small-medium size forest streams, *Hydrobiosella cognata*, *Archaeophylax ochreus*, *Caenota plicata* and *Tasmanthrus galbinomaculatus* were restricted to medium size streams but extended further downstream than *Dipletrona* sp., while *Apsilochorema obliquum*, *Taschorema asmanum* and *Conoesucus digitiferus* ranged from medium size forest streams to the furthest downstream sites, and *Taschorema viridarium*, *Hydrobiosella waddama* and *Plectrocnemia* sp. were restricted to the larger downstream sites.

Taschorema evansi, *Helicopsyche* sp. and *Tamasia variegata* were only recorded from the Lake Ayr outlet stream. In addition, *Asmicridea edwardsi* may be restricted to the outlet stream, as the only other record (site 4) was a single adult male which may have flown downstream from site 3. All of these species are widely distributed at lower altitudes, and physical conditions unique to site 3 probably explains their presence in the study area.

The leptocerid *Triplectidina nigricornis* was only recorded from site 6, which consisted of a series of small isolated pools on Pelion Plain, and was the only species of caddisfly collected from these pools.

Spatial separation of congeners was a striking feature of the genera *Tasmanthrus* and *Hydrobiosella*. *Hydrobiosella waddama* was only recorded from open sites on the Pelion Plain, whereas *H. cognata* appears to be restricted to forest sites further upstream. A single adult male of *H. cognata* collected at site 17 probably flew down from an upstream site. Similarly, *Tasmanthrus angustipennis* was restricted to the three sites downstream from Lake Ayr, while *T. galbinomaculatus* was recorded from upstream forested sites.

Notes on selected taxa. — *Moruya* sp. Larvae of the genus *Moruya* were collected from many sites, but we are unable to identify them to species. At least some specimens are probably *M. charadra*, adults of which were collected from site 17, but it is likely an additional species is also represented in the samples.

Hydrobiosidae: larval species A, B and C. These species were each recorded from a single site, and only one or two specimens were collected. It is likely that at least two of them are species of *Austrochorema*, while a third may be *Ipsebiosis*. Until adults have been reared, they must remain unidentified.

Genus *Tasmanthrus*. Male pupae and associated larval sclerites were collected, as well as numerous larvae, of two distinctive species of *Tasmanthrus*. One species is clearly *T. angustipennis*, while the male genitalia of the second species agrees with *T. galbinomaculatus* described from Cradle Mountain by Jacquemart (1965). Neboiss (1977) has placed the latter species as a synonym of *T. angustipennis*, but this decision will probably have to be reversed.

General comments

The present survey has yielded 80 species of aquatic insects, consisting of 17 species of stoneflies, 13 species of mayflies and 50 species of caddisflies. Species richness varied considerably between individual sites. At running water sites total numbers of species ranged from eight (site 9) to 28 (site 17), with an obvious trend of increasing richness as one moved from small headwater streams to larger downstream sites.

We found no evidence of endemism in the aquatic insects of Pelion Valley. While the Tasmanian mayflies are poorly known, and as a consequence we are unable to assess the wider distribution of the species we collected, Tasmanian stoneflies and caddisflies have been extensively surveyed in recent years. With the exception of *Eusthenia reticulata*, all the stonefly species from Pelion Valley are widely distributed in Tasmania (Hynes, 1989). *E. reticulata* is a species which has rarely been collected, and its presence in Douglas Creek is of significance. Similarly, the caddisfly species which we have identified are well known from elsewhere in the state (Neboiss et al., 1988). The twelve taxa which we are unable to identify were all collected as larvae only, and there is no reason to suggest that any of these are endemic or rare when none of the 38 species which we were able to identify fall into either of these categories.

The survey has highlighted deficiencies in the taxonomy of adults and nymphs of Tasmanian mayflies, and the larval taxonomy of some of the caddisflies. Aquatic insect communities are valuable indicators of general environmental quality, and are responsive to catchment disturbance and changes in such factors as hydrology, temperature and siltation. However, if they are to be fully utilised in environmental monitoring programs, it is important that basic taxonomic information is available, especially for immature stages.

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Table 1. Plecoptera (stoneflies) of Pelion Valley

	Abundance/distribution	Sites
EUSTHENIIDAE		
<i>Eusthenia costalis</i>	Abundant, widespread	7-11, 13-17
<i>E. reticulata</i> (A)	Rare, restricted	4
<i>E. spectabilis</i> group (N)	Common, restricted	3, 11
AUSTROPERLIDAE		
<i>Tasmanoperla thalia</i>	Abundant, widespread	7,8,10,11,13-16
<i>Crypturoperla paradoxa</i>	Abundant, widespread	8-15
GRIPOPTERYGIDAE		
<i>Leptoperla varia</i>	Common, restricted	2
<i>L. beroe</i>	Rare, restricted	1
<i>Cardioperla nigrifrons</i>	Common, limited	8, 10-12
<i>C. falsa</i>	Common, restricted	10
<i>C. sp. A</i>	Rare, restricted	5
<i>Dinotoperla serricauda</i>	Rare, restricted	17
<i>Trinotoperla zwicki</i>	Common, limited	7, 10, 11, 17
<i>T. tasmanica</i>	Rare, restricted	11
NOTONEMOURIDAE		
<i>Notonemoura lynchi</i>	Rare, restricted	8, 16
<i>Austrocerca tasmanica</i>	Rare, restricted	1
<i>A. rieki</i>	Rare, restricted	2, 3
<i>Austrocercella christinae</i>	Rare, restricted	9
<i>Austrocercoides bullata</i> (A)	Rare, restricted	4
<i>A. sp. (N)</i>	Common, widespread	4, 7, 8, 14-16

A = adults, N = nymphs

Table 2. Ephemeroptera (mayflies) of Pelion Valley

	Abundance/distribution	Sites
SIPHONURIDAE		
<i>Ameletoides</i> sp.	Common, widespread	4,7,9-11,14,15
ONISCIGASTRIDAE		
<i>Tasmanophlebia lacustris</i>	Common, limited	1,2,4,14
BAETIDAE		
<i>Baetis</i> sp. A	Common, restricted	5,17
LEPTOPHLEBIIDAE		
<i>Atalophlebia albiterminata</i>	Rare, restricted	3
<i>Nousia</i> sp.5	Abundant, widespread	4,7,14,15,17
<i>Nousia</i> sp.6	Common, widespread	9-11, 15, 16
<i>Nousia</i> sp.7	Abundant, widespread	2,4,7-11,13-17
<i>Nousia</i> sp.8	Common, limited	2,3,5,17
<i>Nousia</i> sp.9	Common, limited	7,14,17
<i>Nousia</i> sp.10	Rare, restricted	13
Genus D sp. <i>tristis</i>	Abundant, widespread	4,7,11,14,15,17
<i>Austrophlebioides</i> sp.4	Common, restricted	5,17
<i>Austrophlebioides</i> sp.5	Common, limited	7,10,11,15

TABLE 3 Distribution of mayfly species, Pelion Valley

	Running water sites increasing catchment size →												Lake outlet sites		Moss face of waterfall	Standing waters	
	2	8	13	9	16	10	15	11	14	7	4	17	5	3	12	1	6
<i>Ameletoides</i> sp.				X		X	X	X	X	X	X						
<i>Tasmanophlebia lacustris</i>	X								X		X						X
<i>Baetis</i> sp.												X		X			
<i>Atalophlebia albiterminata</i>														X			
<i>Nousia</i> sp. 5							X		X	X	X	X					
<i>Nousia</i> sp. 6				X	X	X	X	X									
<i>Nousia</i> sp. 7	X	X	X	X	X	X	X	X	X	X	X	X					
<i>Nousia</i> sp. 8	X											X	X	X			
<i>Nousia</i> sp. 9									X	X		X					
<i>Nousia</i> sp. 10			X														
Genus D sp. <i>tristis</i>							X	X	X	X	X	X					
<i>Austrophlebioides</i> sp. 4												X		X			
<i>Austrophlebioides</i> sp. 5						X	X	X		X							
Number of species	3	1	2	3	2	4	6	5	6	6	5	7	3	2	-	1	-

Table 4. Trichoptera (caddisflies) of Pelion Valley

	Abundance/distribution	Sites
HYDROBIOSIDAE		
<i>Apilochorema obliquum</i>	Abundant, widespread	4,5,7,10,14,15,17
<i>Austrochorema pegidion</i>	Common, limited	7,8,10,13
<i>Ulmerochorema</i> sp.	Common, limited	4,5,7,17
<i>Ethochorema nesydrion</i>	Abundant, widespread	4,7,10,11,14-17
<i>Ethochorema kelion</i>	Rare, restricted	15
<i>Taschorema asmanum</i>	Abundant, widespread	4,5,7,12,14,15,17
<i>Taschorema evansi</i>	Common, restricted	3, 5
<i>Taschorema viridarium</i>	Common, restricted	4, 17
<i>Ptychobiosis nigrata</i>	Rare, restricted	15
<i>Keotonga clivicola</i>	Rare, restricted	14
<i>Moruya charadra</i>	Rare, restricted	17
<i>Moruya</i> sp.	Common, widespread	4,7,10,12,14,15,17
Unident. larva sp.A	Rare, restricted	2
Unident. larva sp.B	Rare, restricted	12
Unident. larva sp.C	Rare, restricted	12
GLOSSOSOMATIDAE		
<i>Agapetus tasmanicus</i>	Rare, restricted	5
PHILOPOTAMIDAE		
<i>Hydrobiosella cognata</i>	Abundant, widespread	7,9-17
<i>Hydrobiosella waddama</i>	Abundant, restricted	5,17
HYDROPSYCHIDAE		
<i>Smicrophylax</i> sp.	Abundant, restricted	17
<i>Asmicridea edwardsi</i>	Abundant, restricted	3,4
<i>Dipletrona</i> sp	Abundant, widespread	8,10,11,13,15,16
POLYCENTROPODIDAE		
<i>Plectocnemia</i> sp.	Rare, restricted	4,7
<i>Tasmanoplegus spilota</i>	Rare, restricted	2
<i>Nyctiophylax repandus</i>	Common, limited	7,14,17

Table 4 continued.

LIMNEPHILIDAE		
<i>Archaeophylax ochreus</i>	Common, widespread	7,10,11,13-16
PLECTROTARSIDAE		
<i>Plectrotarsus tasmanicus</i>	Common, restricted	1
TASIMIIDAE		
<i>Tasimia</i> sp.	Common, restricted	17
HELICOPSYCHIDAE		
<i>Helicopsyche</i> sp.	Common, restricted	3
CONOESUCIDAE		
<i>Costora rotosca</i>	Common, limited	3,11,17
<i>Costora delora</i>	Common, restricted	4,5
<i>Conoesucus fromus</i>	Common, limited	2,4,17
<i>Conoesucus digitiferus</i>	Abundant, widespread	3,4,7,10-12,17
<i>Conoesucus nepotulus</i>	Common, restricted	12,17
CALOCIDAE		
<i>Caenota plicata</i>	Common, widespread	3,4,7,9-11,15
<i>Tamasia variegata</i>	Common, restricted	3
HELICOPHIDAE		
<i>Alloecella grisea</i>	Rare, restricted	10
<i>Alloecella longispina</i>	Common, limited	7,8,12
<i>Alloecella pilosa</i>	Rare, restricted	12
PHILORHEITHRIDAE		
<i>Aphilorheithrus stepheni</i>	Rare, restricted	1, 4
<i>Aphilorheithrus</i> sp.A	Rare, restricted	14
<i>Tasmanthrus angustipennis</i>	Common, limited	3,5,17
<i>T. galbinomaculatus</i>	Abundant, widespread	4,7,10-12,14-16
LEPTOCERIDAE		
<i>Triplectides similis</i>	Rare, restricted	1
<i>Triplectides bilobus</i>	Abundant, widespread	4,7,11,13-17
<i>Symphitoneuria opposita</i>	Common, restricted	1
<i>Triplectidina nigricornis</i>	Common, restricted	6
<i>Notalina parkeri</i>	Rare, restricted	1
<i>Notalina fulva</i>	Common, restricted	1,4
<i>Notalina</i> sp.	Common, limited	1,3,5,17
<i>Condoceris</i> sp.	Common, restricted	2,14
<i>Oecetis scirpicula</i>	Rare, restricted	1

TABLE 5 Distribution of caddisfly species, Pelion Valley (Taxa recorded from only one locality not included)

	Running water sites increasing catchment size →												Lake outlet sites 5 3	Moss face of waterfall 12	Standing waters 1 6
	2	8	13	9	16	10	15	11	14	7	4	17			
<i>Apsilochorema obliquum</i>					X	X		X	X	X			X		
<i>Austrochorema pegidion</i>		X	X		X					X					
<i>Ulmerochorema</i> sp.										X	X	X	X		
<i>Ethochorema nesydrion</i>					X	X	X	X	X	X	X	X			
<i>Taschorema asmanum</i>						X		X	X	X	X		X	X	
<i>Taschorema evansi</i>													X	X	
<i>Taschorema viridarium</i>										X	X				
<i>Moruya</i> sp.					X	X		X	X	X	X			X	
<i>Hydrobiosella cognata</i>		X	X	X	X	X		X	X	X	X			X	

Table 5 continued.

	2	8	13	9	16	10	15	11	14	7	4	17	5	3	12	1	6
<i>Hydrobiosella waddama</i>												X		X			
<i>Asmicridea edwardsi</i>											X			X			
<i>Diplectrona</i> sp.		X	X		X	X	X	X									
<i>Plectrocnemia</i> sp.										X	X						
<i>Nyctiophylax repandus</i>									X	X		X					
<i>Archaeophylax ochreus</i>			X		X	X	X	X	X	X							
<i>Costora rotosca</i>								X				X		X			
<i>Costora delora</i>											X			X			
<i>Conoesucus fromus</i>	X										X	X					
<i>Conoesucus digitiferus</i>						X		X		X	X	X		X		X	
<i>Conoesucus nepotulus</i>												X				X	
<i>Caenota plicata</i>				X		X	X	X		X	X			X			
<i>Alloecella longispina</i>	X								X					X			
<i>Tasmanthrus angustipennis</i>												X		X	X		
<i>Tasmanthrus galbinomaculatus</i>					X	X	X	X	X	X	X					X	
<i>Triplectides bilobus</i>		X		X		X	X	X	X	X	X	X					
<i>Notalina</i> sp.												X		X	X		X
<i>Condocerus</i> sp.	X								X								
Total number of species	4	3	5	3	6	11	12	9	12	16	16	18	9	9	10	7	1

Appendix. Details of sampling locations.

Site 1 D. Cartwright and J. Dean, 16 January 1990, Tas., Lake Ayr, 8114.214695

Site 2 J. Dean and D. Cartwright, 16 January 1990, Tas., Small trickle flowing into Lake Ayr, 8114.214695

Site 3 D. Cartwright and J. Dean, 16 January 1990, Tas., Lake Ayr outlet stream, approx. 100 metres d/s Lake Ayr, 8114.211695

Site 4 J. Dean and D. Cartwright, 16 January 1990, Tas., Douglas Creek, upstream of confluence with Lake Ayr outlet stream, 8114.206691

Site 5 D. Cartwright and J. Dean, 16 January 1990, Tas., Lake Ayr outlet stream, upstream of confluence with Douglas Creek, 8114.205692

Site 6 J. Dean and D. Cartwright, 16 January 1990, Tas., Isolated pools, Pelion Plains, 8114.207692

Site 7 D. Cartwright and J. Dean, 16 January 1990, Tas., Douglas Creek, Pelion Rangers Hut, 8114.208683

Site 8 J. Dean and D. Cartwright, 17 January 1990, Tas., Headwater stream, 100 metres North of Pelion Gap, 8114.217649

Site 9 D. Cartwright and J. Dean, 17 January 1990, Tas., Unnamed creek, approx. 3/4 km, NW of Pelion Gap, 8114.211654

Site 10 J. Dean and D. Cartwright, 17 January 1990, Tas., Unnamed creek, approx. 1 1/2 km, NW of Pelion Gap, 8114.209661

Site 11 D. Cartwright and J. Dean, 17 January 1990, Tas., Sharers Hut Creek, approx., 2 1/2 km NW of Pelion Gap, 8114.208668

Site 12 J. Dean and D. Cartwright, 17 January 1990, Tas., Douglas Creek, approx., 2 1/2 km NW of Pelion Gap, 8114.208669

Site 13 D. Cartwright and J. Dean, 17 January 1990, Tas., Unnamed creek, approx., 2 3/4 km NW of Pelion Gap, 8114.207673

Site 14 J. Dean and D. Cartwright, 18 January 1990, Tas., River Forth, Frog Flats, 8114.172674

Site 15 D. Cartwright and J. Dean, 18 January 1990, Tas., Unnamed creek, approx., 1/4 km E of Frog Flats, 8114.176674

Site 16 J. Dean and D. Cartwright, 18 January 1990, Tas., Unnamed creek, approx., 3/4 km NE of Frog Flats, 8114.179678

Site 17 D. Cartwright and J. Dean, 18 January 1990, Tas., Douglas Creek, High Bridge, near old Pelion Hut, 8114.198690

